

Investigate the Effect of Fatty acids on Rheological Properties and In Vitro Permeability of Escitalopram Oxalate from Hydroxypropyl Cellulose Gel Formulations.

ABSTRACT

The present studies evaluate rheological properties and in-vitro permeability properties of Escitalopram Oxalate (ECO) containing hydroxypropyl cellulose (Klucel HF, HPC) gels prepared with different carbon chain length containing fatty acids. The formulations were prepared by mixing solvent, escitalopram oxalate and kulcel HF (HPC) in homogenizer at 25000 RPM. A controlled stress rheometer was used to study the effect of different number of carbon chain fatty acids on the rheological properties and microstructure of HPC gels. The in-vitro permeability study was performed using human cadaver skin in order to evaluate the enhancing effect of fatty acids. The studies demonstrated that as the carbon chain length increased (C₁₀-C₁₈) the zero-shear viscosity, and yield stress value increased, which suggested that the stability of gel structure was increased with increase in carbon chain of fatty acids. Cohesive Energy was also depended on the carbon chain of fatty acids. There was decreased in cohesive energy as decrease in carbon chain of fatty acids. Temperature loop was created using heating and cooling temperature cycle. Oleic acid (C₁₈) gave the best thermal stability with lowest temperature loop area. Increase in carbon chain length of fatty acids decreased the permeability enhancing effect of Escitalopram Oxalate through human cadaver skin during In-vitro permeability studies. The permeability of ECO through human cadaver skin was found to be in increasing order as capric acid > lauric acid > Oleic acid > No-enhancer. Rheological studies could be useful to investigate the internal structure of HPC gels. Fatty acids alter the rheological properties of HPC gels such as zero shear viscosity, yield stress and cohesive energy. Moreover, In-vitro permeability results demonstrated that HPC gels containing fatty acids could be potential delivery system for transdermal delivery of ECO.

Keywords: Polymer rheology, Microstructure, Fatty acids, Diffusion, Transdermal, Escitalopram oxalate, Viscosity, Cohesive energy

1. INTRODUCTION

Depression is a chronic or recurrent mood disorder that affects both economic and social function of about 121 million people worldwide. The World Health Organization published a research which showed that depression will be second largest disease in the world by the year 2020 for all ages and sexes [1, 2]. In 1980, Tricyclic antidepressant (TCA) and Monoamine Oxidase Inhibitors (MOI) were the first choice of drugs. However, their side effects, toxicity and drug-drug interaction required newer class of agents which affecting central nervous system with fewer side effects [3, 4]. Escitalopram oxalate (ECO) is one of the newer class of anti-depressant agent. It is an S-isomer of citalopram. ECO is well absorbed following oral administration, with a bioavailability of approximately 80%. Peak plasma level of ECO usually 10-30 ng/ml for 10 mg of oral dosage form with steady state plasma concentration of 20-125 nmole/L. The half-life of ECO is between 27-32 hrs. There are severe chances of overdose with current ECO oral dosage form. The possible side effects of overdose include convulsion, coma, dizziness, hypotension, insomnia, nausea, vomiting, sinus tachycardia, somnolence, and ECG changes [5].

29 Furthermore, the saw-tooth pattern of plasma drug concentration following oral administration
30 often causes adverse events at maxima and loss of therapeutic effect at minima, which leads to
31 intolerability in case of many antidepressants [6]. Therefore, studies on the transdermal route for systemic
32 drug delivery of ECO has attracted considerable attention. Furthermore, the skin is an attractive potential
33 route of drug administration because of avoidance of first-pass hepatic metabolism of ECO.

34 Polymers that can form a gel or matrix are key component for transdermal drug delivery system
35 (TDDS) [7, 8]. One of the representative of that polymer system is Hydroxypropyl cellulose (Klucel-HF,
36 HPC), a non-ionic, hydrophilic and pH independent polymer. It is widely used in pharmaceutical industry
37 as a thickening agent or gelling agent to achieve sustained release of the small molecules from different
38 dosage forms. A polymer gel is a solid-in-liquid colloid in which the solid phase forms a network structure
39 that immobilizes the liquid and produces solid or semisolid like properties. Gelation arises either from
40 chemical cross-linking through covalent bond formation or from physical cross-linking through polymer-
41 polymer interactions [9, 10]. Rheological properties of polymer are useful as they can provide some of the
42 fundamental finished product properties such as storage stability, effect of formulation variable on the
43 microstructure, consistency of the product, drug releasing from polymer entrapment and etc. [11].
44 Rheology is a powerful tool to obtain information and characterized microstructure of wide range of
45 materials such as solutions, polymer melts, gels, particulate systems etc. [12]. Rheological measurement
46 mainly performs in two modes: 1) Rotational (shear) mode and 2) Oscillation (Dynamic) mode. During low
47 stress/strain, inter and intra-particle molecules arrangements are only slightly disturbed during
48 measurement which make microstructure in quiescent conditions [13]. This is the primary advantage of
49 oscillation mode over rotational mode, which destroy the internal structure of sample. Moreover,
50 oscillation mode can also useful in illustrating the elastic and viscous behavior of sample [14]. It has been
51 reported that surfactants greatly modify the rheological properties of cellulose ether, poloxamer, poly-co-
52 acrylic acid and carbopol [15-17]. Little attention has been focused to the effect of fatty acids on the
53 rheological properties of transdermal gels. The fatty acids are frequently incorporated to modulate drug
54 release rate by promoting the drug permeability through human cadaver skin [18].

