

Original Research Article

**GLOBAL WARMING POTENTIALS OF MUNICIPAL SOLID
WASTE DUMPSITE IN CALABAR METROPOLIS, CROSS
RIVER STATE****ABSTRACT**

Total anthropogenic greenhouse gases have continued to increase with municipal solid waste dumpsites contributing immensely to the high concentrations of greenhouse gases with high warming potentials. This study is aimed at determining the global warming potentials of municipal solid waste dumpsite in Calabar metropolis. Soil, leachate and solid wastes samples were collected from the dumpsite and subjected to standard microbiological analysis. The samples were examined for temperature, p^H , mean heterotrophic bacterial/ fungal counts and greenhouse gases emissions from the dumpsite using Combustible Gas Leak Detector. The mean temperature values for both soil and leachate samples ranged from 82⁰F-83⁰F while the mean p^H values ranged from 6.57-7.0. Carbondioxide, Methane and Nitrous oxide in the range The mean total viable aerobic heterotrophic bacterial count in both leachate and soil samples ranged from $1.7 \times 10^3 - 8.0 \times 10^3$ cfu/ml and $1.2 \times 10^4 - 8.0 \times 10^4$ cfu/g, while the mean total viable fungal counts for both leachate and soil samples ranged from $1.0 \times 10^5 - 5.0 \times 10^5$ cfu/ml and $2.1 \times 10^3 - 6.0 \times 10^3$ cfu/g. Bacterial isolate from the samples obtained from the dumpsite include: *Escherichia coli*, *Bacillus sp*, *Pseudomonas aeruginosa*, *Enterobacter sp*, *Klebsiella sp*, *Proteus sp*, *Salmonella sp*, *Staphylococcus aureus*, *Micrococcus luteus*, and *Methanococcus sp*. The fungal isolated include: *Candida tropicalis*, *Aspergillus sp*, *Penicillium spp*, *Candida parapsilosis*, *Candida albicans*, and *Saccharomyces sp*. This study reveals the health, environmental, and climate hazard that could result from indiscriminate dumping of untreated wastes.

Key words: Global warming potential, solid waste, Lemna dumpsite, greenhouse gases

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INTRODUCTION

33 The increasing awareness of global warming all over the world,
34 and most recently its association with solid waste biodegradation is a
35 cause for concern as temperature ranges around such waste dumpsites
36 are of increase than normal. Warming of the climate system is
37 unequivocal and since the 1950s with observed changes (IPCC, 2014).
38 Most researches on the causes of this high temperature have reported
39 greenhouse gases (GHG) emitted from microbial activities of solid
40 waste dumpsite (Bassey, 2012). Lemna solid waste dumpsite formerly
41 known as Ikot-Effanga Mkpa dumpsite is an open dump that
42 encourages the breeding of rodents and proliferation of most
43 pathogenic bacterial and fungal species with no report of the dangers
44 associated with these emissions (Bassey *et al.*, 2015).

45 Waste is any substance, solution, mixture for which no direct
46 use is envisaged but which is transported for reprocessing, elimination
47 by incineration or other methods of disposal (Yakowitz, 1988). With
48 urban industrialization, social development and population increases,
49 solid waste production are growing rapidly making pollution a serious

50 problem (Khupe, 1996; Yaliang, 1996).Solid waste disposal poses
51 threat to humans, animals and the environment at large. Like chemical
52 hazards, etiologic agents might be dispersed in the environment
53 through water and wind.

54 Waste management in developing countries is usually equated
55 with land disposal or discharge water (Cilinskis and Zaloksnis, 1996).
56 This method of waste management is unscientific and causes nuisance
57 to the public and most of all results in global warming as a result of
58 GHG emitted into the atmosphere. This gases alter the climate and
59 also deplete the ozone layer when waste is dumped on land, soil
60 micro-organism including fungi and bacteria, readily colonize the
61 waste carrying out the degradation and transformation of degradation
62 (organic) materials in the waste (Stainer *et al.*, 1989). Micro-
63 organisms in the waste dump use the waste constituents as nutrients,
64 thus detoxifying the materials as their digestive processes breakdown
65 complex organic molecules into simple less toxic molecules (Pavoni
66 *et al.*, 1975).

67 Calabar city does not have a sanitary landfill. Improper disposal
68 of untreated municipal solid waste is not only harmful to human's
69 health but also constitute a threat to the ecological environment
70 (Yaliang, 1996). Temperatures around waste dumps seemed to be
71 higher than normal, hence, the need to study keenly the global
72 warming potentials associated with waste biodegradation in Lemna
73 waste dumpsite.

