

Review Article

A review of experimental studies for available experimental evidence on the use of prosthetic material in diaphragmatic hiatal hernia repair

ABSTRACT

Aim: The benefits of prosthetic material in hiatal hernia repair have been well documented. However, the associated risks are substantial and they are related to the technique, but also the choice of material. Experimental data are invaluable to understand and evaluate the interaction of different meshes with the host tissue. The purpose of this article is to summarize the available experimental evidence in the repair of hiatal hernias with the use of prosthetic materials in animal models.

Methods: A review of the literature from January 1990 to December 2014 was carried out for articles presenting experimental data on hiatal hernia repair.

Results: After discarding non relevant articles, 35 articles were identified. A variety of synthetic and absorbable materials were studied. Review of the available studies showed that there is great variability between synthetic materials regarding tissue integration, shrinkage and adhesion formation, however they have greater mechanical strength when compared to however biological/absorbable materials, which have a tendency for better integration in host tissue. Biological adhesives seem to be an effective alternative method of mesh fixation.

Conclusions: Experimental data are essential in order to fully appreciate the process of repair of a hiatal hernia with a prosthetic material. The articles reviewed provide insight into the properties of different prosthetic materials. However, there were large variations in their quality and the methods used. Data from animal studies are an excellent way of evaluating the multitude of materials that have recently become available. Good quality, comparative animal studies are essential in an effort to further improve outcomes for patients who undergo hiatal hernia repair.

Keywords: hiatal, hernia, diaphragmatic, mesh, animal, experimental, review

9 **1. INTRODUCTION**

10
11
12 The introduction of laparoscopic techniques in hiatal hernia repair resulted in a significant increase
13 in the number of annually performed anti-reflux procedures in less than a decade [1]. There are now
14 randomized trials supporting the use of surgical management as a first-line treatment in selected patients
15 [2]. In some patient subgroups, however, such as the patients with a large paraesophageal hernia,
16 recurrence rates can reach 42% [3]. Recurrence usually occurs after disruption of the crural closure and
17 as the tissues being approximated are frequently attenuated and sutured under tension [4].

18 In an effort to overcome these limitations, selective mesh use has been reported since the 1970s.
19 In the first large series of patients published, Carlson et al were able to achieve excellent results with
20 polypropylene repair, without any clinical recurrences in long term follow up [5]. A number of clinical trials
21 have since established the efficacy of prosthetic mesh in preventing recurrence in the hiatus [6], however,
22 the emergence of relatively few, but in some cases devastating, complications such as mesh erosion,
23 highlight the need for further research [7].

24 As new materials are continuously being developed it is important for surgeons to make an
25 informed decision on which material to use. Animal studies are essential in evaluating the interaction
26 between the different prosthetic materials and the host tissue and their relative safety and efficacy in
27 hiatal hernia repair. We have performed a literature review in order to examine the contribution of the
28 available experimental evidence towards selecting the optimal prosthetic material and surgical technique
29 in mesh repair of hiatal diaphragmatic hernia.

30 **2. MATERIAL AND METHODS**

31
32
33
34 We searched for articles on hiatal diaphragmatic hernia repair meeting the criteria outlined below
35 and analyzed them for specific outcomes using the PRISMA guidelines.

36
37 **2.1 ELIGIBILITY CRITERIA**

38
39

40 1) Type of study: Experimental animal (in vivo) study of repair of hiatal/paraesophageal or congenital
41 diaphragmatic hernia using prosthetic material (mesh). Models of congenital diaphragmatic hernia
42 were included in this review, because, although the mesh was not placed in the hiatus in these
43 models, they can be considered orthotopic models, usually involving creation of a hernia by excision
44 of part of the left hemidiaphragm, mimicking conditions like those found in a giant paraesophageal
45 hernia (large defect, attenuation of muscular tissue).

46 2) Language: English

47 3) Publication year: 1990-2014

48

49 **2.2 LITERATURE SEARCH STRATEGY**

50

51 Studies were identified by searching the PubMed/Medline and Scopus databases. The following
52 key words were used as search strings: hiatal, diaphragmatic, mesh, animal, experimental.

53 Potentially relevant articles were identified by the title and abstract and full papers were obtained
54 and assessed in detail by two of the authors (M.S. and P.T., both senior surgeons) prior to their inclusion
55 in the review. The reference list for each article was also screened to identify further relevant publications.

56

57 **2.3 Study selection**

58

59 Eligibility assessment was performed independently by 2 reviewers. Disagreements between
60 reviewers were resolved by consensus.

61

62 **2.4 Data extraction**

63

64 Data collection and analysis were carried out independently by 2 researchers. Studies were
65 classified into two experimental model groups which investigated mesh repair of either hiatal or congenital
66 diaphragmatic hernia. Articles were reviewed for a number of variables examining their design (number
67 and type of animals, mesh implantation time, use of comparative/control group,

69 biomechanical/histopathological analysis) and the technique used (Mesh type and shape, fixation type,
70 surgical technique).