55 The objective of this research was to evaluate the effect of number of carbon chain of fatty acids
56 on the rheological properties of hydroxypropyl cellulose gels. Rheological properties were investigated
57 using viscometry and oscillation mode to evaluate the viscoelastic and internal structure properties of the
58 HPC gels. Furthermore, the effect of fatty acids on the permeability of ECO through human cadaver skin
59 was also evaluated.

60

61 **2. MATERIAL AND METHODS**

62

63 **2.1 Materials:**

64 Escitlopram oxalate (ECO) was purchased from Tacoland Corporation, CA, USA. Klucel HF (HPC) was
65 gifted from Hercules, Wilmington, DE, USA. Human cadaver skin was purchased from NY Firefighter skin
66 bank, NY, USA. Oleic acid, lauric acid, capric acid and sodium azide were purchased from Sigma Aldrich,
67 St. Louis, MO, USA. Polyethylene glycol 400 (PEG-400), propylene glycol (PG), isopropyl alcohol (IPA),
68 dimethyl sulfoxide (DMSO), ethanol 190 proof and glycerin was purchased from Pharmaco-Appear co,
69 Brookfield, CT, USA.

70 **2.2 Methods:**

71 **2.2.1 Gel Preparation:**

72 Appropriate quantities of propylene glycol, polyethylene glycol-400, DMSO, isopropyl alcohol, ethanol,
73 glycerin and water are given in Table 1, were mixed together and required quantities of different fatty
74 acids (8% w/w) was added into the solvent mixture according to Table 1. ECO (5% w/w) was added and
75 mixed until it was dissolved in the solvent mixture. Klucel-HF was dispersed in drug solution, and
76 homogenized at 25000 RPM for 30 min.

77 **Table1: Preparation of ECO containing HPC gel**

Ingredients	Formulation (%W/W)			
	FO-25	FL-25	FC-25	F-25
ECO	5	5	5	5
DMSO	10	10	10	10
Glycerine	6	6	6	6
PEG-400	20	20	20	20
PG	20	20	20	20
IPA	20	20	20	20
Water	10	10	10	10
Oleic Acid	8	-	-	-
Lauric Acid	-	8	-	-
Capric Acid	-	-	8	-
Klucel HF	1	1	1	1
Mixing Speed (X1000) RPM	25	25	25	25

78

79 **2.2.2 Rheological measurements:**

80 An advanced Gemini II rheometer (Malvern Instruments, USA) with a cone-plate configuration (diameter
81 40 mm, Gap 150 μm , and cone angle 4°) was used for the rheological measurements of the gels. The gel
82 sample was gently loaded onto the rheometer peltier plate using a tablespoon. Care was taken to
83 minimize shearing during sample removal and sample loading. Rheological tests were performed in two
84 different modes: 1) Rotational and 2) Oscillation.

85 ***2.2.2.1 Rotational Mode:***

86 **Flow Curve Test:** Viscosity curves were generated in controlled-rate mode, with shear rates ranging from
87 $0.01\text{-}100\text{ S}^{-1}$ at temperatures of 25°C and the apparent viscosity as a function of shear rate was
88 monitored. In each case, the shear rate was increased over a period of 198 s and 50 data acquisition
89 points were recorded. Most pharmaceutical gels showed shear thinning behavior which could be
90 described by Cross model [19]:

$$91 \quad (\eta - \eta_\infty) / (\eta_0 - \eta_\infty) = [1 / \{1 + (K^* \dot{\gamma})^m\}] \quad (1)$$

92 The cross model described the relation between apparent viscosity and shear rate where, η_0 is zero
93 shear viscosity, η_∞ is high shear viscosity, K and m are constant, K has the unit of time and m is
94 dimensionless. The degree of shear thinning can be dictated by value of m ($0 < m < 1$). When m
95 approaches zero, the material called Newtonian and for non-Newtonian shear thinning pharmaceutical

96 gels m have value approaching unity. The value of η_0 and η_∞ can further use to evaluate the structural
97 parameter lambda according to following equation [20]:

$$98 \quad K = 1 - (\eta_\infty / \eta_0)^{1/2} \quad (2)$$

$$99 \quad \lambda = (1 - (\eta_\infty / \eta)^{1/2}) / K \quad (3)$$

100 According to indirect microstructure theories, $\lambda=1$ represent the completely build structure while $\lambda= 0$
101 represents the breakdown structure.

