

Original Research Article

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**IMMUNOHISTOCHEMICAL PATTERN OF BREAST CANCER IN MAIDUGURI,
BORNO STATE**

**Key Words: IMMUNOHISTOCHEMICAL, PATTERN, BREAST, CANCER
MAIDUGURI**

ABSTRACT

Background: Breast cancer in women is a major public health problem throughout the world. It is the most common cancer among women both in developed and developing countries. One out of ten of all new cancers diagnosed worldwide each year, is a cancer of the female breast. It is also the principal cause of death from cancer among women globally.

Objectives: This is carried out to determine the immunohistochemical and histopathological patterns of breast cancer in Maiduguri.

Methodology: One hundred and fifty two cases of female breast cancer were retrieved from the archive of Department of Histopathology, University of Maiduguri Teaching Hospital. ER, PR and HER2 expression was assessed using immunohistochemical staining.

Results:Thirty one of the 152 cases were positive for either one or two of the hormonal antigen,while 121 (79.6%) were completely negative for any of the hormonal antigen, of the 31 positive cases, oestrogenreceptors were detected in 14 (45.2%) cases,progesterone were detected in 10 (32.2%) of the cancer cases while HER 2 were detected in 7 (22.6%). The mean age of all the subjects with breast cancer is 47.6% with highest prevalence at the age range of 32 – 58.Invasive ductal carcinoma account for 88.2% of the total breast cancer followed by invasive lobular carcinoma with 4.0%.

26 **Conclusion:**From this study most cases of breast cancer in this environment are hormone
27 receptor negative as found in most part of African continent in contrast to higher number of
28 hormone receptor positive cases in most western and Arabian countries.

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30 **1. Introduction**

31 Immunohistochemistry is a technique that combines anatomical, immunological and biochemical
32 techniques to identify discrete tissue components by the interaction of target antigens with
33 specific antibodies tagged with a visible label. Immunohistochemistry (IHC) has an expanding
34 role in the diagnosis and management of mammary disease [1]. A growing list of available
35 antibodies, improved antigen retrieval techniques, and a better understanding of biology have all
36 contributed to the broader utility of IHC for solving everyday diagnostic problems in breast
37 pathology [1].

38 The use of immunohistochemistry to further characterize breast cancer globally has introduced a
39 new dimension to our knowledge of the disease. Breast cancer can no longer be regarded as a
40 single entity and morphological features alone cannot completely predict the behavior of breast
41 cancer [2]. The three immunohistochemical markers currently in routine diagnostic use in most
42 countries are estrogen receptor (ER), progesterone receptor (PR) and Human epidermal growth
43 factor2(Her2). These markers determine which tumours are likely to respond to hormonal
44 therapy and Herceptin treatment [2]. It is generally acknowledged that breast cancer is a
45 heterogeneous disease with a wide spectrum of clinical, pathologic and molecular features. The
46 molecular classification is becoming the gold standard for complete characterization of breast
47 cancer and the underlying technology has already generated gene-profiling models to predict
48 outcomes [3]. Despite these remarkable achievements, in general, clinicians still rely on
49 traditional clinic pathologic features and readily available tumor markers such as estrogen

50 receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2
51 (HER2). ER, PR, and HER2, routinely available in breast cancer specimens, are reliable,
52 inexpensive, and useful for therapeutic decision making, and the results of these tests are
53 recorded in cancer registries allowing for population-based research which make them a
54 reasonable substitute for the more expensive molecular sub typing [4].

55 Breast cancer in women is a major public health problem throughout the world. It is the most
56 common cancer among women both in developed and developing countries [5]. One out of ten of
57 all new cancers diagnosed worldwide each year, is a cancer of the female breast [5]. It is also the
58 principal cause of death from cancer among women globally. More than 1.38 million cases of
59 breast cancer are diagnosed world-wide in 2008, representing 10.9 % of all cancer [5].

60 It is the second most common cancer now, after lung cancer, when ranked by cancer occurrence
61 in both sexes. About 55% of the global burden is currently experienced in developed countries,
62 but incidence rates are rapidly rising in developing countries [5].

63 In the National Cancer Institute, breast cancer came as number one in ranking malignant tumors
64 constituting 17.5% of total malignancies. Females showed a vast majority of 98.35%, while only
65 1.65% were males [6]. Ductal carcinoma formed a majority of 85.02%, 2.04% of which were
66 intraduct carcinomas. Hormone receptors were positive in 57.8% of cases, while Her-2/neu was
67 positive in 44.5% of cases. Lymph nodes were positive for metastasis in 69.5% of cases [5].

68 Breast cancer is a heterogeneous disease whose evolution is difficult to predict.

69 Consequently, treatment is not as adapted as it should be. Gene expression studies have
70 identified five molecularly distinct subtypes of breast cancer that have prognostic value across
71 multiple treatments and can predict distinct clinical outcomes. These subtypes are termed
72 hormone receptor(s) positive luminal A (luminal A), hormone receptor(s) positive luminal B,

73 luminal HER2/neu, HER2-enriched (i.e, tumors that over express ERBB2-associated genes but
74 do not express genes that define the luminal subtype) and basal-like (triple negative) [7]. These
75 subtypes are associated with differences in clinical outcome, HER2-enriched and basal-like
76 subtypes are hormone receptor negative and have poorer prognosis with shorter survival times
77 than other types [8] .

78 In contrast, the expression of hormone receptor(s) characterizes the luminal breast cancers, with
79 luminal B tumors having intermediate survival time & poorer outcomes than luminal A tumors
80 having the longest survival [9].