71 Study results were specifically assessed for findings relevant to controversial topics in hiatal
72 diaphragmatic hernia repair with prosthetic mesh (Table 1).

73

Table 1: Controversial topics in hiatal hernia repair with prosthetic mesh

1. Mesh shape	
2. Mesh type	a. Infection potential
	b. Handling characteristics
	c. Durability of repair
	d. Adhesion potential, tissue incorporation, fibrosis/stenosis/shrinkage potential
	f. Migration/erosion potential
3. Fixation method	
4. Sutured vs tension-free hiato-plasty	

74

75 **3. RESULTS**

76

77 **3.1 Literature search**

78 Our search strategy initially returned 924 studies which we evaluated based on title and
79 abstract and we selected 21 articles based on our inclusion criteria. The full text of these articles was
80 downloaded and another 9 studies were obtained from their reference lists. After excluding 2 articles
81 studying hiatal hernia repair in the context of fetal tissue engineering, 28 articles were assessed in detail
82 (Figure 1).

83

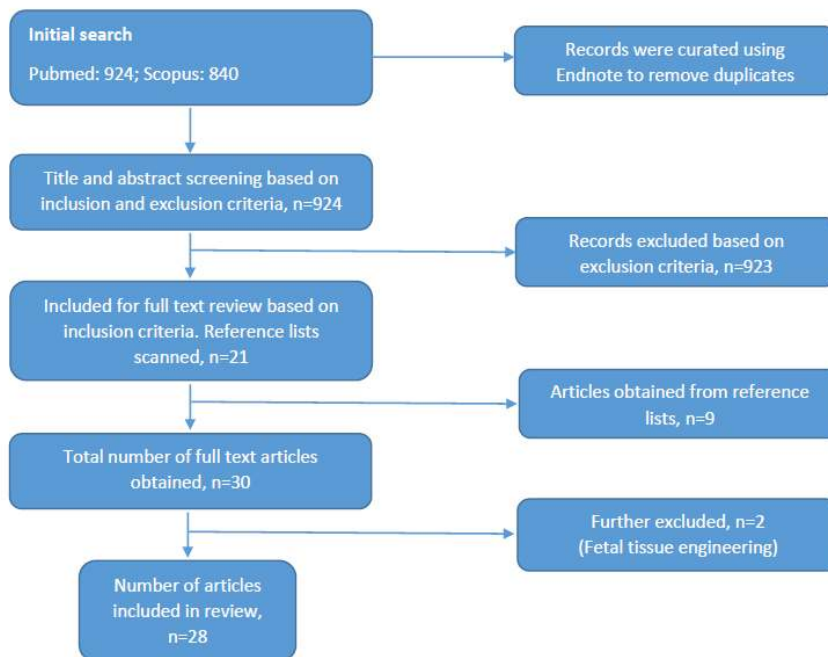
84 **3.2 Study design**

85 Large animals (swine, dogs) were used in most studies. The number of animals in each study was
86 small (6-36 animals). Implantation time ranged from 2 weeks to 12 months (Table 2). A majority of studies

87 included a comparison or control group and histopathological analysis, however only a few studies used
88 endoscopic or radiological assessment or biomechanical analysis.

89 **3.3 Mesh characteristics and surgical technique**

90 A variety of meshes were evaluated, including conventional (polypropylene,
91 polytetrafluoroethylene - PTFE) and newer (polypropylene/ polyglactin 910 - PP-PG, poly(lactic-co-
92 glycolic acid) - PLGA) synthetic materials, biologically derived materials such as bovine pericardium and
93 newer biologic meshes (Small intestinal submucosa - SIS, acellular dermal matrix - Alloderm). Most
94 authors used a rectangular piece of mesh, but circular and U-shaped meshes were also used. The
95 surgical technique used in most studies was mesh fixation in the hiatus using an open technique, with or
96 without excision of part of the left hemidiaphragm, while in two studies an endoscopic approach was
97 utilized: laparoscopic creation of a defect in the left hemidiaphragm and repair in one study and
98 thoracoscopic creation of a paraesophageal hernia and subsequent laparoscopic repair in another.
99 Finally, mesh fixation was achieved with sutures in most cases, while a few of the authors used biological
100 adhesives, such as fibrin glue and polyethylene glycol (Table 3).