102 **2.2.2.2 Oscillation Mode:**

103 **Amplitude Sweep Test:** Amplitude Sweeps were performed in controlled strain mode over the strain
104 range of 0.01- 10 at a frequency of 1 Hz at 25 °C respectively. During each sweep, 20 data points were
105 collected. A double logarithmic axis was used and the linear Viscoelastic (LVE) range, that is, the
106 deformation ranges over, which the elastic modulus value remains relatively linear and constant, was
107 calculated. During LVE range stress and strain have same value. Amplitude sweep can be used to
108 calculate the cohesive energy. Cohesive energy is the energy required to break all the bonds associated
109 with one of its constituent molecules. It is, therefore measure of the inter-molecular energy for a
110 formulation. Cohesive energy increases the intensity of molecule interaction. Cohesive energy of different
111 gel system could be calculated according to following equation [21]:

$$112 \quad \text{C.E.} = \frac{1}{2} (G' \times A^2) \quad (4)$$

113 Where, C.E. is the cohesive energy and G' is the storage modulus (Pa). During amplitude sweep, with
114 increasing strain, the value of G' and G'' is linear during LVE range and after that it significantly dropped
115 due to structural break down. The strain at this turning point is A.

116 **Temperature Sweep:** The thermal stability of formulations was evaluate using two types of temperature
117 ramps, namely a temperature cycle test and temperature ramp tests. In the *temperature cycle test* a
118 constant deformation 0.1 strain was applied at frequency of 1 Hz. The temperature was increased linearly
119 from 5 °C to 80 °C at a rate of 1 °C/min with 800 data points acquisition. Thereafter temperature was
120 decreased back to 5 °C with the same parameters. Thus, heating and cooling rates were kept constant 1
121 °C/min. These heating and cooling ramp created a temperature loop. During the *temperature ramp test*, a
122 constant deformation of 0.1 strain was applied at frequency of 1 Hz. The temperature was increase
123 linearly at the rate of 1 °C/min from 5 °C to 80 °C.

124 **2.2.3 In-Vitro permeation studies:**

125 All In-vitro permeation studies were carried out using vertical type Franz-diffusion cells with diffusional
126 area of 1.76 cm². These cells have a static receiver solution reservoir with a side arm sampling port
127 design. During the course of an experiment, 0.5 ml of the receptor solution were collected at pre-
128 determined time for analysis and replaced with a same amount of fresh receiving media. The receptor
129 compartment (volume 13 ml) was filled with receiving media (PBS pH=7.4 (0.01% Na Azide)) after
130 degassing to avoid air bubbles. The receptor compartment was maintained at 32 °C by means of a water
131 bath circulator and a jacketed surrounding cell. The solution in receiver compartment was continuously
132 stirred by means of coated magnetic stirrer. Human cadaver skin was soaked in PBS for 1 hr. before
133 mounting between the donor and receiving compartment and was secured by means of a pinch clamp. 1
134 gm of each formulation was placed in donor compartment. Each formulation was run 3 times to get better
135 average of permeation of Escitalopram Oxalate through the human cadaver skin.

136 **2.3 Data Analysis:**

137 The effect of different carbon chain length fatty acids on the rheological properties were evaluated using
138 Microsoft-Excel 2016. Data analysis were performed using Minitab 17 to evaluate area under the loop for
139 temperature cycle data.