81 Although some luminal B tumors can be identified by their expression of HER2, the major
82 biological distinction between luminal A and B is the proliferation signature, including genes
83 such as MKI67 (encoding Ki67), which has higher expression in luminal B tumors than in
84 luminal A tumors. Thus, a distinction between luminal A and B tumors that is based on
85 proliferation status among hormone receptor(s) positive luminal patients may be important to
86 breast cancer biology and prognosis since luminal B tumors having a higher rate of tumor cell
87 proliferation and poorer prognosis than luminal A tumors. Thus luminal A and B breast cancers
88 appear to be distinguished by the expression of estrogen receptor (ER), progesterone receptor
89 (PR), HER2, and Ki-67 proteins [10].

90 The Nottingham modification of the Scarff-Bloom-Richardson (NSBR) histological grading
91 system for invasive breast cancer has been recommended by the World Health Organization
92 (WHO) [11] .

93 In the NSBR system, histological grading consists of three components: tubule formation,
94 nuclear pleomorphism and mitotic count. Each of these are allocated a score of 1–3, and the final
95 histological grade is determined according to the sum of the three components (grade 1: sum=3–

96 5; grade 2: sum=6–7; and grade 3: sum=8–9). Patients with the luminal A subtype were less
97 likely to have grade 3 tumors while patients with triple negative tumors had the greatest
98 likelihood of having grade 3. The high cost of gene expression profiling has limited its
99 incorporation into most randomized clinical trials, and therefore, immunohistochemistry-based
100 surrogate assay is proposed to distinguish between various breast cancer subtypes with emphasis
101 on the role of the Ki-67 labeling index as a clinically valuable biomarker for the luminal B
102 subtype [12].

103 **2. Methodology**

104 **Study area**

105 The study was carried out at the Department of Histopathology University of Maiduguri
106 Teaching Hospital, Maiduguri.

107 **Study design**

108 Formalin fixed paraffin embedded sample was obtained from the archive of the Department of
109 Histopathology UMTH. 5 years (January 2011- December 2015) breast cancer positive cases were
110 considered. The case to study composed of all diagnosed breast cancers one representative block
111 was selected from each case if more than one block were retrieved from the archive.

112 **Immunohistochemical method**

113 Paraffin blocks was sectioned at four micrometer thickness, mounted on a slide and placed in the
114 oven for 30mins. Section was deparaffinised by passage through changes of xylene 5 min each
115 and subsequently rehydrated in descending grades of alcohol. It was then washed in buffer. The
116 slide was incubated in hydrogen peroxide block for 10 minutes (to reduce non specific

117 background staining due to endogenous peroxidase).It was then washed 4 times in buffer,ultra
118 V block was applied and incubated for 5 minutes to block nonspecific background staining.
119 primary antibody was applied for 30 minutes, then washed 4 times in buffer,primary antibody
120 enhancer was applied and incubated for 10 minutes at room temperature,HPR polymer was
121 applied and incubated for 15 minutes at room temperature,it was then washed 4 times in buffer,
122 1 drop of DAB plus chromogen substrate was added to 2mls of DAB plus substrate. It was
123 mixed, applied to the tissue and it was finally washed 4times in distilled water,it was
124 counterstain with heamatoxlyne and mount with DPX mountant [13].

125 **Interpretation of slides**

126 Staining intensity of immunohistochemically stained sections was semiquantitatively evaluated
127 using the Quickscore scoring system for PR and ER and DAKO scoring system for HER2.

128 The proportion of positive cells(scored on a scale of 0 to 5) and staining intensity (scored on a
129 scale of 0 to 3) were summed to produce total scores of 0 to 2 though 8.A score of 0 to 2 were
130 regarded as negative while 3 to 8 as positive. For HER2, a zero score defines tumors with no
131 staining or membrane staining in less than 10% of the tumor cells, while 1+ refers to tumors with
132 a faint membrane staining in more than 10% of the tumor cells. A weakly positive result
133 characterized by weak to moderate complete membrane staining in more than 10% of the tumor
134 cells is represented by a 2+ score, while a strongly positive result defined as strong complete
135 membrane staining in more than 10% of the tumor cells is represented as 3+. Scores of 0, 1+ was
136 classified as negative, while a score of 2+ and 3+ Was regarded as positive [14].

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139 **3. Results**

140 The result of the study carried out to determine the immunohistochemical pattern of breast
141 cancer in Maiduguri over the period of five years revealed a breast cancer prevalence of 13.9%.A
142 total of one hundred and fifty two (152) cases of breast cancer specimen found over the period of
143 the study had immunohistochemistry done on them..The result revealed only 31(20.4%) of the
144 one hundred and fifty cases of breast cancer were positive for either one or two of the hormonal
145 antigen while 121 (79.6%) were completely negative for any of the hormonal antigen.Of this 31
146 positive cases, oestrogen receptor were detected in 14(45.2%) cases,progesterone receptor were
147 detected in 10(32.2%) of the cancer cases while HER2 were detected in 7(22.6%) of all breast
148 cancer cases.(Table 4.1). The mean age of all subjects with brain cancer is 46.7 (53.3%) with
149 highest prevalence of cancer at the age range of 32 -52 followed closely by 53- 67 age range
150 having 23% prevalence. (Table 4.2). The result of histopathological pattern of the breast cancer
151 in this environment showed 134 (88.2%) were invasive ductal carcinoma followed by invasive
152 lobular carcinoma (4.0 %) and the other ranging from 1-2% prevalence. (Table4 .3)