74 **MATERIALS AND METHODS**

75 **The Study Area and Sampling Site**

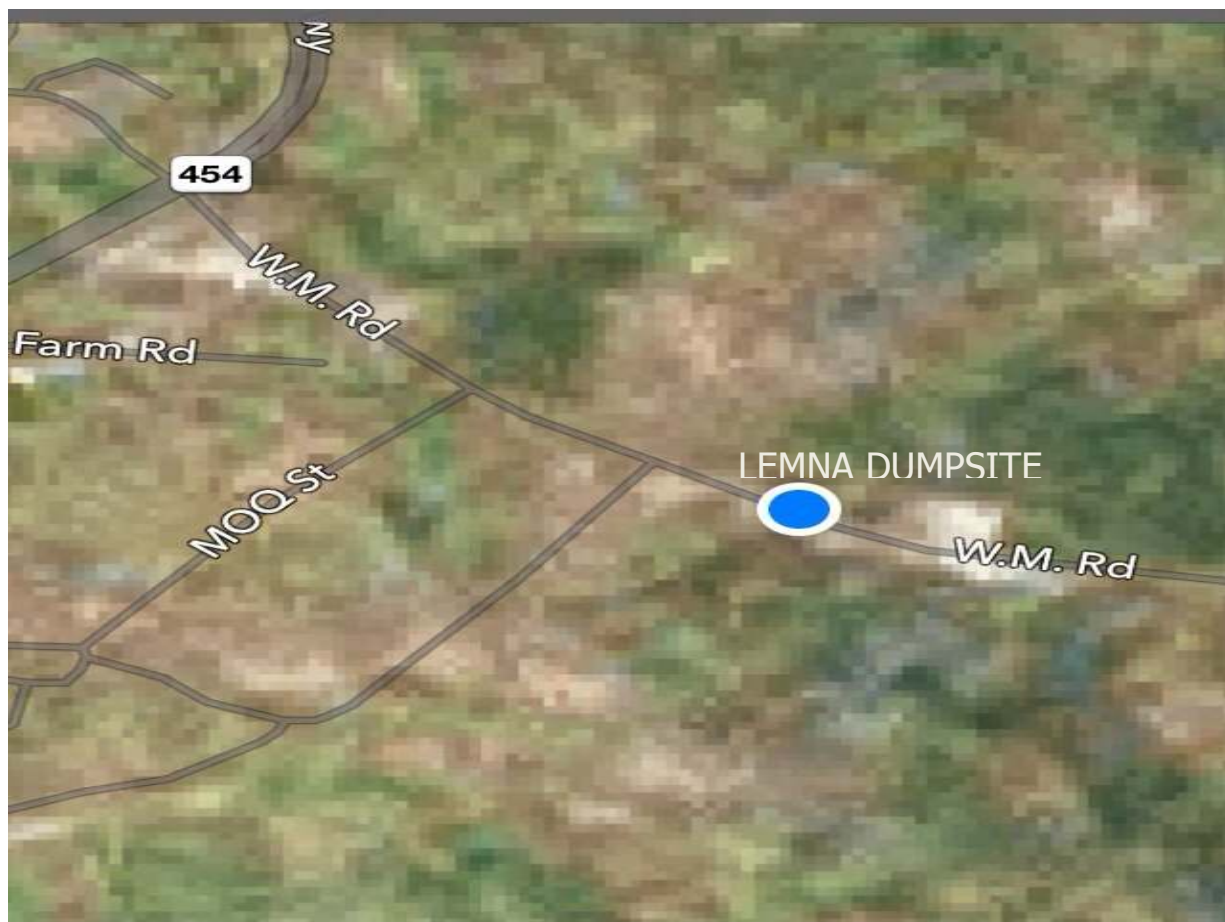
76 The Lemna dumpsite in Calabar Municipality which is where this
77 study was carried out, lies geographically along longitudes $08^{\circ}18'E$ and
78 $08^{\circ}26'E$ Greenwich meridian and latitudes $04^{\circ}55'N$ and $04^{\circ}58'N$ of
79 the equator. In the North, Calabar Municipality is bounded by
80 Odukpani and Akamkpa Local government Area, at the East by the
81 Great Kwa River. At the south, it is bounded by the Calabar River and
82 Calabar South Local government Area. It has an area of
83 331.551 square kilometres. Calabar municipality is characterized by a
84 double maxima rainfall regime which occurs in June and September.

