

1 **Anti-Inflammatory activity and *In-silico* Approaches on root extract of *Decalepis hamiltonii***  
2 **Wight & Arn**

3 **ABSTRACT**

4 **Aims:** The present study was aimed to evaluate anti-inflammatory and antioxidant  
5 potential of methanol root extract of *Decalepis hamiltonii*. **Results** The methanolic extract  
6 showed *in vitro* anti-inflammatory activity inhibition cells 96.1%µg/ml. **MTT (3-(4,5-**  
7 **dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide)** assay was significant inhibited by the  
8 methanol (96.2) and inhibition addition the **Nitric oxide** production 25µg/ml 95% inhibition. The  
9 methanol extract of *Decalepis hamiltonii* shows a Thus, a very strong approach to In-silico  
10 docking and in vitro anti-inflammatory study for whole with these compounds/extract.  
11 **Conclusion.** From the study it is concluded that the root extract of *Decalepis hamiltonii*  
12 showing a strong approach to *In-silico* docking and anti inflammatory activity.

13 **Keywords:** *Decalepis hamiltonii*; **MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium**  
14 **bromide); Nitric oxide; Ferric reducing antioxidant power**

15 **1. Introduction**

16 Medicinal plants have been used as age long remedies for human diseases because they  
17 contain components of therapeutic value. The plants are naturally large source of antioxidants,  
18 anticancer, antidiabetic agents they might be serve as leads for the development of novel drugs.  
19 The research and analysis on plants employed as pain-relievers and anti-inflammatory agents in  
20 traditional ethno medicine is one of the productive and logical strategies in the search for new  
21 drugs [1-2]. Free radicals, which have one or more un-paired electrons (superoxide, hydroxyl,  
22 peroxy) are produced in normal or pathological cell metabolism and the compounds that can  
23 scavenge free radicals have great potential in ameliorating the diseases and pathological cells [3-

24 6]. The increase in the prevalence of multiple drug resistance has shown down the development  
25 of new synthetic anti-inflammatory drugs and the new drug is necessary to search for new anti-  
26 inflammatory from alterative sources. Phytochemicals from medicinal plants showing anti-  
27 inflammatory activities have the potential of filling this need because of the structures of  
28 phytochemical different from those of the more studied and they're those of the more action may  
29 too very likely differ [7]. In this growing interest, many of the phytochemical bioactive  
30 compounds from medicinal plants have shown many pharmacological activities [8-11]  
31 Inflammation are pathological condition in which highly reactive species are produced. Nitric  
32 oxide (NO) is a small diffusible molecule responsible for vasodilatation, neurotransmission and  
33 inflammation. This molecule is produced by the organism at a basal concentration. Nevertheless,  
34 under stimulation by pathogens, NO is generated in higher amounts by the inducible nitric oxide  
35 synthase (iNOS) in activated macrophages. The over production of NO is involved in the  
36 pathogenesis of septic shock, tissue damage, multiple organ dysfunctions and carcinogenesis  
37 processes [12]. Natural products have contributed significantly towards the development of  
38 modern medicine. Recently traditional medicine worldwide has been re-evaluated by extensive  
39 research on different plant species and their active therapeutic principles. The rich wealth of  
40 plant kingdom can represent a novel source of newer compounds with significant anti-  
41 inflammatory activities. The major merits of herbal medicine seem to be their perceived efficacy,  
42 low incidence of serious adverse effects, and low cost. This *Decalepis hamiltonii* (Wight& Arn),  
43 popularly known as swallow root in English, belongs to the family *Asclepiadaceae*. It is a  
44 climber with stout, smooth branches shrub and a native of the forests of Deccan peninsula and  
45 Western Ghats of India. The leaves are curvaceous, orbicular or elliptical, with rounded tip. Its  
46 tubers are consumed as the pickles and the juice for its alleged health promoting properties The

47 Root has a sweet sarsaparilla-like taste; contains 9.2% fleshy matter and 8% woody core [13]. In  
48 this context the present research work is on antioxidant and anti-inflammatory activity of  
49 *Decalepis hamiltonii* (Wight & Arn).