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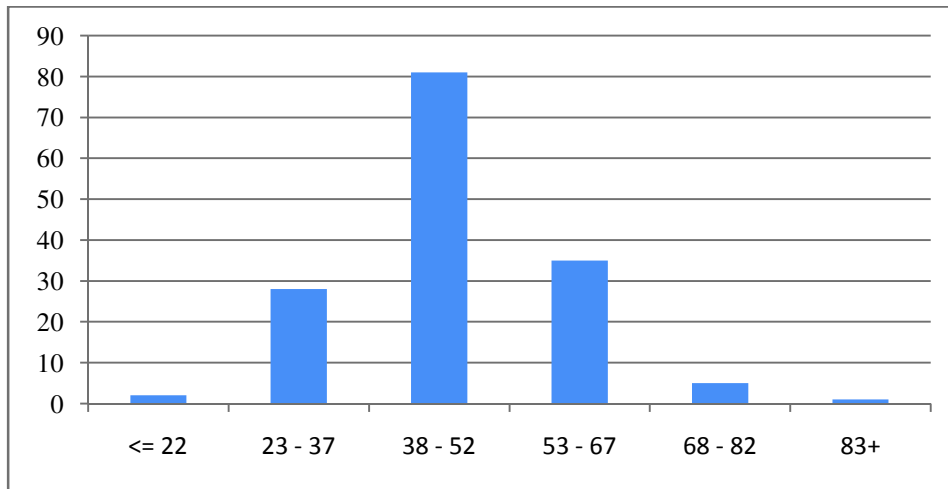
154 **Table 4.1: Frequency of distribution of breast cancer patients by age groups**

Age group	Frequency	Percent
<= 22	2	1.3
23 – 37	28	18.4
38 – 52	81	53.3
53 – 67	35	23.0
68 – 82	5	3.3
83+	1	.7
Total	152	100.0

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157 **Figure 4.1; Histogram of the frequency distribution by age groups of the Patients**



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161 **Table 4.2: Distribution of breast cancer by clinicopathological features**

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DIAGNOSIS	FREQUENCY	PERCENT
IDCA	134	88.2
METAPLASMIC CA	1	.7
ILCA	6	4.0
MEDULLA CA.	2	1.3
INV. PAPILLARY CA	5	3.3
ADENO CA	1	.7
APOCINE CA	1	.7
MUCINOUS CA	1	.7
CARCINOSARCOMA	1	.7
Total	152	100.0

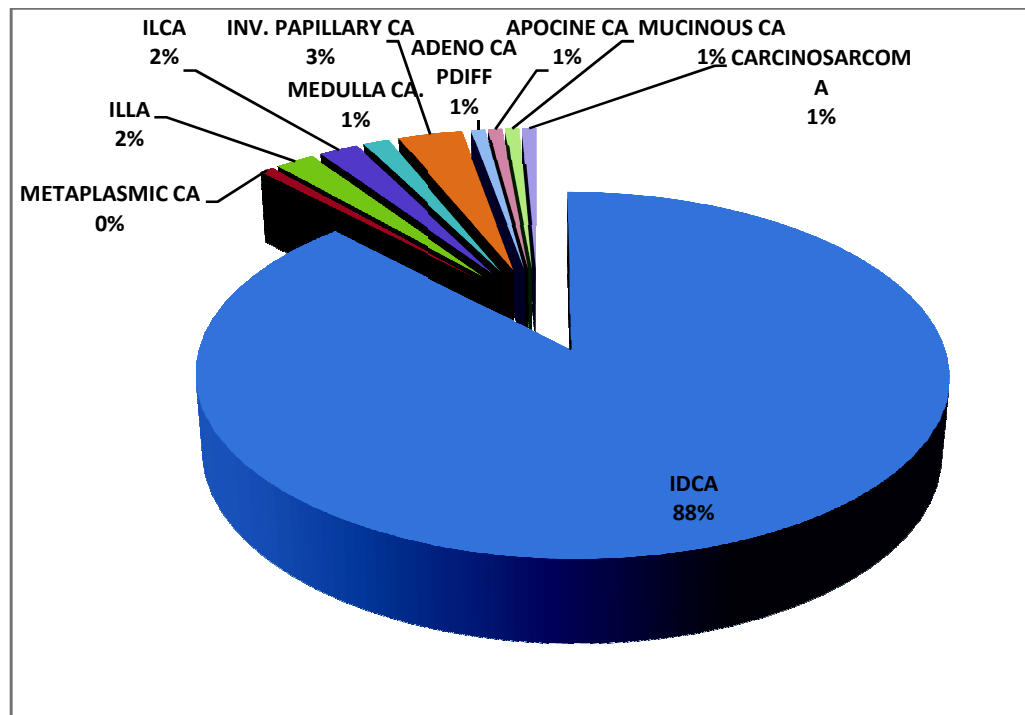
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167 **Figure 4.2: Chart of breast cancer by clinicopathological features**



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179 **Table 4.3: Expression of ER, PR and HER2 in cases**

Marker	Positive (>3)	Negative (0-2)	Total
ER	14 (45.2%)	37(72.5%)	51
PR	10 (32.2%)	41 (80.4%)	51
HER2	7 (22.6 %)	43 (86%)	50
Total	31	121	152