85 The annual temperature is 28⁰cwith a high evapotranspiration and an
86 average humidity of 90%.

87 The vegetation of the study area is characterized by mangrove
88 swamp and rainforest, but due to human activities like cutting down of
89 tress, for roads, building of houses, schools, petrol stations, hotels and
90 market it has resulted in the depletion of the rainforest.

91 It has an annual rainfall of 3000mm and a harmattan wind blowing
92 over the area in December and January respectively.

93 The sampling site was a dump located within Lemma road
94 Calabar Municipality. It is a waste dump site used majorly by the
95 Calabar urban development authority. The dump site was measured
96 with tape and mater rule. The length of the landfill was 960m, the
97 width 430m. Sampling station was established on the waste dumpsite
98 and was represented as soil (S), leachate (L), solid waste (SW) and air
99 sample (AS).



101

Figure 1: Map of the study area

102



103

104 **Figure 2: Lemna road waste dumpsite**

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106

107

108 Sample Collection**109 Collection of Soil Sample**

110 For soil sample collection, the wastes were first removed using
111 a garden rake to expose the soil under the waste dump from where the
112 soil samples were collected. The soil samples were taken at about
113 15cm depth with the use of hand-driven auger and immediately
114 transported to the laboratory in labeled polythene bags in ice-cold
115 boxes at approximately 4°C for microbiological analysis. A control
116 sample was taken from a location of about 500 meters away from the
117 dumpsite.

118 Collection of Decomposing Solid Waste Sample

119 A sterile large wooden spatula that had been properly rinsed
120 with sterile distilled water was used to collect the decomposing wastes
121 into sterile glass petri- dishes and sealed with masking tape. The
122 samples were immediately transported to the laboratory for
123 microbiological analysis or stored in a refrigerator at 4°C until they
124 were needed for analysis.

125

126 Collection of Leachate Sample

127 The method described by Bassey *et al.*, 2015, was employed for
128 leachates sampling. In this method, Pvc pipes were cut into four parts,
129 each of 1m and 0.5m in length. The base end of each pipe was
130 permanently sealed with a pipe cover and an adhesive while the top
131 ends were just fitted with pipe covers. The pipes (both 1m and 0.5m
132 length) were perforated evenly at considerable distances from their
133 base ends to allow for water (leachates) percolation and collection.
134 The perforated pipes were then buried at the studied dumpsite for a
135 period of 3-4 weeks to allow percolation of leachates. After 3-4
136 weeks, sterile enema pumps or sterile syringes were used to collect the
137 leachate into sterile bottles and transported to the laboratory for
138 microbiological and physicochemical analysis.

139 Preparation of Diluent and Media

140 A ten-folds serial dilution of the sample was made by weighing one
141 gram of sample into 9mls of sterile distilled water to obtain 10^{-1}
142 dilution, further dilutions were made until 10^{-9} diluent was obtained.

143 The media used to grow bacterial were nutrient Agar, Eosin
144 methylene Blue Agar, *Salmonella-shigella* Agar, while the media used
145 to grow fungi was potato Dextrose Agar. All were originally in a
146 powdered form the media is usually prepared by weighing out a
147 certain gram, depending on the number of plates to be prepared. The
148 weighed quantity is then mixed with the appropriate amount of sterile
149 distilled water in a conical flask. The conical flask is usually plugged
150 with enough cotton wool and then placed in an autoclave for
151 sterilization at 121⁰c for 15mins after which the media is removed
152 from the autoclave and allowed to cool.

153 **Determination of P^H and Temperature**

154 The samples were made in suspension (solution) and p^H and
155 temperature probes were inserted into it. The p^H and temperature was
156 recorded accordingly.

157

158 **Microbiological analysis of soil and decomposing waste sample**

159 One gram(1g) of the sample was dissolved in 9ml sterile
160 distilled water from the solution, ten-fold serial dilutions in the range
161 of 10⁻¹ – 10⁻⁹ were prepared .One milliliter(1ml) aliquot from the

162 dilution of 10^{-4} of each samples were aseptically transferred into
163 freshly prepared nutrient agar plates then eosin methylene blue agar
164 and salmonella-shigella agar as selective media for isolation and
165 identification while 0.1ml from the dilution of 10^{-3} was aseptically
166 transferred into potato dextrose agar plates were incubated at 37°c (24
167 hours) for bacterial isolates and $28 \pm 0.2^{\circ}\text{c}$ (3 to 5 days) for fungal
168 isolates. Visible colonies of between 30-300 were multiplied by the
169 reciprocal of the dilution factors and recorded as colony forming units
170 per gram (cfu/g) of waste.