50

## 51 **2. Material and Methods**

### 52 **2.1 Collection of material**

53 The Root of *Decalepis hamiltonii* has been collected from kolli hills, Namakkal district of  
54 Tamilnadu, India. The taxonomic identification of plant was done with comparing the Flora of  
55 Presidency of Madras, by Gamble J.S., 1921.

### 56 **2.2 Preparation of solvent extraction**

57 50gm of *Decalepis hamiltonii* root was packed in Soxhlet apparatus for extraction and  
58 500 ml of methanol was used as solvent. Soxhlet was kept running for 72 hours, until the solvent  
59 color appears in the collection tube. Methanol was removed by evaporation using rotary vapour  
60 at not more than 40°C. The residue was then placed in an oven at 40°C for about 48hours to  
61 remove the moisture. The resulting dried mass was then powdered and used for further studies.

### 62 **2.3 Ferric Reducing Antioxidant Power**

63 The Ferric reducing antioxidant power of different parts of the various crude extracts of  
64 *Decalepis hamiltonii* was evaluated [14]. The samples were mixed with 2.5 ml of 0.2 M Phosphate  
65 buffer (pH 6.6) and 2.5 ml of 1 per cent potassium ferric cyanide. After the mixture was incubated at 50  
66 °C for 20 min, 2.5 ml of 10 per cent Trichloroacetic acid, 2.5 ml distilled water and 0.5 ml of 0.1 per  
67 cent ferric chloride was added and then the absorbance was measured at 700 nm against a blank. The  
68 blank consist of all the reagents without the test sample. The reducing power of Gallic acid was also

69 determined for a comparison. High absorbance of the reaction mixture indicates strong ferric reducing  
70 antioxidant power.

#### 71 **2.4 MTT assay**

72 The **Peripheral blood mononuclear layer** cell culture was trypsinized and the cell count  
73 was adjusted to  $1.0 \times 10^5$  cells/ml using medium containing 10% new born calf serum. To each  
74 well of the 96 well micro titreplate, 0.1 mL of the diluted cell suspension (approximately 10,000  
75 cells) was added. After 24 hours, when a partial monolayer was formed, the supernatant was  
76 flicked off, washed the monolayer once and 100  $\mu$ L of different drug concentrations was added  
77 to the cells in microtitre plates. The plates were then incubated at 37°C for 3 days in 5% CO<sub>2</sub>  
78 atmosphere and microscopic examination was carried out and observations recorded every 24  
79 hours. After 72 hours, the drug solutions in the wells were discarded and 50  $\mu$ L of MTT  
80 (2mg/mL) prepared in Hank's balanced salt solution (HBSS) without phenol red was added to  
81 each well. The plates were gently shaken and incubated for 3 hours at 37°C in 5% CO<sub>2</sub>  
82 atmosphere. The supernatant was removed and 50  $\mu$ L of propanol was added and the plates were  
83 gently shaken to solubilize the formed formazan[15]. The absorbance was measured using a  
84 microplate reader at a wavelength of 540 nm.

#### 85 **2.5 Nitric Oxide Production**

86 The Nitric oxide (NO) production in cell culture were determined by measuring the  
87 nitrite (NO<sub>2</sub>) formation by using Griess assay [16]. Briefly, 100 $\mu$ L of griess reagent (Sigma-  
88 Aldrich, USA) containing 1% sulfanilamide and 0.1% naphthylethyene diamine dihydrochloride  
89 in 2.5% H<sub>3</sub>PO<sub>4</sub> was added to 100  $\mu$ L of cell culture supernatant and incubate for 15 minutes at  
90 room temperature in the dark. The color density was measured at 540 nm using a microplate

91 reader (Molecular Devices, USA). A standard curve was generated in the same method using  
92 NaNO<sub>2</sub>.