101

102 **Figure 1: Flow diagram of literature search**

103 **3.4 Results of individual studies**

104 **3.4.1 Mesh shape**

105

106 Although circular, rectangular and U-shaped meshes were used, no study directly compared meshes of
107 different shapes. In a study of polypropylene meshes of a circular shape fixed by sutures in a rabbit
108 model, the meshes had usually moved from their implantation bed and had eroded into the esophagus
109 [16]. However, in another study a circular polypropylene mesh was fixed in place using fibrin glue in a
110 swine experimental model and in this case the authors reached conflicting results as they found the
111 meshes stayed in position and their inner edge had retracted evenly from the esophagus [14].

112

113 **3.4.2 Mesh type**

114

115

116 *3.4.2.1 Infection potential*

117

118 No study on mesh use in contaminated fields has been carried out.

119

120 *3.4.2.2 Handling characteristics*

121

122 The handling characteristics of each mesh i.e. the ease of its use in laparoscopic surgery was not
123 addressed in any study.

124

125 *3.4.2.3 Durability of repair*

126

127 Most of the studies discussed showed that the mesh repair remained successful during the observation
128 period of up to 12 months. SIS was shown to have equivalent strength to PTFE when applied on the
129 diaphragm [27, 32], although it was not as strong as polypropylene meshes [25, 26]. In another study
130 comparing two forms of SIS mesh in a dog model, the first comprised of 4-ply and the other from 8-ply,
131 the thicker version was shown to be stronger, while both showed more strength than native diaphragmatic
132 tissue [28]. Fascia lata was also shown to be equivalent to PTFE in mechanical strength [29].

133 3.4.2.4 Adhesion potential, tissue incorporation, fibrosis/stenosis/shrinkage potential

134

135 Polypropylene mesh consistently caused formation of strong adhesions [25, 26], which were less
136 pronounced with low-weight polypropylene [9]. Dualmesh showed less extensive adhesions than
137 polypropylene [17], while Surgisis showed less adhesions than PTFE in two studies [11, 32], but dense
138 adhesions comparable to polypropylene in another [25].

139 Mesh shrinkage was shown to be around 50-70% of original size for polypropylene [9, 14, 16], while the
140 percentage of shrinkage was more for the low-weight mesh [9]. When PTFE, polyester and polypropylene
141 were compared, PTFE showed considerably more shrinkage that reached 34.9% of its original size [8].

142 Bohm et al compared two composite synthetic polypropylene meshes (Ultrapro, Proceed) to
143 Surgisis in a rabbit model [25, 26]. Inflammatory reaction at the border of the mesh was more pronounced
144 with Proceed, followed by Ultrapro and Surgisis. On the other hand Surgisis and Ultrapro showed better
145 tissue regeneration compared to Proceed. Collagen maturation was slower for Surgisis compared to the
146 synthetic meshes. A composite polypropylene mesh was compared to a conventional polypropylene
147 mesh and the composite mesh showed better integration and reduced inflammatory response, which
148 could be associated with a lower risk of erosion and postsurgical dysphagia [16]. Histological examination
149 and cross-polarization microscopy showed differences in cell proliferation rate, apoptosis and collagen
150 I/III ratio, which were statistically significant and show better tissue integration for the composite mesh
151 [15, 16] Another study showed excellent integration, for a titanium-polypropylene mesh [13].

152 Polytetrafluoroethylene (ePTFE/ Dualmesh) was evaluated and caused the formation of minimal
153 adhesions except in segments of the mesh where folding exposed its superior surface. There were no
154 erosions or migration noted. Microscopic evaluation showed only an unstable capsule encompassing the
155 mesh underlining the importance of a stable fixation [17, 24, 33].

156 Biologically-derived materials were evaluated in several studies. The authors reported complete
157 mesh replacement by fibrovascular scar tissue with SIS mesh, with significant muscular regeneration,