171 **Microbiological analysis of air at dumpsite**

172 Settle plates method described by Bassey et al., 2015, was
173 adopted for this analysis. According to this method, nutrient agar in
174 triplicate was exposed at the dumpsite (10m, 20m, 30m away from the
175 dump) 1hour between 10-12 noon. The plates were later covered and
176 transported to the laboratory for incubation at $25\text{-}28^{\circ}\text{c}$ for 24 hours. The
177 bacterial colonies that appeared on the plate were counted.

178 **3.6.3 Microbiological analysis of leachate**

179 Leachates from such a dumpsite are often turbid therefore, serial
180 dilutions were made. One milliliter (1ml) of the leachate was dissolved

181 in 9ml of sterile distilled water to give 10^{-1} dilution from which further
182 dilution up to 10^{-9} was made (Asikong and Udofia, 2005). The dilutions
183 were then further analyzed as those of decomposed waste and soil
184 sample previously described above. The colony counts were expressed
185 as colony forming units per milliliter of leachate (cfu/ml).

186 **Isolation, characterization and identification of bacteria in the waste**
187 **dump site**

188 Pure cultures of bacteria were obtained by aseptically streaking
189 representative colonies of different morphology types which appeared on
190 the culture plates onto freshly prepared Nutrient agar plates which were
191 incubated at 28°c for 24hours. Discrete bacteria colonies developed and
192 were sub-cultured on nutrient agar slope, incubated at 28°c for 24hours.
193 The nutrient agar slope served as pure culture stock for subsequent
194 characterization tests. The pure cultures were identified based on their
195 cultural, physiological and morphological characteristics. For tests
196 conducted for characterization and identification, see appendix.

197 **Isolation, characterization, identification of fungi in the waste**
198 **dumpsite**

199 Pure cultures of fungi were obtained by sub-culturing discrete
200 colonies into freshly prepared sabouraud dextrose agar plates and
201 inoculated at 28⁰c for 5-7 days. The fungal isolates which developed
202 were further sub-cultured onto agar slope and incubated at 28⁰c for 5 to
203 7days.The isolates which developed were pure cultures which were
204 stored in the refrigerator as stock cultures for subsequent characteristic
205 test. Macroscopic and microscopic examinations of fungal growth were
206 carried out.

207 **Determination of Greenhouse Gas Emissions From the Dumpsite**

208 The emission of greenhouse gases from the studied dumpsite was done
209 using GX-2012 gas detector

210

211

RESULTS

212 The frequency of occurrence of bacteria and fungi species isolated from
213 samples obtained from Lemna waste dumpsites are presented in Tables 1
214 and 2. The table(s) identifies the bacteria and fungi with the highest and
215 lowest frequency of occurrence. From the results obtained, *Escherichia*
216 *coli* had the highest frequency of occurrence while *Micrococcus luteus*
217 had the lowest frequency of occurrence Table 1. *Penicillium* were

218 observed as the most occurring fungal species while *Candida tropicalis*
219 had the lowest frequency of occurrence.

220 The mean bacterial and fungal counts of isolates from the different
221 sources sampled are presented in Tables 3 and 4. The results revealed
222 *Escherichia coli* with the highest mean counts of 27 ± 15.60 cfu/g of
223 decomposing solid wastes while *Pseudomonas aeruginosa* had the least
224 mean counts of 5 ± 3.00 cfu/g of decomposing solid wastes (Table 3).
225 Table 4 also identified the fungus with the highest and lowest mean
226 count for the different samples. *Candida albicans* had the highest mean
227 count of 20 ± 11.60 while *Candida tropicalis* had the lowest mean count
228 of 5 ± 3.00 . Table 5 and 6 showed the mean temperature and P^H values
229 for both soil and leachate samples obtained from the dumpsite.