93 The compounds namely 2-Hydroxy-4-methoxybenzaldehyde was identified by GC-MS  
94 analysis were screened against the anti diabetic, **anti - inflammatory** and cancer protein. The  
95 compound details were retrieved from the Pubchem database and the chemical structures were  
96 generated from SMILES notation (Simplified Molecular Input Line Entry Specification) by  
97 using the Chemskech Software (www.acdlabs.com).

## 98 **2.7 Protein Data Bank**

99 The Protein Data Bank (PDB) archive is the single worldwide repository of information  
100 about the 3D structures of large biological molecules, including proteins and nucleic acids. The  
101 Protein Data Bank (PDB) is a repository for the three-dimensional structural data of large  
102 biological molecules, such as proteins and nucleic acids. The data, typically obtained by X-ray  
103 crystallography or NMR spectroscopy and submitted by biologists and biochemists from around  
104 the world, are freely accessible on the Internet via the websites of its member organizations  
105 (PDBe, PDBj, and RCSB). The PDB is overseen by an organization called the Worldwide  
106 Protein Data Bank, WWPDB.

## 107 **2.8 Preparation of protein structure**

108 The structural information of the macromolecules determined by x-ray crystallographic  
109 and NMR methods are available in the PDB. The 3D structure protein Receptor (PDB I.D: 4IFI)  
110 was downloaded from the Protein Data Bank (PDB) (<http://www.rcsb.org/pdb/>) the water  
111 molecules were removed from protein file 4IFI before docking. Energy minimization by  
112 applying for CHARMM (Chemistry at Harvard Macromolecular Mechanics) force fields, It's a

113 program for macromolecular dynamics; it can be used for energy minimization, normal modes  
114 and crystal optimizations and also incorporates free energy methods for chemical and  
115 conformational free energy calculations.

## 116 **2.9 PUBCHEM**

117 The Pubchem bioassay data (<http://pubchem.ncbi.nlm.nih.gov>) is a public repository for  
118 biological Activities of small molecules and small interfering RNAs (siRNAs) hosted by the US  
119 National Institutes of Health (NIH). It archives experimental descriptions of assays and  
120 biological test results and makes the information freely accessible to the public. A Pubchem  
121 Bioassay data entry includes an assay description, a summary and detailed test results. Each  
122 assay record is linked to the molecular target, whenever possible, and is cross-referenced to other  
123 National Center for Biotechnology Information (NCBI) database records. ‘Related Bioassays’  
124 are identified by examining the assay target relationship and activity profile of commonly tested  
125 compounds. A key goal of PubChem Bioassay is to make the biological activity information  
126 easily accessible through the NCBI information retrieval system- Entrez, and various web-based  
127 PubChem services. An integrated suite of data analysis tools is available to optimize the utility of  
128 the chemical structure and biological activity information within PubChem, enabling researchers  
129 to aggregate, compare and analyze biological test results contributed by multiple organizations.  
130 Describe the PubChem Bioassay database, including data model, bioassay deposition and  
131 utilities that PubChem provides for searching, downloading and analyzing the biological activity  
132 information.

## 133 **2.10 Preparation of Ligand structures**

134 The identified Chemical compound namely 2-Hydroxy-4-methoxybenzaldehyde was  
135 derived from *Decalepis hamiltonii* Wight & Arn and this compound structure was retrieved  
136 from Pubchem online server both of these compounds were under investigation of ChemsSketch  
137 (Chemically intelligent drawing interface free ware developed by Advance Chemistry  
138 Development, Inc., (<http://www.acdlabs.com>) was used to construct the structure of the ligands.  
139 The ligand molecules were generated and the three dimensional optimizations were done and  
140 then saved MOL file (a file format for holding information about the atoms, bonds, connectivity  
141 and coordinates of a molecule).

## 142 **2.11 Drug likeliness prediction**

143 Ligand property was predicted by using “Lipinski drug Filters” ([http://www.scfbio-](http://www.scfbio-iitd.res.in/utility/LipinskiFilters.jsp)  
144 [iitd.res.in/utility/LipinskiFilters.jsp](http://www.scfbio-iitd.res.in/utility/LipinskiFilters.jsp)). Lipinski rule of five helps in distinguishing drug-like and non-  
145 drug-like properties and predicts high probability of success or failure due to drug likeliness for  
146 molecules. The Lipinski filter helps in early preclinical assessment and thereby avoiding costly  
147 late stage preclinical and clinical failures.