158

159

Table 2: Design of experimental studies in hiatal diaphragmatic hernia repair

	Author	Pub year	Animal type	N	Implantation time	Comparative/control group	Biomechanical analysis	Histopathological analysis	Endoscopic/radiological assessment
<i>Hiatal hernia repair models</i>									
1	Muller-Stich [8]	2014	Swine	24	8 weeks	✓		✓	
2	Senft [9]	2014	Swine	24	8 weeks	✓			
3	Krpata [10]	2012	Swine	20	30 days				
4	Vereczkei [11]	2012	Dogs	3	1/3/6 months	✓		✓	
5	Jenkins [12]	2011	Swine	32	2 weeks				
6	Fortelny [13]	2010	Swine	7	4 weeks			✓	
7	Muller-Stich [14]	2008	Swine	9	6 weeks			✓	
8	Otto [15]	2008	Rabbits	20	3 months	✓		✓	
9	Jansen [16]	2007	Rabbits	20	3 months	✓	✓		✓
10	Smith [17]	2007	Swine	18	3/28 weeks	✓		✓	
11	Desai [18]	2006	Dogs	6	12 months			✓	✓
<i>Congenital diaphragmatic hernia repair models</i>									
12	Brouwer [19]	2013	Lambs	7	6 months	✓		✓	
13	Zhao [20]	2013	Rats	52	1, 2, 4, and 6 months	✓		✓	
14	Brouwer [21]	2013	Rats	36	12 weeks	✓		✓	
15	Brouwer [22]	2013	Rats	36	2/12 weeks	✓		✓	
16	Brouwer [23]	2013	Rats	25	2/4/8/12/24 weeks	✓		✓	
17	Gonzalez [24]	2011	Swine	20	6 months	✓		✓	
18	Bohm [25]	2010	Rabbits	33	4 months	✓		✓	
19	Bohm [26]	2010	Rabbits	33	4 months	✓	✓		
20	Urita [27]	2008	Rats	24	1-3 months				
21	Sandovalb [28]	2006	Dogs	11	6 months	✓	✓	✓	
22	Suzuki [29]	2002	Dogs	24	15/30 days				
23	Upadhyaya [30]	2001	rat	8	3 weeks	✓	✓	✓	
24	Steinau [31]	2000	pigs	24	3/6 months	✓	✓	✓	
25	Lantis II [32]	2000	Rabbits	32	6/12 weeks	✓	✓	✓	
26	Kimber [33]	2000	Lambs	12	1,3,6 months	✓	✓		
27	Dalla Vecchia[34]	1999	Rats	87	2 weeks - 4 months	✓		✓	
28	Lally [35]	1993	Rats	37	400 gr	✓		✓	

160

161 without any erosion in surrounding hollow viscera [18]. The 8-ply SIS mesh shows a slower rate of
162 degradation compared to the 4-ply, which can in turn lead to better integration into host tissue [28]. SIS
163 shows equivalent capillary ingrowth to Alloderm (acellular human cadaveric dermis), but a higher level of
164 thinning [34]. When compared to PTFE, SIS shows better integration [11], more collagen deposition and
165 skeletal muscle regeneration and neovascularization [24]. Finally, fascia lata showed superior integration
166 and capillary ingrowth to ePTFE [29] and, in a separate study, excellent tissue integration and
167 neovascularization, along with a mild to moderate inflammatory reaction [11].

168

169 *3.4.2.5 Migration/erosion potential*

170

171 In a study of polypropylene meshes of a circular shape fixed by sutures in a rabbit model, the meshes
172 had usually moved from their implantation bed and had eroded into the esophagus [16]. However, in
173 another study a circular polypropylene mesh was fixed in place using fibrin glue in a swine experimental
174 model and in this case the authors reached conflicting results as they found the meshes stayed in
175 position and their inner edge had retracted evenly from the esophagus [14].

176 The level of migration and the extent of foreign body reaction were higher when a conventional
177 polypropylene mesh was used compared to a composite one [15, 16]. The part of the mesh close to the
178 diaphragm showed less mechanical stability compared to the one close to the esophagus. In a
179 comparative study of PTFE and SIS in a pig model of congenital diaphragmatic hernia repair, the authors
180 were able to demonstrate PTFE has a poorer integration into host tissue compared to SIS and tends to
181 migrate and fold [24].

182

183 **3.4.3 Fixation method (sutures/tacks/glue)**

184

185 Biologically compatible adhesives like fibrin glue and polyethylene glycol were used with no evidence of
186 migration, no evidence of any adverse effect to the incorporation of the mesh and equivalent strength to
187 suture fixation. Krpata et al used an acellular porcine dermal matrix and compared fibrin sealant to
188 fixation with sutures [10]. Meshes fixed with fibrin glue showed no folding, while there was minimal folding

189 in the control group. Esophagograms did not exhibit any signs of strictures. The authors used a “peel” test
190 to compare the force needed to separate the mesh from the crura and found no difference between the
191 two techniques, whilst the introduction of glue between the crura and the mesh did not result in a
192 significantly different cellular response. Use of fibrin sealant resulted in a significant reduction in operative
193 time. Jenkins et al compared two biological adhesives and found both equally effective in mesh fixation
194 [12]. In conclusion data from 6 experimental studies show that both adhesives seem very promising as an
195 alternative, safe, faster fixation method in hiatal hernia repair.

196

197 **3.4.4 Sutured or tension free hiatoplasty**

198 In reviewing the available published studies we did not find any study comparing sutured to tension-free
199 hiatoplasty.

