

GENERATION OF BIOELECTRICITY AND SIMULTANEOUS TREATMENT OF WASTE WATER USING MICROBIAL FUEL CELLS (MFC)

ABSTRACT

This era is definitely an era of renewable energy generated from resource which are naturally replenished on a human timescale. The human inquisitiveness in search of a sustainable and environment friendly energy source has led to several advancement in this field. The microbial fuel cell (MFC) is a new form of renewable energy technology that can generate electricity from what would otherwise be considered waste. The reasons for this recent interest in using bacteria to generate electricity are a combination of the need for new sources of energy, discoveries about microbial physiology related to electron transport, and advancement of fuel cell technologies. The MFC performance was successfully carried out for about 40 days with two different microbes (*E.coli* and *P.aeruginosa*) which were isolated, inoculated and identified by Gram's staining. Biochemical analysis confirms the purity of the respective microorganisms. The physical parameters estimated and studied showed *E.coli* as an efficient source of degradation for BOD, TDS, COD and DO in all the selected samples. The efficiency of salt degradation like nitrate, chloride, sulphate and phosphates) was shown more by *P.aeruginosa* compared to *E.coli* in all the samples. Further the efficiency of microbes in degrading waste materials and simultaneous production of electricity was statistically proven with ANOVA table showing the best voltage production in two samples by *P.aeruginosa* (419.8mV and 380.7mV) in lake water and apartment samples respectively. Similarly, the third sample collected from sewage treatment plant (STP) the maximum volt efficiency of 344.16mV was shown by *E.coli*.

Keywords : (Renewable energy, Bacteria, *E.coli*, *P.aeruginosa*, Biodegradation, Electricity, Microbial fuel cell)

1. INTRODUCTION

Energy calamity in India is rising each year, as there is constant acclivity in the price of fuels and also due to depletion of fossil fuels to a larger level. The demand for an alternative fuel has erupted extensive research in discovering a potential, economical and reusable source for energy manufacturers. For constructing a sustainable world we require to minimize the expenditure of fossil fuels as well as the pollution generated.

34 These two aims can be accomplished all together by treating the waste water (From
35 disposing waste to using it). Industrial waste, agricultural waste and household waste are
36 ideal substrates for energy productions as they are rich in organic contents. .

37 Microbial fuel cell (MFCs) have emerged in recent years as a promising yet challenging
38 technology. MFCs are the major type of bio-electrochemical systems (BESs) which
39 convert biomass spontaneously into electricity through the metabolic activity of the
40 microorganisms. MFC is considered to be a promising sustainable technology to meet
41 increasing energy needs, especially using wastewaters as substrates, which can
42 generate electricity and accomplish wastewater treatment simultaneously.

43 Microbial fuel cells are devices that directly convert chemical energy to electricity through
44 catalytic activities of microorganisms. Electricity has been generated in MFCs from
45 various organic compounds including carbohydrates, proteins and fatty acids. One of the
46 greatest advantages of MFCs over conventional fuel cells like hydrogen and methanol
47 fuel cell is that a diverse range of Organic material can be used as fuels. A microbial fuel
48 cell (MFC) is a device that converts chemical energy to electrical energy with the aid of
49 microorganisms (Pandey *et al.*, 2011). The fact that bacteria can oxidize the substrates
50 to produce electricity makes MFCs an ideal solution for wastewater treatment and
51 domestic energy production (Schwartz, 2007).

52 A MFC consists of anode and cathode separated by a cation specific membrane.
53 Microbes in the anode oxidize fuel here bacteria gain energy for metabolism by
54 transferring electrons from an electron donor, such as glucose or acetate to an electron
55 acceptor such as oxygen and the resulting electrons and protons are transferred to
56 cathode through the circuit and the membrane respectively. Electrons and protons are
57 consumed in the cathode, reducing oxidant usually oxygen. The electrons obtained from
58 this oxidation are transferred into anode chamber where they get departed through an
59 electrical circuit before entering into cathode region (Uma *et al.*, 2016). Since the
60 microbial cells are electrochemically inactive due to nonconductive cell surfaces
61 structure, mediators are employed to facilitate electron transfer from the microbial cells to
62 the anode in MFCs. Production of electrical energy using microorganisms through
63 microbial fuel cells (MFC) is one such renewable and sustainable technology that is
64 considered to be one of the most efficient (HaoYu *et al.*, 2007; Salgado, 2009) and
65 carbon neutral energy sources (Lovley, 2006).