## 148 **2.12 Docking analysis:**

149 The docking analysis is performed by Argus lab 4.0.1 for the antidiabetic and cancer  
150 protein interacts with **Gas-Chromatography** of *Decalepis hamiltonii* 2-Hydroxy-4-  
151 methoxybenzaldehyde compound. The compound or ligand selected for based on Lipinski's rule of  
152 five. Fitting points are added to hydrogen bonding groups on the protein. The interaction between  
153 the binding pockets of target protein, antidiabetic protein and 3v6r inflammation protein and  
154 investigation compound to find out the accurate binding model for the active site of protein. The  
155 mechanism of ligand placement is based on binding site position. The protein ligand docking

156 energy values performance of this compound was based on the Scoring functions which is  
157 implemented in docking program to make various assumptions and implications to fit best  
158 complexes, which includes terms of hydrogen bonds employed by Argus lab 4.0.1 to rank the  
159 docked bases and to assess the binding site and the number of rotatable bonds present.

### 160 **2.13 Ligand binding sites prediction**

161 After docking the docked structure was saved as “.Pdb” file and further explored to  
162 predict the binding sites using “ligand explorer” software. The predicted binding sites, based on  
163 the binding energy, and amino acids make up the binding cavity. Here ligand binding site  
164 represents the site where the ligands most efficiently bind with the protein, among all the active  
165 site.

### 166 **2.13 Discovery Studio Visualizer**

167 The docking results were visualized using Accelrys Discovery Studio 4.1 Visualizer. The  
168 discovery studio visualizer is also a free viewer that is designed to offer an interactive  
169 environment for viewing and editing molecular structures, sequences, X-ray reflection data,  
170 script and other data. DS Visualizer is handier for analyzing the docking results.

## 171 **3.0 Results and Discussion**

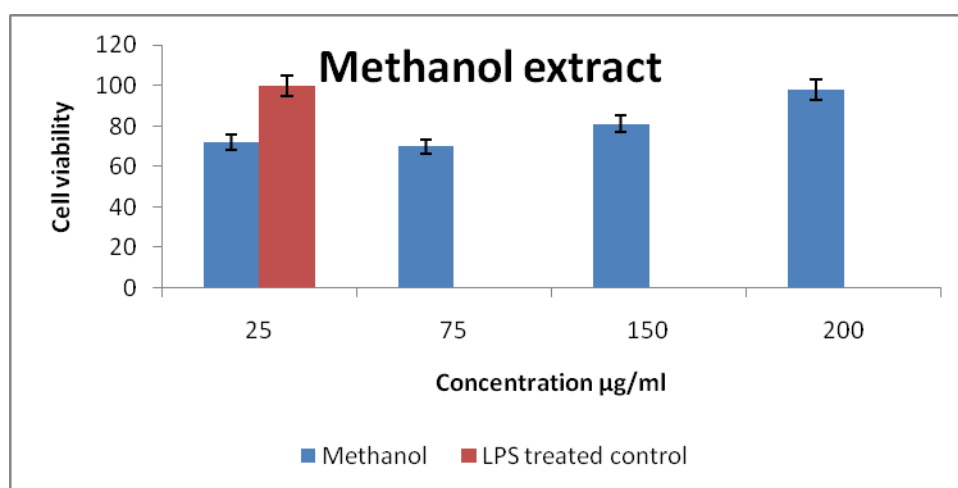
### 172 **3.1 In-vitro Cytotoxicity activity**

#### 173 **Cytotoxicity effect of the root extract of *Decalepis hamiltonii* in Peripheral Blood** 174 **Mononuclear cells (PBMC)**