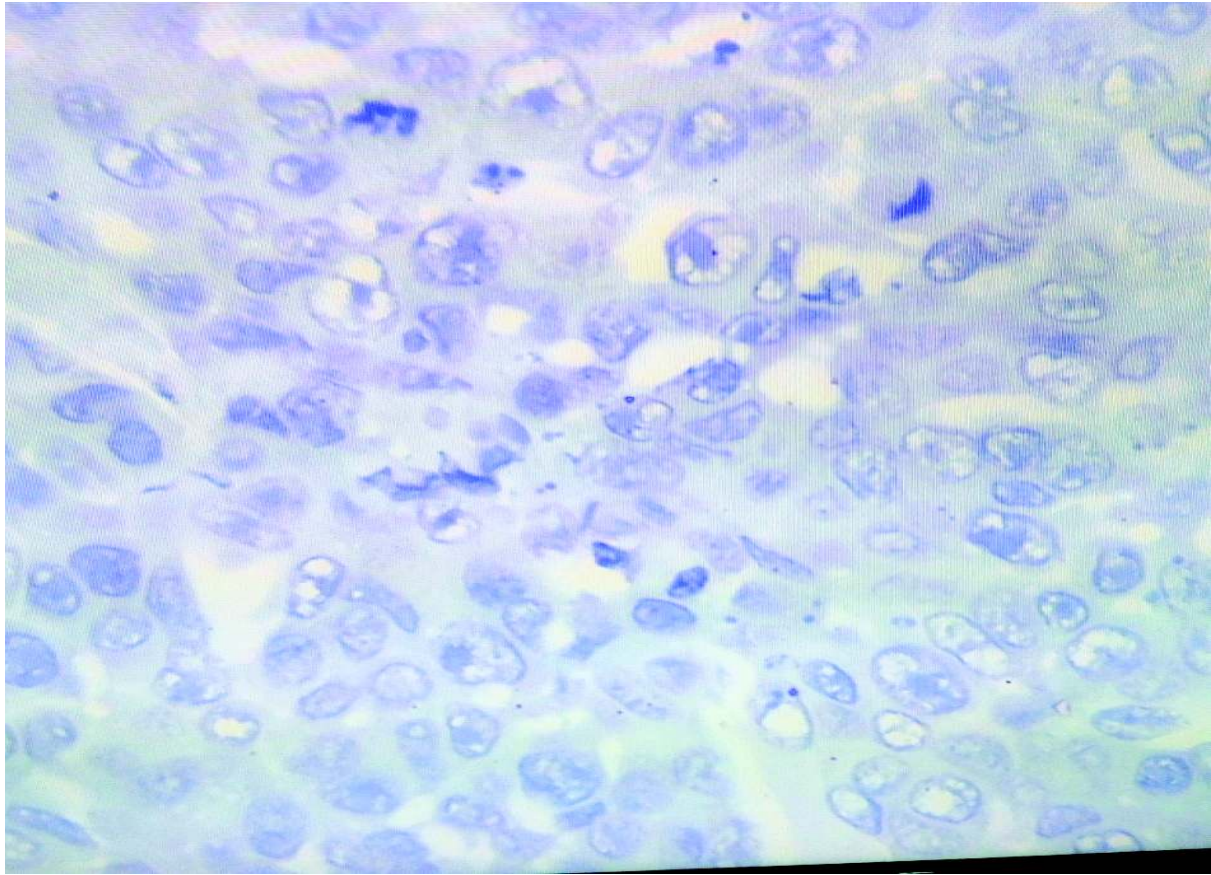
180 ER=Estrogen receptor; PR=Progesterone receptor; HER2/neu=Human epidermal growth factor

181 receptor 2

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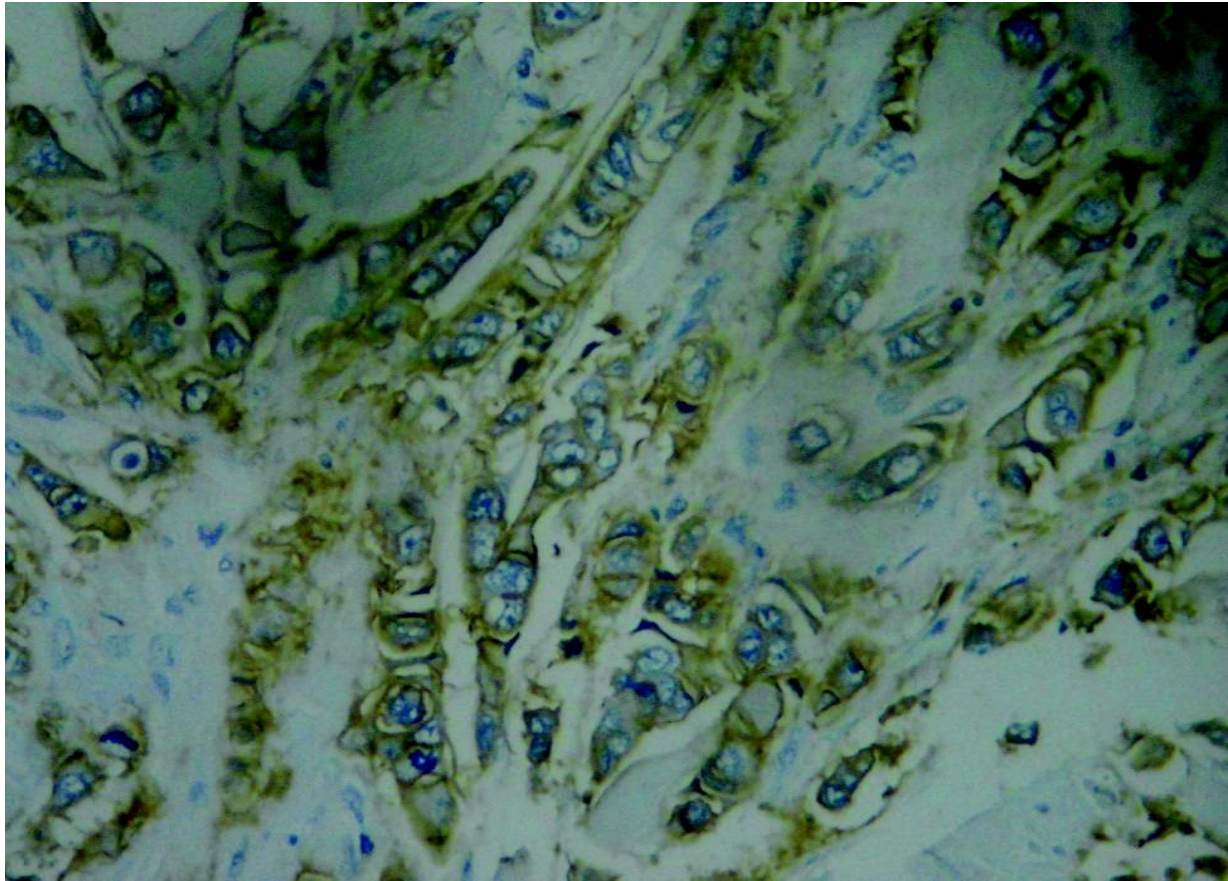