230

231 **Table 1:** Distribution and frequency of occurrence of bacteria species
 232 isolated from the dumpsite samples

Isolated Bacteria species	Frequency of occurrence (%) of Bacteria from different sources.			
	S	L	SW	AS
<i>Escherichia coli</i>	80(16.50)	55 (10.83)	44 (17.0)	25 (7.50)
<i>Bacillus sp</i>	63(6.19)	48 (9.45)	39 (15.1)	30 (9.00)
<i>Pseudomonas aeruginosa</i>	52(10.72)	38 (7.48)	20 (7.72)	15 (4.50)
<i>Enterobacter sp</i>	37(7.63)	50 (9.84)	30 (11.50)	20 (6.00)
<i>Klebsiella sp</i>	49 (10.10)	33 (6.50)	27 (10.42)	40 (12.00)
<i>Proteus sp</i>	40(8.25)	44 (8.70)	15 (5.80)	38 (11.34)
<i>Salmonella sp</i>	39 (8.04)	30 (5.91)	18 (6.94)	40 (12.00)
<i>Staphylococcus aureus</i>	43 (8.87)	64 (12.50)	21 (8.11)	45(13.43)
<i>Micrococcus luteus</i>	29 (5.98)	74 (14.40)	- (-)	56 (16.71)
<i>Methanococcus sp</i>	53 (10.93)	73 (14.40)	45 (17.40)	26 (7.80)
Total	485	508	259	335

233

234 **Key:** S = soil, L = leachate, Sw = solid waste , AS = Air sample

235

236

237

238 **TABLE 2:** Distribution and frequency of occurrence of fungal species
 239 isolated from dumpsite samples.

Isolated Fungal Species	Frequency of occurrence (%) of Fungal from different sources			
	S	L	SW	AS
<i>Candida albicans</i>	60(22.00)	45(23.40)	35(16.00)	0(0)
<i>Aspergillus sp</i>	40(15.00)	30(15.62)	38(17.40)	25(45.50)
<i>Saccharomyces sp</i>	45(16.50)	32(16.70)	58(26.50)	30(45.60)
<i>Candida tropicalis</i>	20(7.33)	15(7.80)	18(8.23)	0(0)
<i>Penicillium sp</i>	58(21.23)	39(20.30)	50(22.83)	0(0)
<i>Candida parapsilosis</i>	50(18.30)	31(16.15)	20(9.13)	0(0)
Total	273	192	219	55

240

241 **TABLE 3:** Mean Count of bacteria species isolated from dumpsite
 242 samples.

Isolated Bacterial species	Mean count of Bacteria from different sources			
	S	L	SW	AS
<i>Escherichia coli</i>	27 ±15.6	18.3± 10.6	14.7±8.50	8.3 ± 4.8
<i>Bacillus sp</i>	21 ± 1.20	16 ± 9.30	13±8.00	10 ± 6.00
<i>Pseudomonas aeruginosa</i>	17 ± 1.00	12.7 ±7.3	7 ±4.00	5 ± 3.00
<i>Enterobacter sp</i>	12 ± 7.00	17 ± 9.60	10±6.00	7 ± 4.00
<i>Klebsiella sp</i>	16.3 ±9.40	11 ± 6.40	9±5.20	13.3±7.70
<i>Proteus sp</i>	13.3±7.70	15 ± 8.50	5±3.00	12.7±7.30

<i>Salmonella sp</i>	14.3 ±8.30	21.3 ±12.3	7 ±4.00	15 ± 8.70
<i>Staphylococcus aureus</i>	13 ± 8.00	10 ± 6.00	6±3.50	13.3±7.70
<i>Micrococcus luteus</i>	15.6 ±9.00	19.6 ±11.4	0(0)	22.0±13.00
<i>Methanococcus sp</i>	17.7 ± 10	24.3	15 ±8.70	8.6 ± 5.00
		±14.00		

243

244 **TABLE 4:** Mean Count of fungal species isolated from dumpsite
245 samples.

Isolated Fungal Species	Mean Count of Fungal from different sources			
	S	L	SW	AS
<i>Candida albicans</i>	20± 11.60	15 ±8.70	11.7 ± 6.70	0(0)
<i>Aspergillus sp</i>	13.3± 7.70	10±5.80	12±7.30	8.3±4.80
<i>Saccharomyces sp</i>	15± 8.70	10.7±6.20	19.3±11.20	10±6.00
<i>Candida tropicalis</i>	6.7± 4.00	5±3.00	6±3.50	0(0)
<i>Penicillium sp</i>	19.3± 11.20	13±7.30	16.7±9.60	0(0)
<i>Candida parapsilosis</i>	17± 9.60	10.3± 6.00	7± 4.00	0(0)
Total	273	192	219	55