175 MTT (3-(4,5 dimethylthiazole- 2yl) - 2, 5-Diphenyl Tetrazolium Bromide) assay is  
176 widely used in the *in vitro* evaluation of the biosafety of plant extracts. Determination of



177 cytotoxic effect of plant extracts is needed before studying the mechanism of action in various  
178 cells / cell lines. The cytotoxicity activity of methanolic root extract of *Decalepis hamiltonii* on  
179 cell viability was determined using cell line culture at various concentrations is shown in Fig 1.  
180 The values of cell viability were found to be in the range of 68% to 98% at various concentrations  
181 tested. The highest concentration of 200µg/ml showed moderate Cytotoxicity. Cell viability was  
182 evaluated by MTT assay, hence the concentrations of sample (cell viability  $\geq 90$ ) were selected for  
183 subsequent iNOS inhibition experiment.



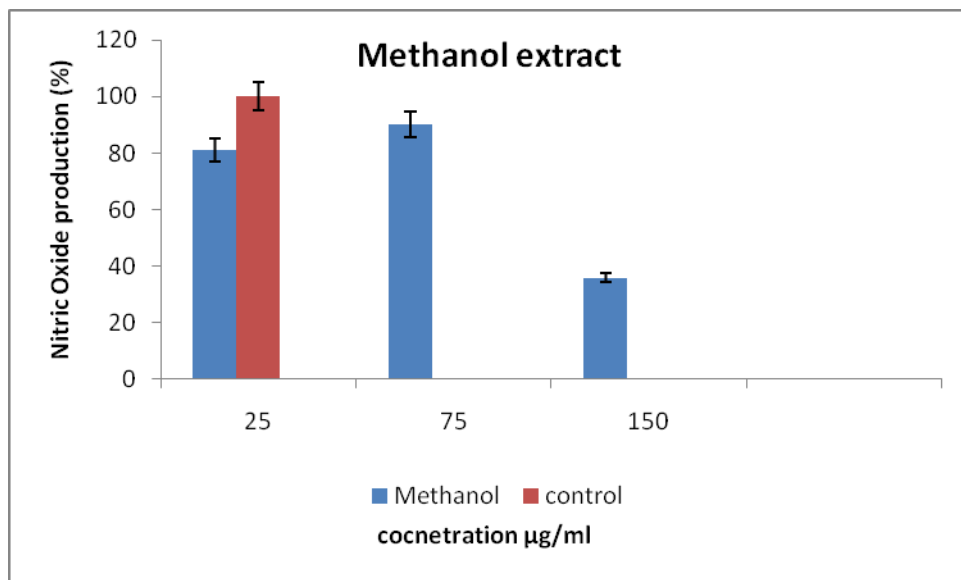
184

185 Results are expressed as mean  $\pm$  Standard deviation

186 **Figure. 1 Cytotoxicity effect of the root extract of *Decalepis hamiltonii***

### 187 **3.2 Inhibition of Lipopolysaccharide – induced iNOS production by root extracts**

188 To investigate the inhibitory effects of the methanolic extracts from the LPS–induced  
189 iNOS production in peripheral blood mononuclear cell (PBMC) the accumulation of iNOS in  
190 the culture medium is measured. As shown in Figure 2, root extracts significantly suppressed  
191 LPS-induced iNOS product in a dose-dependent manner. The root extract at 75µg/ml  
192 significantly inhibited the production of iNOS up to  $90.2 \pm 0.86$ .



193

194

Results are expressed as means  $\pm$  standard deviation

195

**Fig. 2 Nitric oxide effect of the root extract of *Decalepis hamiltonii***

196

### 3.3 Ferric reducing antioxidant power

197

Ferric reducing antioxidant power of Petroleum ether, Chloroform, Ethyl acetate and