66 Application of MFCs for wastewater treatment is very attractive due to energy recovery
67 from waste as well as reducing production of excess sludge, disposal of which is very
68 expensive. It is expected that this process would generate much less sludge than a

69 conventional activated sludge process, since the major part of energy available from the
70 oxidation of the organic contaminants is converted to electricity, and the remaining
71 energy is used for microbial growth.

72 This study aimed to 1) Isolate ,Identify and Procure of the effective micro-organisms
73 (*Escherichia coli*, *P.aeruginosa*) and 2) evaluation of the effective microbes generating
74 the maximum electricity from the given waste sample.

75 **2. MATERIALS AND METHOD:**

76 **2.1 WATER COLLECTION:**

77 Waste water was collected from three different places 1) Apartment waste 2) Lake water
78 3) Secondary treatment plant waste around 1000ml of each waste water surrounded by
79 Hebbal region of Bangalore.

80 **2.2 CONSTRUCTION OF THE MFC SET UP**

81 **PREPARATION OF SALT BRIDGE:** For the preparation of salt bridge, a water solution
82 containing concentrations of 3% sodium chloride and 1.6% agar was autoclaved and
83 poured into PVC pipe of length 10 cm and diameter 22.2mm covered at one end with
84 polythene. The set up was thereafter allowed to cool for nearly two hours inside High
85 efficiency Particulate Air Filter. The salt bridges were thus ready for use. **(Fig**

86 1).



87 Fig 1: Preparation of salt bridge

88 **DOUBLE CHAMBER SET UP**

89 The set up consisted of two reagent bottles made of plastic. The two were connected by
90 making an opening on one side of each bottle such that the salt bridge could be fitted
91 into them. The corners of the openings were sealed with M-seal to ensure that the
92 apparatus was made completely leak proof (Fig 2).

93 The 3 distinct anode chambers were filled with 500ml of apartment waste, lake water,
94 STP sample waste water and 5ml of *E. coli* culture was inoculated along with methylene
95 blue as electron mediator respectively. Same way to the other set of samples *P.*
96 *aeruginosa* was inoculated respectively. The cathode chambers were filled with 500ml of
97 phosphate buffer to those 3 sets of sample respectively of pH-more than 7. Clean
98 graphite electrodes (extracted from 1.5 Volt batteries) were coiled with copper wire and
99 introduced into the chambers. The set up was left 25 days under anaerobic conditions.

100 Cathode chambers would be equivalent of the oxygen sink at the end of the electron
101 transport chain, only now it would be external to the biological cell. The solution would be
102 an oxidizing agent that would pick up the electrons at the cathode. Potassium
103 ferricyanide can be used an oxidizing agent which can be added to the cathode to accept
104 electrons. It is very reactive with the graphite electrode. Ferricyanide has a fairly positive
105 potential compared to the organic matter in the anode and helps to drive the flow of
106 electrons. The set ups were kept for 30days with intermittent addition of inoculum of 3ml
107 to the respective bottles after every 6 days.



108

109 Fig 2: Construction of double chamber set up

110 2.3 DEVELOPMENT OF PURE MICROBIAL CULTURE:

- 111 ➤ Isolation and identification of *Escherichia coli* and *P.aeruginosa* by GRAM
112 STAINING.

113 ➤ Biochemical analysis by Catalase test, Indole test, MR test, Voges Proskauer test
114 and oxidase test.

115 **2.4 PRELIMINARY AND POST TREATMENT WATER ANALYSIS WAS CARRIED OUT BY:**

116 Determination of PH, Dissolved Oxygen, Biological Oxygen demand, Chemical
117 Oxygen Demand, Total Dissolved Solids, Calcium estimation, Phosphate estimation,
118 Sulphate estimation, Chloride estimation & Nitrate estimation.