200

201 **4. DISCUSSION**

202

203

204

205 There are a number of controversial points regarding the best surgical technique in hiatal and
206 paraesophageal hernia surgery [36], the most controversial of which concerns the placement of mesh in
207 the oesophageal hiatus [37-39]. There are reports of a significant reduction in recurrence rates when
208 mesh is used in the surgical repair of hiatal hernia [40, 41]. On the other hand, the surgical community is
209 now conscious that there are important drawbacks in the form of mesh-related complications, reports of
210 which were scarce for two decades and have now begun to appear in the literature [7, 42, 43]. A mesh
211 placed in the diaphragm is subjected to the constant movements of breathing, which are likely to affect its
212 incorporation to the host tissue. Therefore, although there are multiple articles available studying mesh
213 use in animal models for a variety of indications, it is important to evaluate results from animal models of
214 hiatal diaphragmatic hernia repair.

215 Our literature review showed that there were large variations in the quality of experimental studies,
216 only a few of which incorporated histopathological, biomechanical, endoscopic and radiological
217 assessment. The number of animals was small and the implantation time was limited. The surgical
218 technique used in most cases is a disadvantage, since

219

Table 3: Mesh characteristics and surgical technique

Mesh type	Mesh shape	Surgical technique	Fixation method	
<i>Hiatal hernia repair models</i>				
1 [8]	PP/ PET/ PTFE	Circular	Open hiatoplasty and placement of patch in the oesophageal hiatus	Fibrin glue
2 [9]	heavyweight small-porous/heavyweight large-porous/lightweight large-porous PP	Circular	Open hiatoplasty and placement of patch in the oesophageal hiatus	Fibrin glue
3 [10]	acellular porcine dermal matrix	U shaped	Laparoscopic hiatal hernia repair	Sutures/Fibrin sealant
4 [11]	Pericardial and fascia lata patches	Rectangle	3x3 cm patches fixed on muscular part of diaphragm	Polypropylene 3/0
5 [12]	SIS	U shaped	Laparoscopic placement of patch in oesophageal hiatus	Fibrin glue/ polyethylene glycol
6 [13]	Titanium polypropylene mesh	Keyhole	Open placement of the patch without prior hiatoplasty	Fibrin glue
7 [14]	Heavy-weight polypropylene	Circular	Open hiatoplasty and placement of patch in the oesophageal hiatus	Fibrin glue
8 [15]	PP/ PP-polyglactone 25 composite	Circular	Open hiatoplasty and placement of patch in the oesophageal hiatus	Polypropylene 6/0
9 [16]	PP/ PP-polyglactone 25 composite	Circular	Open hiatoplasty and placement of patch in the oesophageal hiatus	Polypropylene 6/0
10 [17]	DualMesh	U shaped	Open transabdominal excision of left hemidiaphragm and open placement of patch in the oesophageal hiatus without prior hiatoplasty	Interrupted ePTFE
11 [18]	SIS	U shaped	Thoracoscopic creation of diaphragmatic hernia and subsequent laparoscopic repair, with hiatoplasty and placement of patch	Interrupted 2/0 polyester
<i>Congenital diaphragmatic hernia repair models</i>				
12 [19]	Collagen-Vicryl	Rectangle	Posterolateral 3x1.5 cm diaphragmatic defect	Running 4/0 prolene
13 [20]	poly(ϵ -caprolactone) and collagen type I	Rectangle	Excision of 70% of the left hemi-diaphragm (approximately 2-3 cm ²)	Interrupted 6/0 Prolene
14 [21]	Dual layered collagenous scaffolds	Rectangle	12 mm diameter right diaphragm defect	Interrupted 6/0 Prolene
15 [22]	Cross-linked collagenous scaffolds	Rectangle	12 mm diameter right diaphragm defect	Interrupted 6/0 Prolene/ interrupted 5/0 Vicryl
16 [23]	Cross-linked collagenous scaffolds	Rectangle	Excision of 1/3 of the right hemidiaphragm	Interrupted 6/0 Prolene
17 [24]	SIS, ePTFE	Rectangle	Excision of the left hemidiaphragm	Running 3/0 prolene
18 [25]	SIS, PP plus Polyglactone-25, and PP plus polydioxanone and cellulose plus Tachosil	Rectangle	A defect of 1cm in diameter was made into the lateral left diaphragm at the interface of tendon and muscle	Running 5/0 Prolene
19 [26]	SIS, PP plus Polyglactone-25, and PP plus polydioxanone and cellulose plus Tachosil	Rectangle	A defect of 1cm in diameter was made into the lateral left diaphragm at the interface of tendon and muscle	Running 5/0 Prolene
20 [27]	PLGA - collagen mesh	Rectangle	Open transabdominal left hemidiaphragm excision and repair	N/A
21 [28]	SIS	Rectangle	Open transabdominal left central hemidiaphragm excision and repair	N/A
22 [29]	Autologous fascia lata/ ePTFE	Rectangle	Left thoracotomy, left hemidiaphragm excision and repair	N/A
23 [30]	Integra	Rectangle	Open excision of left hemidiaphragm and patch repair	Interrupted 6/0 Vicryl
24 [31]	lyophilized dura/ transverse abdominal bovine pericardial serosa	Rectangle	Open excision of left hemidiaphragm and patch repair	Polypropylene 3-0
25 [32]	SIS/ PTFE	Rectangle	Open transabdominal left hemidiaphragm excision and repair	N/A
26 [33]	PTFE/ fluoropolymer-coated PET	Rectangle	Laparoscopic creation of 2x2 cm defect in left hemidiaphragm and repair	3-0 braided polyester
27 [34]	SIS/AlloDerm	Rectangle	Open transabdominal left central hemidiaphragm excision and repair	N/A
28 [35]	ePTFE/oxidized cellulose/polyglactin 910	Rectangle	Excision of the	Running 4-0 silk.