198 Methanol extracts of root plant parts of *Decalepis hamiltonii* is shown in Fig. 3. IC<sub>50</sub> values were

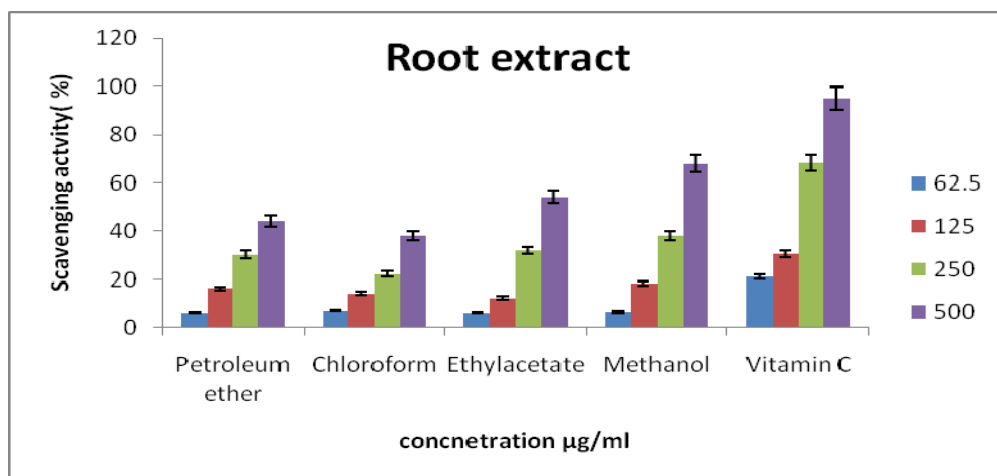
199 calculated as the amount of antioxidant present in the sample necessary to decrease the initial

200 ferric reducing antioxidant concentration by 50%. Petroleum ether, Chloroform, Ethyl acetate

201 and Methanol root extracts of *Decalepis hamiltonii* exhibited the highest activity. The IC<sub>50</sub>

202 values of Methanol extracts of *Decalepis* root and Gallic acid values were 392.7, 442.5, 311.4

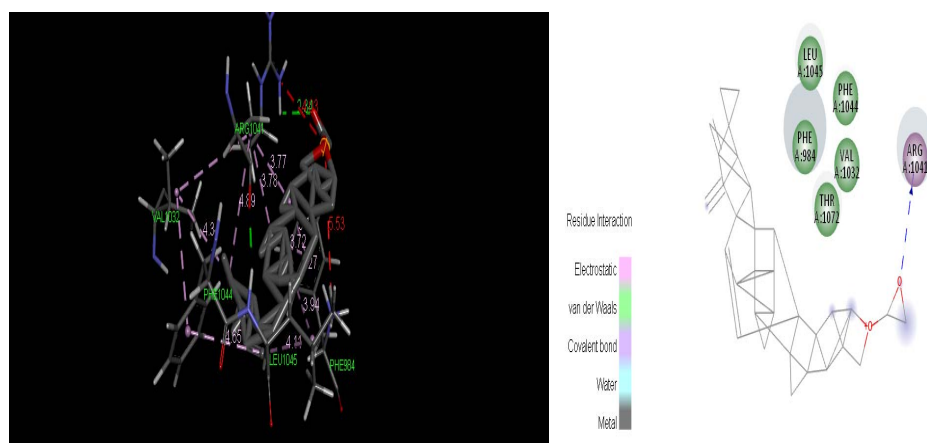
203 and 171.02 µg/ml respectively.



204

205 **Fig. 3 In silico analysis of anti- inflammatory activity of 2-hydroxy-4-methoxybenzaldehyde**  
 206 **from root extract of *Decalepis hamiltonii***

207 The molecular interaction between the ligand and the protein (receptor) was investigated  
 208 using the Ligand fit protocol. The 2-Hydroxy- 4 -Methoxy Benzaldehyde was docked with active  
 209 site inflammation receptor protein 3V6R-docking energy-17.45 kcal/mol. 3D structure and 2D  
 210 structure interaction to interact with Targeted protein; amino acid region and distance. The  
 211 docking studies further confirmed the anti-diabetic property of the bioactive compounds via GC-  
 212 MS analysis and suggested that first time demonstrated a strong 3v6r (Inflammation) inhibitory  
 213 property of this extract, maybe due to the identified bioactive compounds (Figure.4).