140

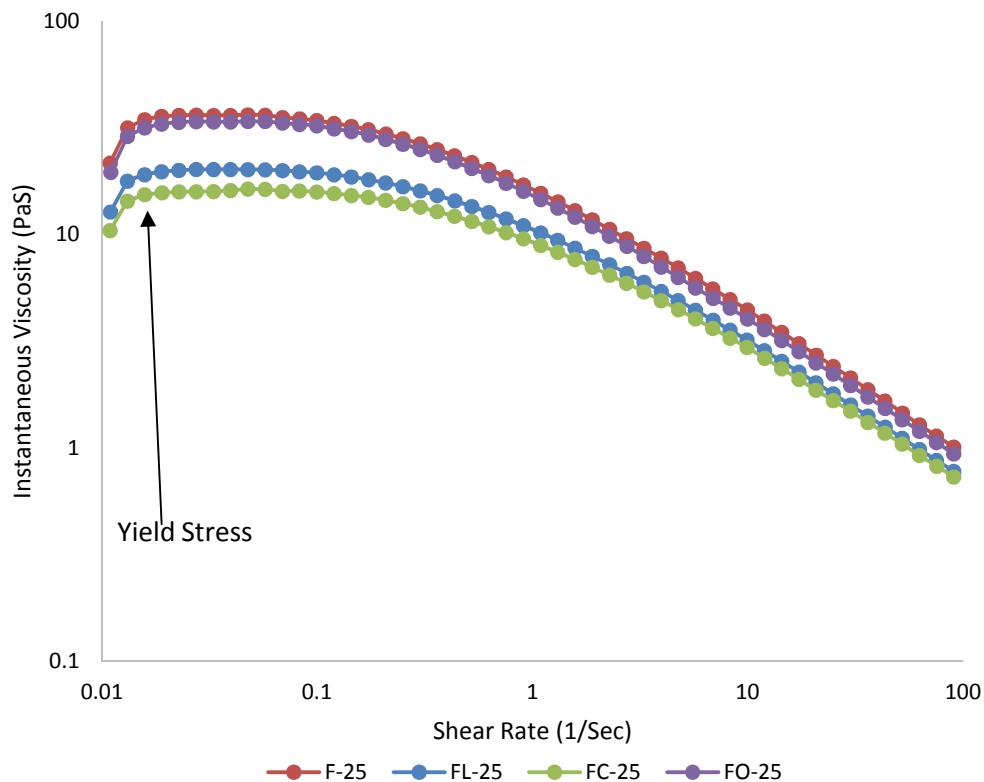
141 3. RESULTS AND DISCUSSION

142

143 3.1 Effect of Fatty acids on Rheological Properties:

144

145 Most of the pharmaceutical gel formulation showed thixotropic behavior that describes a
146 degradation of the polymer structure during the loaded phase; thus, a reduction in viscosity with time
147 occurs when shear stress/shear rate is applied. Therefore, a thixotropic material would have shear-
148 thinning (increase polymer disentanglement) behavior when a gradually increasing shear was applied [22,
149 23]. Figure 1 represents the instantaneous viscosity vs. shear rate profile for formulations. Application of
150 the cross model to instantaneous viscosity vs. shear rate profile indicated that the gels are significantly
151 shear thinning with 'm' approaching unity, and at low shear rates yield stress was also observed.
152



153

154 **Fig. 1. Instantaneous Viscosity vs. Shear Rate profile master curve for FO-25, FL-25, FC-25 and F-**
155 **25 (n=3)**

156 A different carbon chain length fatty acids containing gels exhibited similar shear thinning
157 behavior ($m= 0.90$) but significantly low zero-shear viscosity value. The observed zero-shear viscosity
158 values were proportional to the carbon chain length of fatty acids present in the formulation. The zero-
159 shear viscosity value for C_{18} carbon chain containing fatty acid was almost 108% higher than the
160 formulation containing C_{10} carbon chain fatty acid. The increase in zero-shear viscosity could be due to
161 viscous nature of oleic acid.
162

163 The cross-model's time constant K is related to the shear dependent structural breakdown. A high
 164 value of K implies a relatively large shear dependent contribution to structural breakdown. In another
 165 word, when K is large, breakdown occurs at relatively low shear rate. Observed value of K was also
 166 proportional to the number of carbon chains in fatty acids.

167
 168 Most of the pharmaceutical gels have physically cross-linked structure [23]. In cross-linked
 169 microgel structure individual particles are closely packed with their neighbor was responsible for the yield
 170 stress. The magnitude of the yield stress is a measure of the strength of the closed-pack structure that
 171 must be exceeded for the formulation to flow appreciably [24]. The observed yield stress values were
 172 depended on the fatty acid's carbon chain length. Observed yield stress value for capric acid was 0.73
 173 Pa, which was almost 45% lower than C₁₈ chain containing fatty acid. (Table-2)