219

220 neither the creation of a hiatal hernia nor minimally invasive techniques were used, although similar
221 experimental models have been described as in the article by Desai et al, where a study incorporating the
222 creation of a diaphragmatic hernia and its subsequent repair using laparoscopy is presented, in a model
223 closely resembling the current clinical practice and enabling the surgeon to appreciate the handling
224 characteristics of each mesh [18]. Finally, due to the heterogeneity of the studies quantitative analysis of
225 the results was not possible.

226 We evaluated studies regarding specific topics and a number of these were not addressed at all
227 (infection potential, handling characteristics, sutured/ tension free hiatoplasty), while there was limited
228 data on the impact of mesh shape and the migration/erosion potential and durability of each mesh.

229 A test of the durability of the hiatoplasty should ideally compare biological/bioabsorbable
230 prostheses to materials like polypropylene or PTFE, the efficacy of which has been demonstrated in
231 randomized trials [40, 41]. Biological meshes made from small intestinal submucosa have been shown to
232 reduce recurrence rates in a randomized trial with short-term follow-up [44] and can also be used as a
233 control to evaluate newer biological materials. The ability of the mesh to prevent recurrence can be
234 investigated at autopsy or radiologically. However, new recurrences have been known to occur for a long
235 time after surgery. Indeed, long term observation of the patients in the previously mentioned trial showed
236 no benefit in recurrence rates with SIS mesh [45]. The practical limitations of observation time in animal
237 studies lead authors to perform biomechanical evaluation of mesh materials to evaluate the durability of
238 the repair. Results confirm the better results obtained clinically with polypropylene compared to biologics,
239 but are surprising since PTFE was weaker than expected [40].

240 Most of the authors focused on the potential of adhesion formation, biocompatibility and tissue
241 integration of each mesh and adequate experimental data on several materials is available. The safety
242 profile of each material i.e. its potential to adhere to and erode into viscera, to cause extended fibrosis
243 resulting in oesophageal stenosis, or to migrate from its position is the most pressing issue, since it is the
244 reason mesh-augmented hiatoplasty is not widely used in clinical practice. Polypropylene meshes
245 showed good integration, but also caused significant adhesions. Experimental studies will be very useful

Table 4: Summary of results

1. Mesh shape : no differences
2. Mesh type
 - a. Infection potential : no differences
 - b. Handling characteristics : no differences
 - c. Durability of repair : Polypropylene stronger than PTFE, biologics
 - d. Adhesion potential, tissue incorporation, fibrosis/stenosis/*shrinkage* potential:
 - More adhesions with synthetic meshes, especially polypropylene. Significant amount of shrinkage for PTFE, but also polypropylene.
 - Better integration for composite compared to conventional polypropylene, unstable integration for PTFE.
 - Better tissue integration and regeneration for SIS compared to synthetic materials.
 - f. Migration/erosion potential : PTFE has a poorer integration compared to SIS, while composite is better than conventional polypropylene,
3. Fixation method: Biologically compatible adhesives comparable to suture fixation

246

247 for the comparison of the new generation lightweight meshes, so as to evaluate their advantages
248 compared with standard polypropylene meshes. PTFE resulted in less adhesions, but poor integration
249 with the host tissue. SIS mesh showed an excellent safety profile in experimental studies. These results
250 are in accordance with those obtained from clinical trials including a prospective randomized trial [44] but
251 a marked fibrous response was observed in a previous comparative experimental study, published in
252 abstract form, where significant esophageal stenosis was shown [46]. This finding is significant since
253 esophageal stenosis have been reported in clinical series of patients operated on with SIS mesh [7].