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186 **Fig4.3 Photomicrograph of IDC showing negative membrane staining for HER2 X 100**

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190 **Fig 4.4 Photomicrograph of IDC showing positive membrane staining for HER2 X 100**

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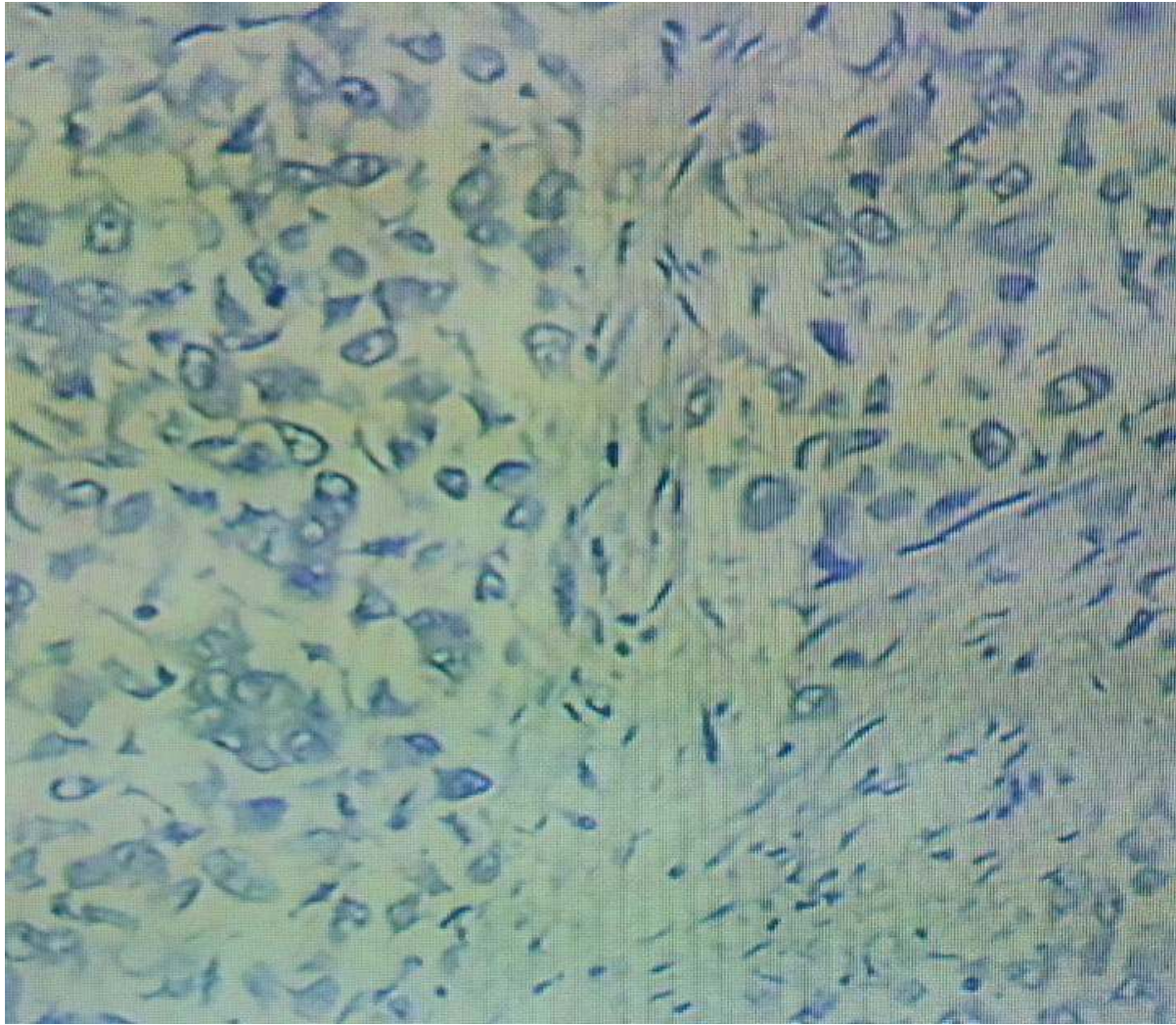
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199 **Fig 4.5 photomicrograph of IDC showing negative nuclei staining for ER X100**