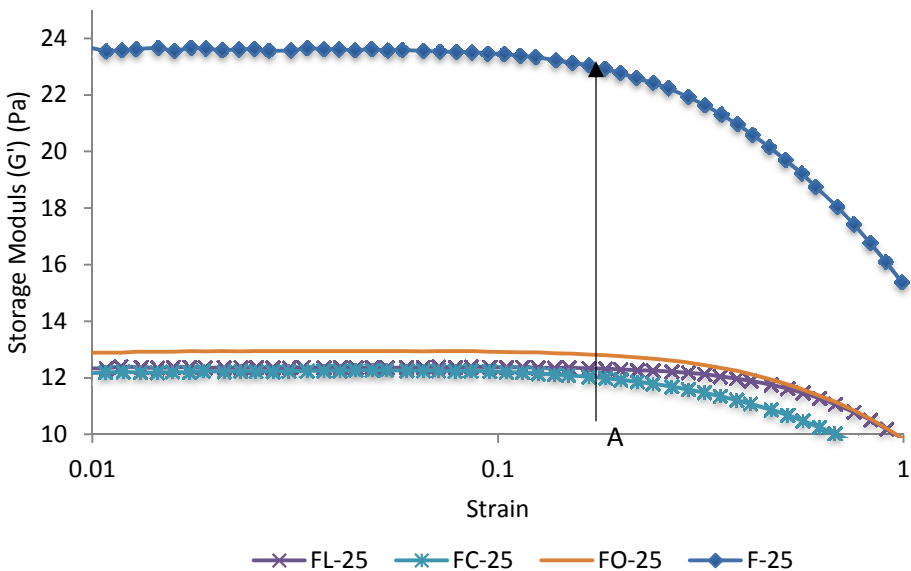
174
 175
 176 **Table 2. Flow Curve Parameters (n=3)**

Formulation	Cross Model Parameters					λ^*	Yield Stress (Pa) \pm S.D.
	η_0 (PaS)	η_∞ (PaS)	M	K (Sec)	R ²		
FO-25	33.15	0.47	0.9035	1.004	0.9822	0.893	1.33 \pm 0.02
FL-25	19.93	0.37	0.8801	0.7962	0.986	0.896	0.85 \pm 0.3
FC-25	15.92	0.35	0.8887	0.6585	0.987	0.896	0.73 \pm 0.1
F-25	35.8	0.46	0.8848	1.06	0.9834	0.899	1.43 \pm 0.07

177 * : Cross model and λ was calculated from the master curve of three replicates

178 Cellulosic derivative polymer has a –OH group in their structure, which forms hydrogen bonds
 179 between polymer chains. These hydrogen bonds partially or fully destroyed by the interaction between –
 180 OH group of solvents. These interactions affect the internal structure of polymer gels [25]. Addition of
 181 fatty acid disrupt the stable structure and this could be due to formation of hydrogen bonding between
 182 fatty acid and polymer chains. This hypothesis could be supported by evaluating cohesive energy for
 183 each formulation, which was related to the carbon chain of fatty acids. Table-3 showed that as increase in
 184 carbon chain of fatty acid, cohesive energy was also increased. This was suggested that energy required
 185 to break H-bond between oleic acid and polymer chain could be more energy consuming than the H-bond
 186 between lauric acid or capric acid and polymer chains.

187
 188 Furthermore, storage modulus (G') is correlating with the deformation energy stored in sample
 189 during shear process and represents as an elastic behavior of sample. According to Figure 2, formulation
 190 F-25 had the highest storage modulus (G') value, which suggested incorporation of fatty acids in
 191 formulation lower the elastic behavior of the HPC gel. However, the LVE region for F-25 was lower, which
 192 suggested that the stability of HPC gel was increased due to the presence of fatty acid in the formulation.



193

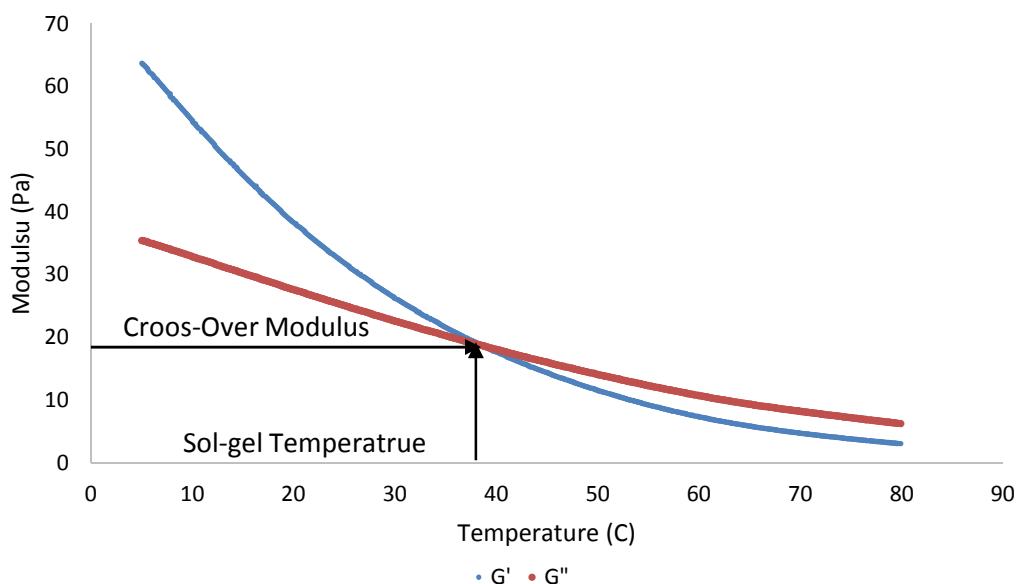
194 **Fig. 2. Representative graph of Storage Modulus (G') vs. Strain Profile for FO-25, FL-25, FC-25**
 195 **and F-25**