119 **2.5. STATISTICAL ANALYSIS:**

120 The statistical significance was determined by one-way analysis of variance (ANOVA) to
121 determine if the data obtained was significantly varied from one another. Statistical
122 significance between different microorganisms (*E.coli* and *P.aeruginosa*) on 3 different
123 samples (Lake Water, Apartment waste, STP sample) was determined. P value of <0.05
124 was considered significant.

125

126 **2.6. ELECTRICAL MEASUREMENT**

127

128 The voltage readings were taken with the help of multimeter and it is converted into
129 current. Current was measured at a resistance (R) from the microbial fuel cell. I (mA) is
130 the current, and R (Ω) is the external resistance.

131 **4. RESULTS AND DISCUSSION**

132 The present research work entitled “**Generation of Bioelectricity and Simultaneous**
133 **Treatment of Waste Water Using Microbial Fuel Cells (MFC)**” was carried out at
134 Indian Academy Degree College, Centre for Research and PG studies, Bangalore.

135 For the performance MFC the two different microbes (*E.coli* and *P.aeruginosa*) were
136 isolated and inoculated which were identified by Gram’s staining and further biochemical
137 analysis confirms the purity of the respective microorganisms.

138 **4.1. ISOLATION AND IDENTIFICATION OF MICRO-ORGANISMS:** *Escherichia coli* was

139 isolated from the sewage sample using Eosin Methylene Blue Agar medium and
140 *P.aeruginosa* were isolated using *P.aeruginosa* isolation agar medium. The given micro-
141 organisms were identified by performing gram staining. Further the conformity of the
142 identified micro-organisms was tested at the biochemical level. On the basis of
143 morphological, cultural and bio-chemical characteristics, the isolated *E.coli* bacteria was
144 Gram negative in nature and showed positive response to Catalase, indole, MR test and
145 showed negative results for oxidase, VP test, Urease Test while the *P.aeruginosa*

146 bacteria showed positive response to Catalase, oxidase, tests and showed negative
147 response for indole, MR test, VP test.. These results are in consent with the work
148 reported by Priya Iyer¹ *et al.*, 2013;

149 **4.2. IMPACT OF MFC ON WASTE WATER QUALITY**

150 To check the efficiency of microbes certain physical analysis of three different samples
151 (Lake Water, Apartment waste, STP sample) were carried out before and after treatment
152 in MFC (with microbial inoculum).

153 The results analysed indicates that *E.coli* was well performed in treating all 3 samples by
154 the reduction in the level of BOD, TDS, COD, DO and CALCIUM. The lake water,
155 apartment waste and STP samples are all rich in great biomass sources (organic
156 matters) for MFCs, (Suzuki *et al* 1978; Oh and Logan 2005; Min *et al* 2005 and Zuo *et al*
157 2006). Liu and Logan (2004) suggested the vital role of the chamber setup (double)
158 which greatly influences the COD level of waste water. This setup removes up to 80% of
159 COD present in water sample. The present study showed that the role of
160 microorganisms (*E.coli*) is another added feature to convert rich organic matter into
161 generation of electricity by reducing concentration of COD BOD and DO. This is in
162 consent with the work done by Mercy *et al* 2012. Further there is an important role of
163 formation of biofilm in the MFC setup in degradation of waste. It was found that the
164 reduction of dissolved solids increased with the increase in biofilm concentration
165 (Hampannavar *et al* 2011). The study revealed the formation of increased biofilm in
166 *E.coli* cultures compared to *P.aeruginosa*. The calcium removal capacity with respect to
167 *E.coli* and *P.aeruginosa* in MFCs is not reported in the literature and further
168 investigations are necessary to confirm the mechanism behind the nitrogen removal and
169 its effect on overall power production in the MFC. But in case of other parameters like
170 NITRATE, CHLORIDE, SULPHATE, PHOSPHATE estimations *P.aeruginosa* performed
171 efficiently in reducing these amounts in samples. Since *P.aeruginosa* is a sulphide
172 oxidizing microorganism it has been reported that they are efficient anaerobic
173 microorganisms which involve in reducing the concentration of sulphate in the samples (
174 Dambe, T; Quentmeier *et al* 2005). Sulfate reduction in a MFC fed with carbohydrates
175 has been described previously (Rabaey,K *et al* 2004). According to Vanita Roshan Nimje
176 *et al* 2012, as a result of substrate oxidation, liberated electrons which gradually
177 decrease the level of nitrate but the role of microbes in reducing nitrates were not
178 described earlier. Even the phosphates and chlorides removal capacity with respect to
179 *E.coli* and *P.aeruginosa* in MFCs were also not reported in the literature and further

180 investigations are necessary to confirm the mechanism behind the nitrogen removal and
 181 its effect on overall power production in the MFC.