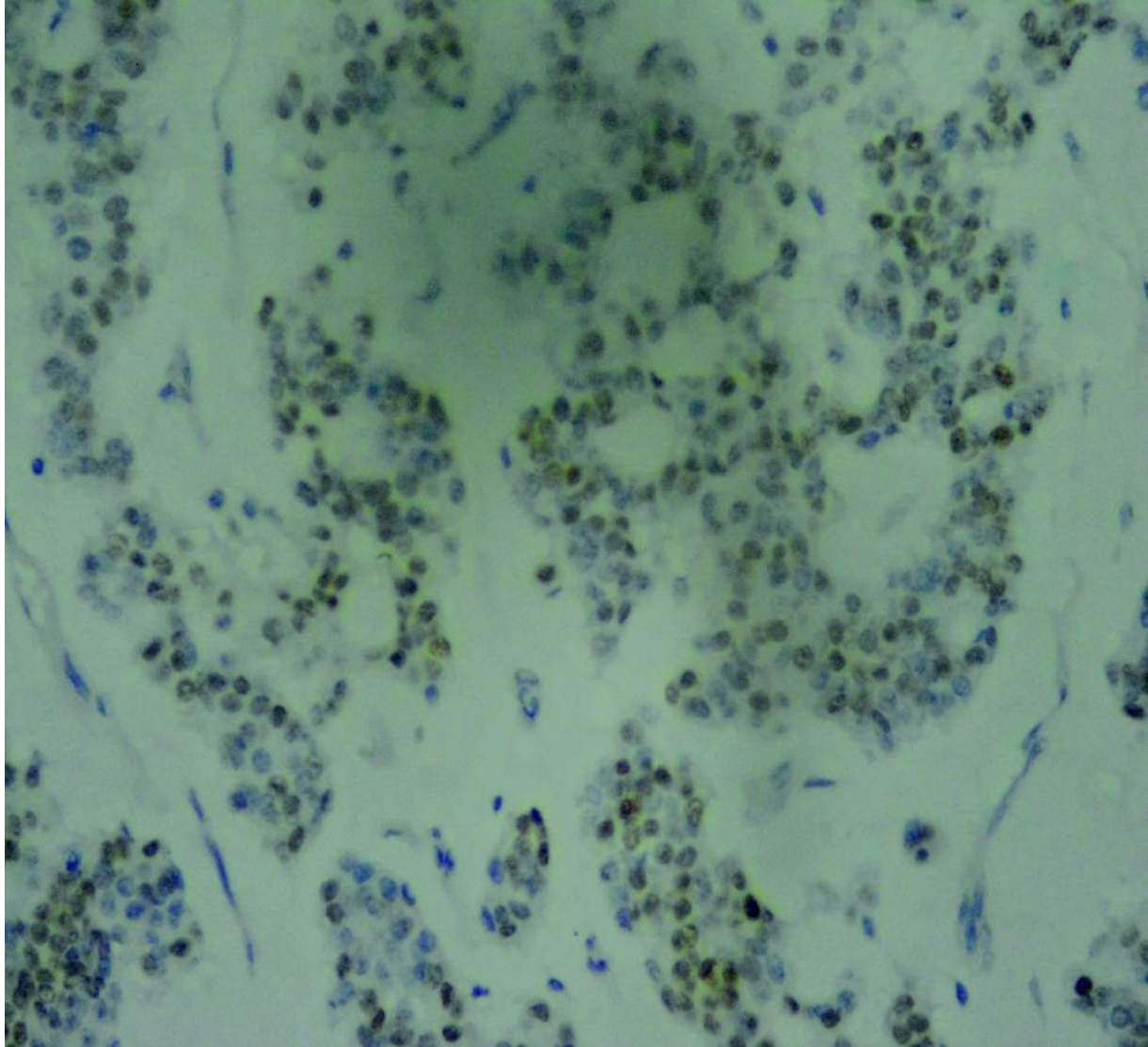
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206 **Fig 4.6 Photomicrograph of IDC showing positive nuclei staining for ER X100**

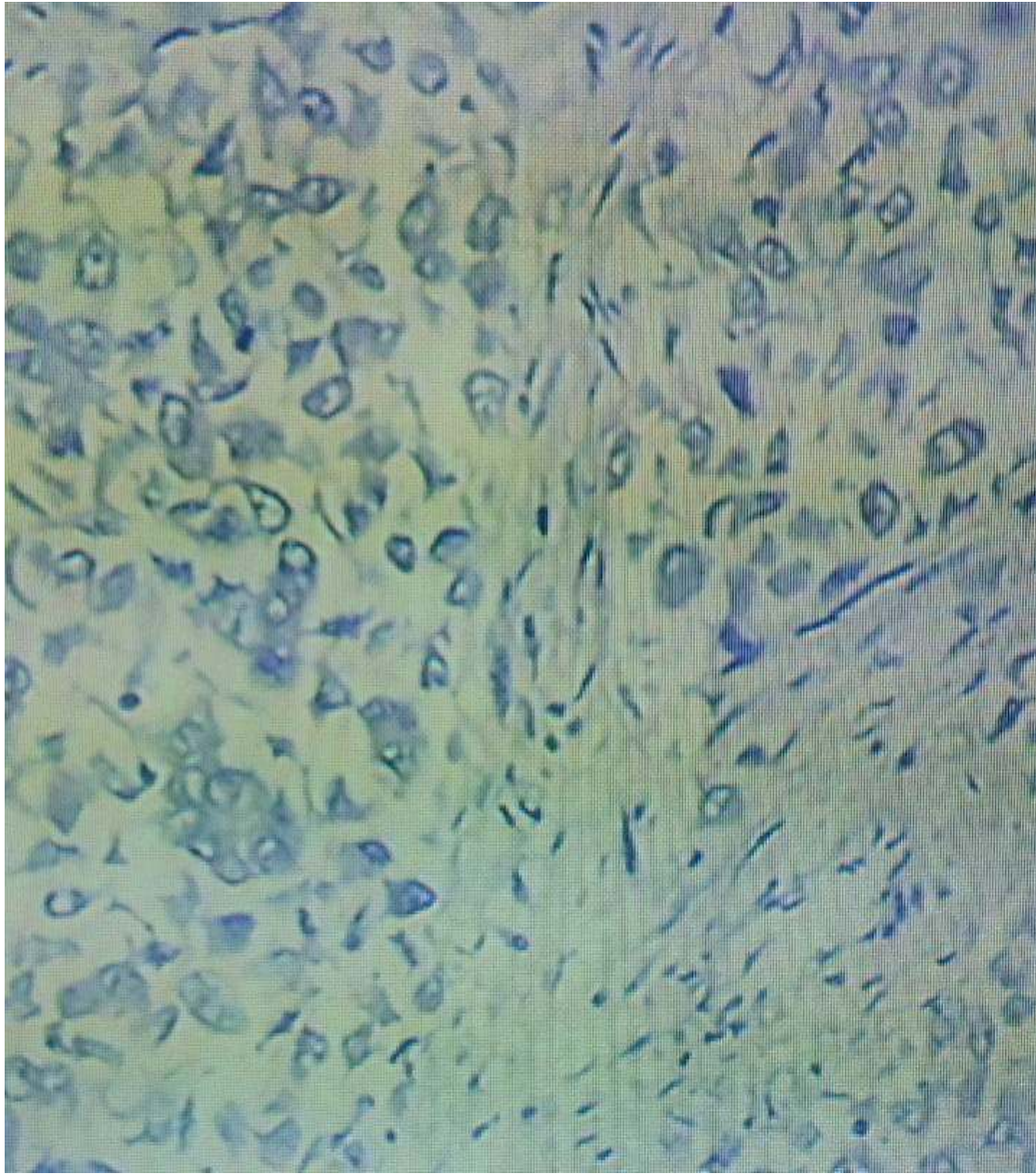
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213 **Fig 4.7 photomicrograph of IDC showing negative nuclei staining for PR X 100**