246

247 **TABLE 5:** Temperature of the samples

Temperature Reading (Days)	Sample code			
	S ₁	S ₂	L ₁	L _{2a}
1	82	82	83	82

2 82 83 83 82

Total 164 165 166 164

Mean 82 82.5 83 82

248 **Key:** S_1 = soil sample I, S_2 = Soil sample II, L_1 = Leachate sample I,

249 L_2 = Leachate sample II

250 **Note:** Temperature in $^{\circ}\text{F}$ to $^{\circ}\text{C}$ ($T^{\circ}\text{f} - 32 \times 5/9$)

251 Temperature in $^{\circ}\text{C}$ to $^{\circ}\text{F}$ ($T^{\circ}\text{c} \times 9/5 + 32$)

252 **TABLE 6:** Mean pH values of samples collected from the refuse

253 dump site.

pH Reading (Days)	SAMPLE CODE			
	S_1	S_2	L_1	L_2
1	7.0	7.0	7.0	7.0
2	7.0	6.57	6.9	7.0
Total	14	13.6	13.9	14.0
Mean	7.0	6.98	6.95	7.0

254 **DISCUSSION**

255 The gradual increase of temperature around municipal solid waste
256 dumpsites within Calabar Metropolis were glaring over the years with serious
257 impacts to the immediate environment and biodiversity. The increase is
258 attributed to the continuous emissions of greenhouse gases (GHGs) from the
259 municipal solid waste dumpsites (Bassey, 2012; Bassey *et al.*, 2015). The
260 temperature values of soil and leachate samples from this study ranged from
261 82⁰F to 83⁰F. While the mean temperature value for soil samples was 82.3⁰F
262 while mean values for leachate was 83⁰F. The pH values ranged from 6.57 to
263 7.0. The mean pH value for soil sample was 6.9 while mean value was 7.0. The
264 observed temperature ranges in this study are such that support the growth of
265 mesophilic bacteria (mesophiles). Mesophiles are bacterial species involved in
266 biodegradation i.e digestion and decomposition of organic matter. During the
267 decomposition process, psychrophylic bacteria start the process (active at lower
268 temperature up to 55⁰F) and generate heat in the process. When the temperature
269 inside the decomposing layer reaches 50-100⁰F, the psychrophylic bacteria are
270 now displaced by the mesophilic bacteria which then continue the
271 biodegradation. The microbial activities of the mesophilic bacteria often elevate
272 the temperature of the compost which now gives way for the thermophilic flora
273 such as *Bacillus*, *Aspergillus* as reported in this study. The degree of acidity
274 (pH), reported in this study for all the stations of the waste dump site ranged

275 from 6.5 to 7.0. According to Hagerty *et al.*, (1973), they reported that, the
276 initial pH of solid waste is between pH 5.0 and 7.0 for refuse which is about 3
277 days old. The pH of the waste drops to 5.0 or less in the parts 2-3 days of
278 composing and then begins to rise to about 8.5 for the remainder of the aerobic
279 process.

280 According to the soil classification by Odu *et al.*, (1985), the degree of
281 acidity for sampled soil and leachate ranged from slightly acidic to neutral. The
282 frequency of occurrence (%) of isolated bacterial for the different samples
283 showed the organisms that occurred most. Among all the bacteria isolated from
284 the different samples only *Micrococcus* was not isolated from solid waste (SW).

285 The frequency of occurrence of isolated fungi for the different samples showed
286 the fungal that occurred most. Among all the fungi isolated from the different
287 samples *Candida albicans*, *Candida tropicalis*, *Penicillium spp*, *Candida*
288 *parapsilopsis* was not isolated from the Air sample.

289 The total aerobic bacterial count was highest in the sample leachate and
290 lowest in solid waste (SW). The order of decrease in the bacterial counts in all
291 the samples was solid waste (259) < Air < soil (485) < leachate (508). The total
292 viable fungal count was highest in the sample soil and lowest in Air. The order
293 of decrease in the fungal counts in all the samples was Air (552) < leachate
294 (192) < solid waste (219) < soil (273). The bacterial counts were higher than the

295 corresponding fungal courts. This is because the temperature range (82⁰f to 83⁰f)
296 supports the proliferation of the bacterial isolates.