182

183 **Table 1: Comparative analysis of physical parameters of 3 samples before and**
 184 **after treatment by isolated micro organisms**

Physical parameters	Lake water			Apartment waste			STP waste		
	Before treatment	After treatment		Before treatment	After treatment		Before treatment	After treatment	
		<i>E.coli</i>	<i>P.aeruginosa</i>		<i>E.coli</i>	<i>P.aeruginosa</i>		<i>E.coli</i>	<i>P.aeruginosa</i>
PH	8.6	8.3	7.4	7.3	8.6	8.0	7.4	8.3	7.5
BOD	3.2mg/Lt	1.6mg/Lt	2.4mg/Lt	4.5mg/Lt	1.2mg/Lt	1.6mg/Lt	4.4mg/Lt	2mg/Lt	3.2mg/Lt
TDS	20.4g/Lt	0.92mg/Lt	1.08g/Lt	12.4g/Lt	1.84g/Lt	2.08g/Lt	10.8g/Lt	1.76g/Lt	1.88g/Lt
COD	136mg/Lt	64mg/Lt	92mg/Lt	596mg/Lt	244mg/Lt	316mg/Lt	364mg/Lt	152mg/Lt	184mg/Lt
DO	2mg/Lt	0.8mg/Lt	1.2mg/Lt	5.7mg/Lt	3.6mg/Lt	4.1mg/Lt	4.8mg/Lt	2.5mg/Lt	3.4mg/Lt
NITRATE	0.39mg/ml	0.15mg/ml	0.09mg/ml	0.36mg/ml	0.24mg/ml	0.20mg/ml	0.90mg/ml	0.60mg/ml	0.57mg/ml
CHLORIDE	192.4mg/Lt	72.4mg/Lt	49.9mg/Lt	417.3mg/Lt	139.9mg/Lt	124.9mg/Lt	342.3mg/Lt	112.4mg/Lt	92.49mg/Lt
SULPHATE	1.8mg/ml	0.5mg/ml	0.3mg/ml	1.9mg/ml	0.2mg/ml	0.0mg/ml	2.6mg/ml	0.4mg/ml	0.1mg/ml
PHOSPHATES	6.4mg/ml	4.2mg/ml	4.0mg/ml	7.4mg/ml	3.6mg/ml	2.6mg/ml	15mg/ml	6.8mg/ml	3.2mg/ml
CALCIUM	1.52mg/Lt	0.32mg/Lt	0.72mg/Lt	1.68mg/Lt	0.48mg/Lt	0.80mg/Lt	0.72mg/Lt	0.32mg/Lt	0.56mg/Lt

185

186 **4.3 PERFORMANCE OF MFC**

187 Once the apparatus was set up on combination of 3 different samples with respective
 188 micro-organisms i.e., Lake Water with *E.coli* and Lake water with *P.aeruginosa* followed

189 by Apartment waste with *E.coli* and Apartment waste with *P.aeruginosa* finally, STP
 190 sample with *E.coli* and STP sample with *P.aeruginosa*. The voltage in mili volts was
 191 checked with the help of multimeter in both control as well as in the samples with
 192 respective inoculum for about 30 days at a time point per day the readings were taken
 193 for two times (10.0AM and 4.0PM) under room temperature.

194 The voltage reading in the control samples was gradually found to decrease as the day's
 195 move on approximately till the day 12 the voltage was obtained, later the values were
 196 found to be as 0.00mV. Simultaneously the voltage values (mV) obtained in the samples
 197 treated with the respective micro-organisms (*E.coli* and *P.aeruginosa*) initially shown
 198 high values in the *E.coli* containing samples than *P.aeruginosa*.