196 The stability of any gel depends on the mobility of particles, which are present in the sample. By
 197 increasing mobility, probability of particle-particle interaction increase and stability of the product
 198 decrease [26]. In order to determined temperature stability of gels, it was important to monitor the
 199 rheological properties of sample through temperature cycle. When complex modulus was monitored
 200 throughout the temperature cycle, it created the loop. The area of the loop determined the stability of gels.
 201 For thermally stable gels, the microstructure was not disturbed with temperature and it showed smaller
 202 loop area as compare to thermally unstable gel formulations. Furthermore, if the energy required to break
 203 the bonds is lower, than the formulation will not able to tolerate the higher temperature and the structure
 204 will break down at lower temperature. This assumption was confirmed by running the temperature cycle.
 205 Formulation with oleic acid showed lowest temperature loop area, while formulation F-25 showed highest
 206 temperature loop area.

207

208 In temperature sweep, when G' is equal to G'' , It is called sol-gel transition temperature. During
 209 the whole range of temperature ramp G' and G'' was inversely propositional to temperature. (Figure 3)
 210 The initial decrease of modulus could be related to the increase in fluidity with increasing temperature.
 211 This decrease might also be attributed to the energy dissipation movement of the molecules and
 212 decreased in intermolecular interactions, which in turn decreased the energy needed for the flow, thus
 213 decreased the interference of the hydrodynamic domains [27, 28]. Table-3 provided all thermos-
 214 rheological parameters for formulations

215



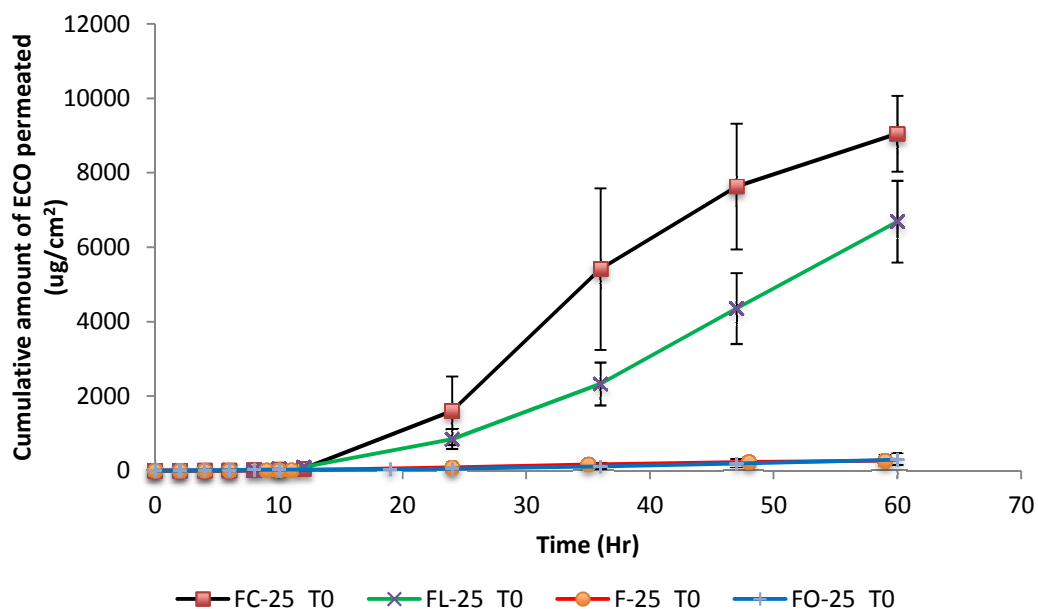
216
217 **Fig. 3. Representative graph of Temperature sweep for Formulation F-25**