297 This study shows the types of bacteria and fungi and their frequency of
298 isolation from the waste dump site at Lemna road. The bacteria isolated from the
299 dumpsite include: *Escherichia coli*, *Micrococcus luteus*, *Bacillus* sp, *Proteus* sp,
300 *Pseudomonas aeruginosa*, *enterobacter* sp, *Klebsiella* sp, *Salmonella* sp,
301 *Methanococcus* sp, *Staphylococcus aureus*, of all the different general of
302 bacteria isolated from the waste dumpsite, *Escherichia coli* had the highest
303 frequency of occurrence while *pseudomonas aeruginosa* had the lowest
304 frequency of occurrence.

305 All the bacterial isolates reported in this research have been reported to be
306 associated with waste and waste biodegradation. *Bacillus* and *Pseudomonas*
307 species were reported by Gray (1967) to be associated with waste. *Bacillus*, *E.*
308 *coli*, *Klebsiella* and *Pseudomonas* were also reported by cook et al, (1964).
309 Ekundayo, (1977) reported *Bacillus*, *E. coli*, *Proteus*, *Pseudomonas*,
310 *Staphylococcus*, *Streptococcus* among others. *Pseudomonas* has been widely
311 reported to be associated with waste (Sabry, 1992). Fungal species isolated from
312 the Lemna road dumpsite include *Candida albicans*, *Aspergillus* spp,
313 *Saccharomyces* sp, *Candida tropicalis*, *Penicillium* spp, *Candida parapsilosis*.
314 From the results obtained, *Candida tropicalis* recorded the lowest frequency of
315 occurrence while *Saccharomyces* sp recorded the highest frequency of isolation.

316 All fungi isolates reported in this study have been reported to be associated with
317 waste and waste biodegradation (Ekundayo, 1977; Ikpedu, 1980) reported
318 *Aspergillus* and *Saccharomyces*.

319 The study has revealed the presence of various bacteria and fungi known
320 to be associated with waste biodegradation with subsequent release of
321 greenhouse gases that has contributed immensely to the upscale of temperatures
322 within the study area. If the activities of these bacteria, fungi and yeast are
323 properly harnessed it can be used in future treatment plant in Nigeria in
324 escalating the bioconversion of waste compost into organic fertilizer for
325 optimum use in agriculture and horticulture. According to cook *et al.*, (1967)
326 and Monica Chesborough (2005) all the bacterial genera reported in this
327 research with exception of enterobacter are potential pathogens. Also, all the
328 fungal genera reported in this research with exception of *Penicilium* are
329 potential pathogens. The presence of these potential pathogens (bacterial and
330 fungal) reported in this study may be attributed to the disposal of raw human
331 faecal discharges and other human wastes at the dump site.

332 The indiscriminate dumping of waste around residential areas and other
333 ecologically sensitive area plays and inevitable role in global warming and also
334 pose serious health challenges. In a case where wastes are dumped
335 indiscriminately, microbes set in to carry out decomposition. During the process
336 of decomposition leachate are formed, if close to a river source the leachate

337 formed flows in and contaminate the river which then becomes hazardous to the
338 community nearby. Also, during the process of decomposition greenhouse gases
339 are formed such as methane which contributes to the depletion of the ozone
340 player. All these effect as a result of the indiscriminate dumping of waste cannot
341 be under estimated, so therefore, Nigeria as a whole should direct her efforts
342 towards the treatment of waste before disposal as to minimize the health hazards
343 associated with dumping of waste.

344 **Conclusion**

345 The present study reveals the potential hazards of indiscriminate dumping
346 of solid waste on the environment at large, with regards to global warming
347 contributions from waste biodegradation. Therefore, the results highlight the
348 importance of proper collection, transportation, disposal and treatment of solid
349 waste in an environmentally friendly manner that will not deface the aesthetics
350 of the ecosystem. Calabar with a vast rising population of over 400,000 people
351 has led to high generation of municipal solid waste with decreased space for
352 disposal. The high concentrations of carbon dioxide, methane and nitrous oxides
353 around the studied dumpsite is a wakeup call for more research tailored towards
354 generating necessary measures that can arrest the risk associated with the
355 continual emission of these greenhouse gases (GHGs) from the dumpsites.