199 The voltage values were obtained on intermittent addition of inoculum was carried out for
 200 efficient voltage result. 3ml of inoculum was added for every 6 days into the respective
 201 samples in the MFC set up.as per the values the voltage was found to decrease
 202 gradually but when the inoculum was reloaded into the medium the value was found to
 203 be increasing. The cycle was carried out for 30days as tabulated in **TABLE 1**.

204 The voltage values were converted into amps with the help of formula. The current
 205 produced is calculated as follows:

$$I = V/R$$

206
 207
 208 Where, I= Current in mA

209 V=Voltage in mV

210 R= Resistance in Ω

211 Ω-100Ω

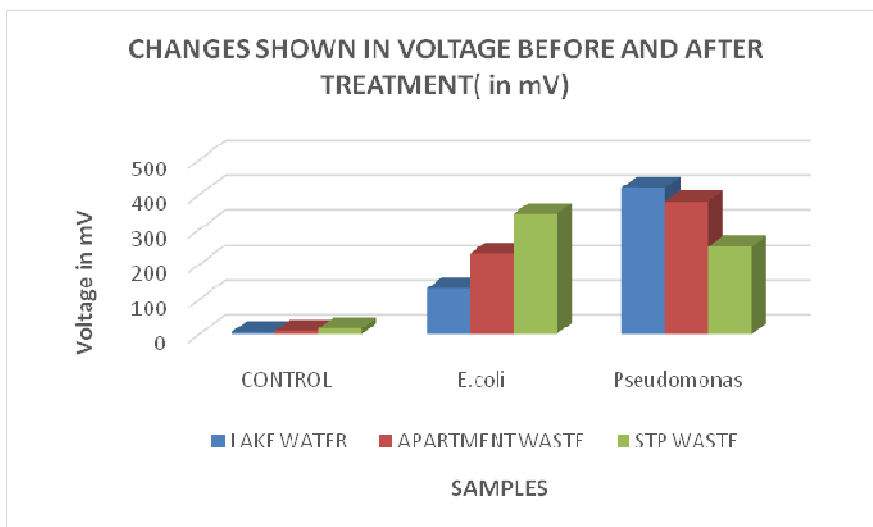
212

213 **Table 2: Changes shown in voltage (mV) before and after treatment.**

SAMPLES	CONTROL(mV)	<i>E.coli</i>(mV)	<i>P.aeruginosa</i>(mV)
LAKE WATER	5.88333333	131.18	419.80
APARTMENT WASTE	9.96666667	231.18	380.72
STP WASTE	16.58333333	344.17	251.82

214

215



216

217 **Graph 1: Effect of microbial inoculum in generation of voltage (mV) against**
 218 **samples compared with control.**

219

220 Followed by the voltage comparative results to see the efficiency of microbes between
 221 them in generating best voltage and as per the **TABLE 2** it was seen that in lake water
 222 *P.aeruginosa* was resulted in better voltage (**419.8mV**) when compared to that of *E.coli*
 223 (**131.18mV**) and same in case of apartment waste (*E.coli*-**231.18mV**, *P.aeruginosa*-
 224 **380.7mV**) but there was little difference was appeared in case of STP sample, *E.coli*
 225 were shown better voltage (**344.16mV**) than that of *P.aeruginosa* (**251.81mV**). Graphical
 226 representation was well established to compare the activity of inoculum in different
 227 sample (**GRAPH 1**). Similarly the control and treated values were obtained by the OHM's
 228 law thereby converting mV into mA. One way ANOVA was performed for average
 229 voltage (mV) readings to show the level of significance separately for three different
 230 samples and it was found that the Lake water, Apartment waste, STP sample there was
 a significant as obtained *P* value is less than 0.05.

231

232 Cumulative statistical analysis was performed to calculate the fold change and compare
 233 the *P* value obtained for voltage generated (mV) between treatment vs microbial
 inoculum (**TABLE 3**).

234

235

236

237 **Table 3:** Cumulative statistical analysis for voltage generated (mV) between treatment vs
 238 microbial inoculum.