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217 4. Discussion

218 Immunohistochemistry based classification of both ER, PR, and HER2 status provide prognostic
219 and therapeutic information not achievable from either alone. The use of IHC in breast cancer has
220 become an integral part of a complete and comprehensive histopathology report, in terms of
221 prognosis and prediction of response to treatment, in addition to histological grade and tumor sub
222 types, hormone marker ER, PR and HER2 has become the mainstay requirement for the
223 oncologist in the developed world, assessment for hormonal receptors expression status is
224 required to determine patient eligibility for hormonal therapy. However, in the developing
225 countries clinicians administer hormonal therapy without any knowledge of their patient
226 receptors status. ER, PR and HER2 expression status is not routinely determined in the
227 developing countries because of limited resources and relatively high cost of testing.

228 The result of the immunohistochemical pattern of breast cancer in this study revealed that ER
229 was positive in 45.2%, PR was positive in 32.2 % while HER 2 was positive in 22% cases.

230 This is a little slightly lower than the report carried out in Ibadan by [1] that show 65.1% ER
231 positively, 54.7% PR positively and 79.7% HER 2 negative. But inline with the report of
232 Nwotoret *al.*,2014 with ER positive in 54.2% cases while PR was seen in 50% with HER 2
233 present in 31%. Recently [15] reported a similar study in Abuja with ER positive in 46.3% and
234 PR positive in 42.6%.

235 In Ile-Ife a studied carried out by [16] reported ER positively in 34.6% PR positively in 25% and
236 HER 2 positivity in 38.2% which is also in line with this study.

237 In Ghana, it was reported an ER, PR and HER2 receptor positivity of 32.1%, 25.6% and 22.5%
238 respectively, recently in AI Khobor Saudi Arabia (S.A) the rate of positive hormone receptor and

239 HER2 in breast cancer using IHC were 69.2%, 61.5 % and 25.1% for ER, PR and HER2
240 respectively. In China ER was positive in 53%, PR was positive in 51.5% and HER 2 in 46.2%
241 [17]. In the Arabian countries, the frequency of the IHC positive hormone receptor and HER2
242 show great variation, Runnak and colleagues in 2012 investigated 514 cases of breast cancer in
243 Iraq females of different origin, Arabic and Kurdish, they found that 73% were ER positive,
244 64.2% where PR positive only 20.4% of breast cancer cases were HER2 positive. The low rate of
245 IHC staining positive for ER, PR and HER22 in Maiduguri is in harmony and fall in the same
246 range of other populations in Nigeria [18] and Ghana on the other hand the rate of positivity in
247 ER, PR and HER2 in Iraq, Egypt and USA [19, 20]. Shows high rate of positivity.

248 Alternatively contributing factor to those finding could be biological and lifestyle aspect.

249 The mean age of all subject in the study was 46.7 years, this is similar to mean age of 49.7 years,
250 48.1 years and 47.5 years reported in Nigeria, Senegal and India respectively but less than mean
251 age of 55-58 years reported in Western countries like USA [21].

252 This might be as a result of good screening programme in this developed countries and also
253 presence of good diagnostic facility that will enable early diagnosis and treatment.

254 The majority of breast cancer in this study were Invasive ductal carcinoma with 88.2%.

255 **5. Conclusion**

256 From this study, it can be concluded that most cases of breast cancers are hormone receptor
257 negative as found in most part of the African continent in contrast to highest number of hormone
258 receptor positive cases of breast cancer in most Western and Arabian countries. The prevalence
259 of hormone receptors positive breast cancer stand at 20.4% with ER accounting four 45.2% of

260 the hormone receptor positive cases while PR positive account for 32.2% and HER 22.6%.The
261 mean age of the subject is 46.7. The histopathological pattern of breast cancer in this study
262 revealed that 88.2% of all breast cancer are invasive ductal carcinoma.