254 The method used to fix the prosthesis to the crura presents a problem since laparoscopic suturing
255 is challenging and time consuming (and potentially risky for the inexperienced) and use of tacks, although
256 fast and effective, places large vessels and the heart at risk of serious injury with potentially catastrophic
257 results [47]. The stability of the mesh depends on both the fixation method but also the material itself and

258 the strength of its incorporation. T-peel testing is an elegant method of quantifying the strength of mesh
259 incorporation [10]. Directly observing the tendency of the mesh to migrate and cause adhesions is
260 tempting, however results must be interpreted with caution; failure of known complications to emerge in
261 these studies could be caused by the relatively short observation time (erosions occur up to nine years
262 after surgery) [7], but could also be interpreted to a lesser extent as proof of the importance of surgical
263 technique (i.e. to strengthen the argument that the reported complications are not inherent in the mesh
264 type but rather are a result of inadequate surgical technique). Indeed, there was a striking difference in
265 mesh migration of polypropylene mesh in the article by Fortelny et al compared to the study of Jansen et
266 al [13, 16]; the difference in the ratio of thickness between the mesh and the tissues of the two different
267 animal models was offered as an explanation.

268 **5. CONCLUSION**

269

270

271 The introduction of the new collagen-based biomaterials and the preliminary encouraging results
272 from their use raised great expectations for improved outcomes. The biomaterials from porcine small
273 intestinal submucosa and porcine or human acellular dermis are already widely used in clinical practice
274 (they were being used in 1/3 of all mesh-augmented hiatal diaphragmatic hernia repairs a few years ago
275 [48]) and experimental data are invaluable to further our understanding of their incorporation in host
276 tissue. Fascia lata has been used as a prosthetic material in the hiatus during the 1970s with mixed
277 results, showing efficacy but also some complications [49]. It is, however, in our opinion a very interesting
278 material because it is the only easily obtainable strong autologous patch and has been shown to possess
279 strength equal to PTFE and better incorporation on the diaphragm [28]. Although, the reviewed articles
280 study most of the types of meshes currently in clinical use, new biological and bioabsorbable materials
281 are being introduced in clinical practice without any available published experimental data [49, 50].

282 There is an ever growing need for experimental studies, which should also be well-designed in
283 order to also tackle ethical concerns with regards to animal sacrifice. Studies on mesh-augmented
284 hiatoplasty should include a laparoscopic animal model, biomechanical evaluation and histopathological
285 evaluation of no less than two different biomaterials at the very minimum. In the absence of good quality
286 clinical trials, which are invariably difficult to put together due to the relatively small number of patients,

287 good quality, comparative animal studies are essential in order to identify the mesh with the best
288 safety/efficacy profile, determine the optimal shape and fixation method and enable surgeons to make an
289 inform decision on the merits of using mesh in hiatal hernia repair.

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314
315
316
317
318
319
320
321
322
323
324
325
326
327
328
329
330
331
332
333
334
335
336
337
338
339
340
341
342
343

REFERENCES

1. Finks JF, Wei Y, Birkmeyer JD. The rise and fall of antireflux surgery in the United States. *Surg Endosc.* 2006;20(11): 1698-1701.
2. Mehta S, Bennett J, Mahon D, Rhodes M. Prospective trial of laparoscopic nissen fundoplication versus proton pump inhibitor therapy for gastroesophageal reflux disease: Seven-year follow-up. *J Gastrointest Surg.* 2006;10(9): 1312-6; discussion 1316-7.
3. Hashemi M, Peters JH, DeMeester TR, Huprich JE, Quek M, Hagen JA et al. Laparoscopic repair of large type III hiatal hernia: objective followup reveals high recurrence rate. *J Am Coll Surg.* 2000;190(5): 553-560; discussion 560-551.
4. Soper, N. J. and D. Dunnegan. Anatomic fundoplication failure after laparoscopic antireflux surgery. *Ann Surg.* 1999; 229(5): 669-676; discussion 676-667.
5. Carlson MA, Condon RE, Ludwig KA, Schulte WJ. Management of intrathoracic stomach with polypropylene mesh prosthesis reinforced transabdominal hiatus hernia repair. *J Am Coll Surg.* 1998; 187(3): 227-230.
6. Granderath FA, Carlson MA, Champion JK, Szold A, Basso N, Pointner R et al. Prosthetic closure of the esophageal hiatus in large hiatal hernia repair and laparoscopic antireflux surgery. *Surg Endosc.* 2006;20(3): 367-379.
7. Stadhuber RJ, Sherif AE, Mittal SK, Fitzgibbons RJ Jr, Michael Brunt L, Hunter JG et al. Mesh complications after prosthetic reinforcement of hiatal closure: a 28-case series. *Surg Endosc.* 2009;23(6): 1219-26.