SAMPLES	<i>E.coli</i> (mV)	<i>P.aeruginosa</i> (mV)	Fold change	P value
Lake water	131.1833	419.8	3.200102	2.58E-23
Apartment waste	231.1833	380.7167	1.646817	1.89E-07
STP sample	344.1667	251.8167	1.366735	0.005454

239

240 The control voltage values and treated voltage values were taken for 30 days
 241 respectively during that period of time intermittent (for every 6 days) addition of inoculum
 242 facilitated to show fluctuation in voltage readings up to maximum level and decline phase
 243 this is because when the microbial inoculum at exponential phase it shows high potential
 244 and it gradually decreases due to stationary and death phase S. Veer Raghavulu et al
 245 2011.

246 Upon addition of *E.coli* and *P.aeruginosa* in three different samples showed change in
 247 voltage reading (mV) as well as current reading (mA) among which in lake water and
 248 apartment waste, *P.aeruginosa* resulted in high voltage of **419.8mV and 380.7m V**
 249 respectively whereas *E.coli* resulted voltage in lake water and apartment waste was
 250 found to be **131.18mV and 231.18mV**. Among the pure cultures studied *P.aeruginosa* is
 251 electrochemically active in nature (Logan 2009) and *E. coli* is electrochemically inactive,
 252 while mixed consortia are comprised of both of them. Electrochemically active nature
 253 supports the pumping out of redox powers (H⁺ and e⁻) from the outer membrane of
 254 biocatalyst. According to S. Veer Raghavulu et al 2011, higher bio potential observed
 255 with *Pseudomonas aeruginosa* might be because of its electrochemical nature and the
 256 involvement of soluble shuttlers for the redox powers. In case of STP sample the voltage
 257 was seen high in the *E.coli* treated sample (**344.16mV**) than *P.aeruginosa* (**251.81mV**).
 258 The efficiency of *E.coli* in STP is due to the substrate level where the micro-organism
 259 oxidize organic compounds present in waste water, electrons are released yielding a
 260 steady source of electrical current.

261 The relation between volume of waste water, its characteristics, organic content,
 262 microbial load in anode chamber, the potential of electrode etc need to be standardised
 263 for improving the efficiency of the MFC.

264

265 **SUMMARY AND CONCLUSION**

266 Under present investigation, bioelectricity was successfully generated from Lake Water,
267 apartment and Secondary Treatment Plant waste water using Microbial Fuel Cell
268 Technology (MFC).

269 The physical parameters estimated and studied showed *E.coli* as an efficient source of
270 degradation for BOD, TDS, COD and DO in all the 3 samples. The efficiency of salt
271 degradation (NITRATE, CHLORIDE, SULPHATE AND PHOSPHATES) was shown more
272 by *P.aeruginosa* compared to *E.coli* in all the 3 samples.

273 According to the results obtained from this study, the electricity generation in Lake water
274 was facilitated more by the organism *P.aeruginosa* compared to *E.coli* which was proven
275 statistically by the ANOVA table, similarly in case of apartment waste *P.aeruginosa* was
276 giving best voltage and current but there was slight difference seen in case of STP
277 sample where *E.coli* gave the maximum voltage when compared with that of
278 *P.aeruginosa*. Since the ANOVA values obtained in the all three samples was less than
279 *P* value (0.05) hence the results were found to be significant.

280 MFC is a promising technology for bioelectricity generation and waste water treatment.
281 Recent research and development and analysis of literature review showed that higher
282 power densities can be obtained from improved MFC designs with the use of cost
283 effective materials. Hence, large scale MFC's can solve the future energy crisis
284 undoubtedly. More research and development is required for assessing suitability of
285 microorganisms for better efficiency for electricity production. The ultimate achievement
286 in MFCs will be when they can be used solely as a method of renewable energy
287 production.

288 So the present investigation led to a conclusion that the two suitable microorganisms
289 *E.coli* and *P.aeruginosa* proved to be equally efficient in degrading the waste depending
290 on the source with simultaneous production of electricity.

291 Further, a detail study for this upcoming technology for power generation need to be
292 studied and developed.

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