263 REFERENCES

- 264 [1]. Adebamowo CA, Famooto A., Ogundiran T. O., Aniagwu T., Nkwodimmah C., and Akang
265 E. E. (2008). Immunohistochemical and molecular subtypes of breast cancer in Nigeria,
266 *Breast Cancer Research and Treatment*; **110**: 183-188.
- 267 [2]. Bauer K.R., Brown M., Cress R.D., Parise C.A., and Caggiano V. (2007). Descriptive
268 analysis of estrogen receptor (ER)-negative, progesterone receptor (PR)-negative, and HER2-
269 negative invasive breast cancer, the so-called triple-negative phenotype: a population-based
270 study from the California cancer Registry. *Cancer*; 1721–1728.
- 271
- 272 [3]. Berry D. A., Cirrincione C., Henderson I. C., Citron M. L., and Budman D. R.
273 (2006).Estrogen receptor status and outcomes of modern chemotherapy for patients withnode
274 positive breast cancer.*Journal of American Medicine* 295: 1658-1667.
- 275
- 276 [4]. Bocker W., Bier B., and Freytag G. (1992). An immunohistochemical study of the breast
277 using antibodies to basal and luminal keratins, alpha-smooth muscle actin, vimentin,
278 collagen IV and laminin, part II: epitheliosis and ductal carcinoma insitu.*VirchowsArchieves*
279 *of Pathology Anatomic Histopathology*.**421**: 323–330.
- 280 [5]. Farley J., Shin H. R., and Bray F. (2010). Estimation of worldwide burden of cancer in
281 2008.*International Journal of Cancer*,**127**,pp. 2893-2917.

- 282 [6]. Mokhtal A, and Shaghir E.I (2007).Estrogen-receptor status and outcomes of modern
283 chemotherapy for patients with node-positive breast cancer.*Journal of American*
284 *Medicine***295**: 1658–1667.
- 285 [7]. Carley L. A., Dees E. C., and Sawyer L. (2012).The tripple negative paradox:primary tumor
286 sensitivity of breast cancer. *Clinical cancer Research*, **13**:2329-2334.
287
- 288 [8]. Sorlie T., Tibshirani R., Parker J., Hastie T., Mairon J. S., and Nobal A. H. (2003).Repeated
289 observation of breast tumor subtype in independent gene expression data sets. *Journal of*
290 *National Health Academic Sciences USA*.**100**: 8418-8423
- 291 [9]. Cheang M. C., Chia S. K., Voduc D., Gao D., and Leung S. (2009). Ki67 index, HER2
292 status, and prognosis of patients with luminal B breast cancer. *Journal of National Cancer*
293 *Institute* **101**: 736–750.
294
- 295 [10]. Klintman M., Bendahl P. O., Grabau D., Lövgren K., Malmström P., and Fernö M.
296 (2010). South Sweden Breast Cancer Group. The prognostic value of Ki67 is dependent on
297 estrogen receptor status and histological grade in premenopausal patients with nodenegative
298 breast cancer. *Modern Pathology* **23**(2): 251–259.
- 299 [11]. Dent R., Trudeau M., Pritchard K. I., Hanna W. M., Kahn H. K., Sawka C. A., Lickley LA,
300 Rawlinson E., Sun P., and Narod S. A. (2007). Triple-negative breast cancer: clinical
301 features and patterns of recurrence. *Clinical Cancer Research***13**: 4429–4434.
302

- 303 [12]. Nelson HD, Humphrey LL, Nygren P, Teutsch SM, Allan JD(2004): Postmenopausal
304 hormone replacement therapy: scientific review. *Journal of American*
305 *Medicine* 288:872.
- 306 [13]. Shan-Rong Shi, James Guo, Cote L. C., Debra Hawes, Yan Shi, Sandra Thu, and Clive R.
307 (1999).*Applied Immunohistochemistry and Molecular Morphology*.7: 201-208.
308
- 309 [14]. Hammond M. E., Hayes D. F., and Dowsett M. (2010). American Society of Clinical
310 Oncology/College of American Pathologists guideline recommendations for
311 immunohistochemical testing of estrogen and progesterone receptors in breastcancer.*Journal*
312 *of Clinical Oncology*, **28**: 2784–2795
313
- 314 [15]. Madukwe U.A, Jonathan I, and Obama Yibala (2016). Triple negative breast cancer in a
315 private immunohistochemistry laboratory in Abuja, *Nigeria advance in biological*
316 *research***10**(1): 58-64.
317
- 318 [16]. OmoniyiEsan O. O., Olaosa O. O., Aremu O. A. and Omonisi A. E. (2015). Hormonal and
319 HER2 receptor Immunohistochemistry of breast cancer in Ile-Ife Nigeira, *Austine journal*
320 *of women health* ,**3**: 121-123
- 321 [17]. Chow L. W., and Hop A. A. (2000). Hormonal receptor determination of 1,052 Chinese
322 breast cancers. *Journal of Surgical Oncology*, **3**: 172-5.
- 323 [18]. Nwotor C.C, and So Keshinro (2014). Pattern of hormone receptor and human epidermal

- 324 growth factor receptor 2 status in sub Saharan breast cancer cases, *Nigeria journal of*
325 *chemical practice*,**18**:553-558.
- 326 [19]. Sterer M, Rosen H, and Weber R (1993). Immunohistochemical and biochemical
327 measurement of estrogen and progesterone receptors in primary breast cancer.
328 correlation of histopathology and prognostic factors. *Journal of Annals of Surgery***1**: 13-
329 21
- 330 [20]. Iqbal M., Davies M. P., Shoker B. S., Jarvis C., Sibson D. R. and Sloane J. P. (2001).
331 Subgroups of non-atypical hyperplasia of breast defined by proliferation of oestrogen
332 receptor-positive cells. *Journal of Pathology* ,**193**:333.
333
- 334 [21]. Kallel I., Khabir A., and Boujelbene N. L. (2012). EGFR overexpression relates to triple
335 negative profile and poor prognosis in breast cancer patients in Tunisia. *Journal of*
336 *Receptor Signal Transduction Research***3**: 142-149
337