218
219 **Table 3. Thermo-Rheological Parameters (n=3)**

Formulation	Cohesive Energy (J/m ³) ± S.D.	Temperature loop area (J°C/m ³) ± S.D.	Cross-Over Temperature (°C) ± S.D.	Cross-Over Modulus (Pa) ± S.D.
FO-25	6.3 ± 0.02	215.91 ± 0.5	32.7 ± 0.5	15.1 ± 0.17
FL-25	5.45 ± 0.05	330.32 ± 1.0	30.1 ± 1.0	13.8 ± 0.11
FC-25	4.76 ± 0.1	337.6 ± 0.7	27.5 ± 0.3	13.6 ± 0.14
F-25	6.65 ± 0.07	389.04 ± 1.2	38.2 ± 1.0	14.8 ± 0.13

220
221
222 **3.2 Effect of fatty acid on permeability of Escitalopram Oxalate:**

223
224 The flux of each formulation was obtained through the slope of cumulative amount of ECO
225 permeated (µg) through human cadaver skin verses Time (Hr) profile (figure 4). The lag time was
226 evaluated by extrapolating the regression line. It showed that different fatty acids affect the flux profile of
227 ECO. Lower carbon chain fatty acids showed higher diffusion of ECO from human cadaver skin as
228 compare to higher carbon chain fatty acids. Enhancement ratio was calculated from equation 5 for each
229 formulation and presented in Table-4



230
231 **Fig.4 Effect of Fatty acids on the permeability of ECO through Human Cadaver Skin (n=3)**

232 **Table 4. Transdermal Parameters (n=3)**

Formulation	Flux ($\mu\text{g}/\text{cm}^2/\text{Hr}$) \pm S.D.	Permeability Co-Efficient X 10^{-4} (cm/Hr)	Enhancement Ratio
FO-25	23.17 \pm 4.66	4.63	4.59
FL-25	184.05 \pm 5.51	36.81	36.45
FC-25	205.58 \pm 5.17	41.12	40.71
F-25	5.05 \pm 2.27	1.01	1.00

233

234
$$E. R. = \frac{\text{Flux of Formulation with enhancer}}{\text{Flux of Formulation without enhancer}} \quad (5)$$

235

236 Several researchers have demonstrated that the fatty acids with carbon chain of C_{10} - C_{12} increase
237 the permeation of hydrophilic drug molecule through human cadaver skin as compare to C_{18} carbon chain
238 fatty acids [29-31]. The permeability coefficient (P) has been calculated using following equation:
239

240
$$P = \frac{J}{C} \quad (6)$$

241

242 Where, J is the flux obtained from the Cumulative amount (Q) versus time ($\mu\text{g}/\text{cm}^2/\text{Hr}$), C is the
243 initial concentration applied in the donor compartment ($\mu\text{g}/\text{ml}$). The decreased in the flux profile of ECO
244 with oleic acid could also be explained by higher yield stress and zero shear viscosity data. Furthermore,
245 the cohesive energy was also found higher as compare to lauric acid and capric acid, which proposed the
246 greater interaction between oleic acid and HPC polymer chains.
247

248 **4. CONCLUSION**

249
250 Rheological studies conducted on HPC gels showed that presence of fatty acids could alter the
251 microstructure of the gel system. Furthermore, HPC gels displayed shear-thinning behavior with observed
252 yield stress value. Moreover, zero shear viscosity of formulation was depended on the carbon chain
253 length of fatty acids. Length of carbon chain in fatty acids altered the microstructure of HPC gels, which
254 was validated through the cohesive energy. Different carbon chain length of fatty acids showed
255 significantly different thermo-rheological properties that could be due to the difference in microstructure of
256 HPC gels. In-vitro permeability results demonstrated that HPC gels containing fatty acids could be
257 potential delivery system for transdermal delivery of ECO.