214

215 **Figure.4 3D and 2D structures for amino acid regions for 2-Hydroxy-4- Methoxy**  
216 **Benzaldehyde interact with Targeted protein**

217 In the present study methanolic root extract of *Decalepis hamiltonii* was found to have maximum  
218 inhibition concentration (99.5% at 200 µg/ml. Similar results were observed [17], who had  
219 shown moderate cytotoxicity of *Polygonum chinensis* extracts (400- 125 µg/ml). [18] also  
220 compared root, stem and leaf extracts of *Jatropha curcas* in anti-inflammatory activity study  
221 towards carrageen an induced *rat paw edema*. Groups of albino rats were injected by  
222 inflammation inducer (Carrageenan) and the levels of inflammation mediators (histamine,  
223 serotonin and kinin) were monitored. Their results showed that root extract inhibited histamine,  
224 serotonin and kinin released up to 70% - 80% compared to leaf (50% - 55%) and stem (40% -  
225 45%). However, these studies did not indicate the nature of compounds involved in the anti-  
226 inflammatory activity.

227 Ferric reducing properties are generally depend on the presence of reductions which have  
228 been shown to exert antioxidant activity by breaking the free radical chain by donating a  
229 hydrogen atom [19].The present study of the ferric radical activity of the all the extracts of the  
230 *Decalepis hamiltonii* leaves, stem and root were analyzed. The root extract was found to increase  
231 in a concentration depended manner. Among all the four extracts, methanol root extract was  
232 recorded the highest scavenging activity of 78.0% at a concentration of 500µg/ml. The results  
233 suggest that methanol extracts have more reducing power than ethyl acetate, petroleum ether and  
234 chloroform but not as efficient as standard gallic acid. Similar type of work has also been carried  
235 out in *Azima tetracantha* [20]. It is believed that antioxidant activity and reducing power are  
236 related [21]. Reuctones donate a hydrogen atom and inhibit LPO by donating hydrogen. Atom  
237 and there by terminating the free radical chain reaction [22].

238 **3.5 Conclusion**

239 Thus, a very strong approach to *In-silico* docking and in vitro anti-inflammatory study for  
240 whole with these compounds/extracts. Further studies regarding isolation and characterization of  
241 bioactive principles responsible for this action may be further carried out in future.

242 **Reference**

- 243 1. .Elisabetsky E, Ahmador TA, Albuquerque RR, Nunes, DS, Carvalho ACT. Analgesic activity  
244 of Pyschotriacolorata(Wild. ex-R. and S.) Muell. Arg. alkaloids. J.Ethnopharmacol. 1995; 48, 77-  
245 83.
- 246 2. Chen IN, Chang CC, Wang CY, Shyu YT, Chang TL., Antioxidant and antimicrobial  
247 activity of Zingiberaceae plants in Taiwan. Plant Foods Human Nutrition, 2008.,63:15-  
248 20.
- 249 3. Dirsch, V.M., Stuppner, H., Vollmar, A.M. The Griess assay: suitable for a bio-guided  
250 fractionation of anti-inflammatory plant extracts? Planta Medica 1998.,64, 423–426.  
251 discovery. Environment Health Perspective.2001; 109:69-75.
- 252 4. Duh, P.D., Y.Y Tu and G.C. Yen. Antioxidant activity of water extract of harnng jyur  
253 (Chrysanthemum Morifolim Ramat). Leb. Wissn. Technol.,1999 32:269-277
- 254 5. Fabricant DS, Fansworth NR., The value of plants used in traditional medicine for drug  
255 6. Gulcin I, Mshvildadze V, Gepdiremen A, Elias R., Antioxidant activity of saponins  
256 isolated from ivy: alpha-hederin, hederasaponin-C, hederacolchiside-E and  
257 hederacolchiside-F. Planta Medica. 2004;70(6):561-563.
- 258 7. Halliwell B, How to characterize an antioxidant: an update. Biochemical Society  
259 Symposia.1995; 61:85-91
- 260 8. Huang, D., B. Ou and R.L. Prior. The chemistry behind antioxidant capacity assays. J.  
261 Agric. Food Chem.,2005., 53,1841.
- 262 9. Kim PKM, Zamora R, Petrosko P, Billiar TR (2001) Int Immunopharmacol 1:1421-1441.
- 263 10. Moncada, S, Plamer, RMJ, Higgs EA. Nitric oxide: Physiology, pathophysiology and  
264 pharmacology. *Pharmacol. Rev.* **1996**, 43, 109–142
- 265 11. Nagarajan,S, Jagan Mohan Rao L, and Gurudutt, K.N. Chemical composition of the  
266 volatiles of *Decalepsis hamiltonii*.(Wight and Arn). Flavour and Fragrances Journal,  
267 2001;16(1)27-29.