356

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REFERENCES

- 358 Asikong, B. E. & Udofia, U. U. (2005). *Introduction to Environmental Microbiology,*
359 *Pollution and Waste Management.* Calabar: MABASS Printers, 105 – 182.
- 360 Bassey, I. U. & Brooks, A. A. (2012). Environmental and Public Health
361 Implications of Municipal Solid Waste Management Impacts of Lemna
362 Dumpsite, Calabar. Nigeria. *An Msc thesis.* 20-102.
- 363 Bassey, I. U., Brooks, A. A., Asikong, B. E. & Andy, I. E. (2015).
364 Environmental and Public Health Aspects of Solid Waste Management at
365 the Lemna Dumpsite in Calabar, Cross River State, Nigeria. *International*
366 *Journal of Tropical Diseases and Health.*
- 367 Cilinskis, E. & Zaloksnis, J (1996). Solid waste Management in the city of
368 Riga, Latvia: Objectives and Strategy. *Ambio.* 25:103-107.
- 369 Climate Change (2014). Synthesis Report Summary for Policy Makers. Pp
370 1-5.
- 371 Cook, H. A., Cremwell D. L., Wilson H. A.(1964). Micro-organisms in house
372 hold refuse and sewage water from sanitary landfills. *Proceedings, West*
373 *Virginia Academy of sciences.* 39(15):107-114..
- 374 Ekundayo, C.(2001). *Determination of the Impacts of Waste Management*
375 *Activities on Greenhouse Emission.* Submitted by KF consulting, Tonic-
376 Smith Associates and Environs _ RIS. 73(25): 127-178
- 377 Ekundayo, J. A. (1977). Environmental consequences of the pollution of the
378 Lagos Lagoon. *Bull Science Association Nigeria* 3(2):210-299.
- 379 Gray, K. R. (1967). Benefits of compost science. *Accelerated Compositing*
380 *Compost science.* 7(3):29-32.
- 381 Ikpendu, C. O. (1980). Microbial enzymes in the conversation of Local organic
382 wastes. In: S. O. Emejuaiwe, O. ogunbi and S. O. Sanni (Editors). *Global*
383 *Impact of Applied Microbiology.* 32(7): 1243-1257.
- 384 Khupe, J. S. N. (1996). Water supply, sewage and waste management for Gaborone,
385 Botswana. *Ambio,* 25, 134-137.
- 386 Inter-govenmental Panel on Climate Change (IPCC), (2014). *Summary for Policy*
387 *Makers. In: Climate Change 2014: Mitigation of Climate change.*
388 Contribution of Working Group III to the Fifth Assessment Report of the

- 389 Intergovernmental Panel on Climate change. Cambridge University Press,
390 Cambridge, United Kingdom and New York, NY, USA. Pp 1-29
- 391 Inter-govenmental Panel on Climate Change (IPCC), (2001).*IPCC Third*
392 *Assessment Report* (McCarthy J. J., Canziani. O. F. Leary N., kokken. D.
393 J. And White, K. S. Cambridge University Press, Cambridge. 44(21):
394 454-563.
- 395 Inter-govenmental Panel on Climate Change (IPCC), (1997).Guidelines for
396 National Greenhouse Gases inventories. In: Tsai: WR, Editor. Bio-energy
397 from landfill gas in Taiwan.*Elsevier Renewable and Sustainable Energy*
398 *Reviews, 11(13):331-44.*
- 399 Monica, C. (2005). *District Laboratory practice in Tropical Countries, part 2.*
400 Biochemical test to identify bacteria. 62-70.
- 401 Odu, C. T., Esuruoso, O. F., Nwoboshi, L. C. & Ogunwale, J. A. (1985).
402 Environmental study of the Nigerian Agip oil company, Operational Area.
403 Soil and Fresh Water Vegetation. Union Graft Publishing, Milan. Pp 22-
404 25.
- 405 Pavoni, J. L., Hoer, Jr. J. E. and Hagerty D. L. (1975).*Handbook of Solid Waste*
406 *Disposal, Materials and Energy Recovery.* Van Nostrand Reinhold
407 Company, New York. 38(18): 456-498.
- 408 Sabry, S. A. (1992).Microbial degredation of shrimp shell waste.*Journal of*
409 *Basic Microbiology.* 32(2): 107-111.
- 410 Stainer, R. Y., Ingraham, J. L., Whelis, M. L. & Painter, P. R. (1990). *General*
411 *microbiology* (5th ed.). London: Macmillan, 482-484.
- 412 Yakowitz, H. (1988). Identifying, classifying and describing Hazardous wastes.
413 In: A. L. Jacqueline (Editor). *Hazardous Waste Management; Industry*
414 *and Environment.*11 United Nations Environment Programme). 34(13):
415 234-256.
- 416 Yaliang, Y., (1996). Changzhou, China: Water supply, sewage treatment and waste
417 disposal strategies for sustainable development. *Ambio*, 25 (2), 86 – 89.