258
259 **CONSENT**

260
261 It is not applicable

262
263 **ETHICAL APPROVAL**

264
265 It is not applicable

266
267 **REFERENCES**

268
269 1. Sampson, S.M. Treating depression with selective serotonin reuptake inhibitors: a practical
270 approach. in Mayo Clinic Proceedings. 2001. Elsevier.
271 2. Smith, R. and M.J. Bogusz, Forensic science. Vol. 6. 2011: Elsevier.
272 3. Kent, J.M., SNaRIs, NaSSAs, and NaRIs: new agents for the treatment of depression. The Lancet.
273 2000;355(9207): 911-18.
274 4. Pacher, P., et al., Current trends in the development of new antidepressants. Current medicinal
275 chemistry. 2001;8(2):89-100.
276 5. Rao, N., The clinical pharmacokinetics of escitalopram. Clinical pharmacokinetics. 2007;46(4):281-
277 90.
278 6. Kilts, C.D., Potential new drug delivery systems for antidepressants: an overview. The Journal of
279 clinical psychiatry. 2003; 64:31-33.
280 7. Kadam, A.S., M.P. Ratnaparkhi, and S.P. Chaudhary, Transdermal drug delivery: An overview.
281 International Journal of Research and Development in Pharmacy and Life science. 2014;3(4):
282 1042-53.
283 8. Latheeshjhal, L., et al., Transdermal drug delivery systems: an overview. International Journal of
284 PharmTech Research. 2011;3(4):2140-48.
285 9. Tabilo-Munizaga, G. and G.V. Barbosa-Cánovas, Rheology for the food industry. Journal of Food
286 Engineering. 2005;67(1):147-56.
287 10. Waigh T.A., Microrheology of complex fluids. Reports on Progress in Physics. 2005;68(3): 685
288 11. Dobos A.M., Onofrei M.D., Stoica I., Olaru N., Olaru L., Loan S., Rheological Properties &
289 Microstructures of Cellulose acetate Phthalate/Hydroxypropyl Cellulose blends. Polymer
290 Composites. 2012;33(11): 2072-83.
291 12. Menard K.P., Dynamic mechanical analysis: A practical introduction. 2008. CRC press
292 13. Goodwin, J.W. and R.W. Hughes, Rheology for chemists: an introduction. 2008: Royal Society of
293 Chemistry.
294 14. Kim J.Y., Song J.Y., Lee E.J., Park S.K., Rheological properties and microstructures of Carbopol
295 gel network system. Colloid and polymer science. 2003;281(7): 614-23.
296 15. Alvarez-Lorenzo C., Concheiro A., Effect of Surfactant on gel behavior. American Journal of drug
297 delivery. 2003;1(2):77-101.
298 16. Bromberg, L., M. Temchenko, and R.H. Colby, Interactions among hydrophobically modified
299 polyelectrolytes and surfactants of the same charge. Langmuir. 2000;16(6):2609-14.

- 300 17. Caykara T., Kiper S., Demirel G., Thermosensitive poly(N-isopropylacrylamide-co-acrylamide)
301 hydrogels: synthesis, swelling and interaction with ionic surfactant. *European Polymer Journal*.
302 2006;42(2): 348-55.
- 303 18. Pathan, I.B. and C.M. Setty, Chemical penetration enhancers for transdermal drug delivery
304 systems. *Tropical Journal of Pharmaceutical Research*. 2009;8(2):173--79
- 305 19. Chen H., Ding Y., Tan C., Rheological behavior of nanofluids. *New Journal of Physics*. 2007;
306 9(10):367.
- 307 20. Mewis J., Wagner N.J., Thixotropy. *Advance in colloid and Interface Science*. 2009; 147:214-27.
- 308 21. Ji, Y., et al., The relationships between rheological rules and cohesive energy of amphiphilic
309 polymers with different hydrophobic groups. *Journal of Polymer Research*. 2015;22(3):26.
- 310 22. Dong, Z., et al., Rheological properties of polymer micro-gel dispersions. *Petroleum Science*.
311 2009;6(3):294-98.
- 312 23. Osada, Y. and A. Khokhlov, *Polymer gels and networks*. 2001: CRC Press.
- 313 24. Islam, M.T., et al., Rheological characterization of topical carbomer gels neutralized to different pH.
314 *Pharmaceutical research*. 2004;21(7):1192-99.
- 315 25. Kamide, K., *Cellulose and Cellulose Derivatives*. 2005: Elsevier Science.
- 316 26. Liu J., Cao D., Zhang L., Molecular dynamics study on nanoparticle diffusion in polymer melts: a
317 test of the Stokes-Einstein law. *The Journal of Physical Chemistry C*. 2008; 112(17): 6653-61.
- 318 27. Colby R.H., Structure and linear viscoelasticity of flexible polymer solutions: comparison of
319 polyelectrolyte and neutral polymer solutions. *Rheological Acta*. 2010; 49(5):425-42.
- 320 28. Hoare T.R., Kohane D.S., Hydrogels in drug delivery: progress and challenges. *Polymer*. 2008;
321 49(8): 1993-2007.
- 322 29. Williams A.C., Barry B.W., Penetration enhancers. *Advanced drug delivery reviews*. 2012; 64:128-
323 37.
- 324 30. Songkros S., An overview of skin penetration enhancers: penetration enhancing activity, skin
325 irritation potential and mechanism of action. *Songklanakarian Journal of Science and Technology*.
326 2009; 31(3):299-321.
- 327 31. Parisi, N., et al., Topical delivery of hexamidine. *International journal of pharmaceutics*.
328 2016;506(1):332-39.
- 329