- 268 12. Nayak. B.S and K.N. Patel. Anti-Inflammatory screening of *Jatropha curcas* root, stem  
269 and leaf in albino rats. Rom J Biol –Plant Biol., 2010.,55:9–13.
- 270 13. Pesewu GA, Cutler RR, Humber DP Antibacterial activity of plants in traditional  
271 medicine of Ghana, with particular reference to MRSA. J. Ethnopharm.2008; 116:102-  
272 111.
- 273 14. PinDer Duh, X. Antioxidant Activity of Burdock (*Arctium lappa* Linné): Its Scavenging  
274 Effect on Free-Radical and Active Oxygen J. Am.oil. chem.soc.,1998.,75:4
- 275 15. Prachayasittikul S, Buraparungsang P, Worachartcheewan A, Isarankura-Na- Ayudhya  
276 C, Ruchirawat S, Prachayasittikul V., Antimicrobial and antioxidant activity of  
277 bioreactive constituents from *Hydnophytumformicarum*Jack. Molecules. 2008;13: 904-  
278 921.
- 279 16. Squadriato GL, Peyor WA., Oxidative chemistry of nitric oxide: the role of superoxide,  
280 peroxy nitric and carbon dioxide. Free Radical Biology and Medicine.1998; 25:392-403,
- 281 17. Srividya, A.R., A. Shalom, R. Chandrasekhar, P. Vijayan and V.J. Vishnuvarthan.  
282 Cytotoxic, antioxidant and antimicrobial activity of *Polygonum chinensis* Linn in vero  
283 cell-lines and L6 cell-lines, Int. J. Pharmaceu Sci and Nanotech.,2012., 4: 1569-1574.
- 284 18. Turker AU, Usta C., Biological screening of some Turkish medicinal plants of  
285 antimicrobial and toxicity studies. Natural Products. 2008; 22:136-146.
- 286 19. Vinoth, B., R. Manivasagaperumal and P. Prakash. Free radical scavenging potential of  
287 different extracts from *Azima tetraacantha* Lam. Int. J. Res. Ayurveda Pharm., 2015.,6(1):  
288 131-137
- 289 20. Vongtau, HO. Abbaha. J. Mosugub, O, Chindo BA.Ngazal, IE.Salawu AO, Kwanashie, HO.  
290 Gamaniel, KS, 2004. Antinociceptive profile of the methanolic extract of *Neorautanenia mitis*  
291 root in rats and mice. J. Ethanopharmacol. 92, 317-324.
- 292 **21. Rangika, BS, Dayananda D. Peiris LDC (2015). Hypoglycemic and hypolipidemic activities of**  
293 **aqueous extract of flowers from *Nyctanthes arbor-tristis* L. in mice. BMC complementary and**  
294 **Alternative Medicine. 15 (289); Article ID: 10.1186/s12906-015-0807-0.**
- 295 **22. Peiris LD C, Jayatilleka H, Dhanushka T. (2015). Evaluation of aqueous leaf extract of**  
296 ***Cardiospermum halicacabum* (L.) on fertility of male rats. Biomed Central International Research**  
297 **2015; Article ID: ID 175726,**