- 344 8. Müller-Stich BP, Senft JD, Lasitschka F, Shevchenko M, Billeter AT, Bruckner T et al. Polypropylene,
345 polyester or polytetrafluoroethylene-is there an ideal material for mesh augmentation at the esophageal
346 hiatus? Results from an experimental study in a porcine model. *Hernia*. 2014;18(6):873-81.
347
- 348 9. Senft J, Gehrig T, Lasitschka F, Linke GR, Shevchenko M, Bruckner T et al. Influence of weight and
349 structure on biological behavior of polypropylene mesh prostheses placed at the esophageal hiatus. *J*
350 *Laparoendosc Adv Surg Tech A*. 2014;24(6):383-90.
351
- 352 10. Krpata DM, Blatnik JA, Harth KC, Phillips MS, Novitsky YW, Rosen MJ. Evaluation of fibrin sealant for
353 biologic mesh fixation at the hiatus in a porcine model. *Surg Endosc*. 2012;Nov;26(11):3120-6.
354
- 355 11. Vereczkei A, Varga G, Tornoczky T, Papp A, Horvath ÖP. A new experimental method for hiatal
356 reinforcement using connective tissue patch transfer. *Dis Esophagus*. 2012;Jul;25(5):465-9.
357
- 358 12. Jenkins ED, Lerdsirisopon S, Costello KP, Melman L, Greco SC, Frisella MM et al. Laparoscopic
359 fixation of biologic mesh at the hiatus with fibrin or polyethylene glycol sealant in a porcine model. *Surg*
360 *Endosc*. 2011;25(10):3405-13.
361
- 362 13. Fortelny RH, Petter-Puchner AH, Glaser KS, Keibl C, Gruber-Blum S, Ohlinger W et al. Fibrin sealant
363 (Tisseel) for hiatal mesh fixation in an experimental model in pigs. *J Surg Res*. 2010;162(1):68-74.
364
- 365 14. Müller-Stich BP, Mehrabi A, Kenngott HG, Fonouni H, Reiter MA, Kuttymorotov G et al. Is a circular
366 polypropylene mesh appropriate for application at the esophageal hiatus? Results from an experimental
367 study in a porcine model. *Surg Endosc*. 2009;23(6):1372-1378.
- 368 15. Otto J, Kämmer D, Jansen PL, Anurov M, Titkova S, Ottinger A et al. Different tissue reaction of
369 oesophagus and diaphragm after mesh hiatoplasty. Results of an animal study. *BMC Surg*, 2008;8: 7.
370

- 371 16. Jansen M, Otto J, Jansen PL, Anurov M, Titkova S, Willis S et al. Mesh migration into the esophageal
372 wall after mesh hiatoplasty: comparison of two alloplastic materials. *Surg Endosc.* 2007;21(12): 2298-
373 2303.
- 374
- 375 17. Smith GS, Hazebroek EJ, Eckstein R, Berry H, Smith WM, Isaacson JR et al. Evaluation of DualMesh
376 for repair of large hiatus hernia in a porcine model. *Surg Endosc.* 2008;22(7): 1625-1631.
- 377
- 378 18. Desai KM, Diaz S, Dorward IG, Winslow ER, La Regina MC, Halpin V et al. Histologic results 1 year
379 after bioprosthetic repair of paraesophageal hernia in a canine model. *Surg Endosc* 2006;20(11): 1693-
380 1697.
- 381
- 382 19. Brouwer KM, Daamen WF, Hoogenkamp HR, Geutjes PJ, de Blaauw I, Janssen-Kessels W et al.
383 Collagen-Vicryl scaffolds for reconstruction of the diaphragm in a large animal model. *J Biomed Mater*
384 *Res B Appl Biomater.* 2014 May;102(4):756-63.
- 385
- 386 20. Zhao W, Ju YM, Christ G, Atala A, Yoo JJ, Lee SJ. Diaphragmatic muscle reconstruction with an
387 aligned electrospun poly(ϵ -caprolactone)/collagen hybrid scaffold. *Biomaterials.* 2013 Nov;34(33):8235-
388 40.
- 389
- 390 21. Brouwer KM, Daamen WF, Reijnen D, Verstegen RH, Lammers G, Hafmans TG et al. Repair of
391 surgically created diaphragmatic defect in rat with use of a crosslinked porous collagen scaffold. *J Tissue*
392 *Eng Regen Med.* 2013 Jul;7(7):552-61
- 393
- 394 22. Brouwer KM, Daamen WF, van Lochem N, Reijnen D, Wijnen RM, van Kuppevelt TH. Construction
395 and in vivo evaluation of a dual layered collagenous scaffold with a radial pore structure for repair of the
396 diaphragm. *Acta Biomater.* 2013 Jun;9(6):6844-51
- 397

398 23. Brouwer KM, Wijnen RM, Reijnen D, Hafmans TG, Daamen WF, van Kuppevelt TH. Heparinized
399 collagen scaffolds with and without growth factors for the repair of diaphragmatic hernia: construction and
400 in vivo evaluation. *Organogenesis*. 2013 Jul-Sep;9(3):161-7
401

402 24. Gonzalez R, Hill SJ, Mattar SG, Lin E, Ramshaw BJ, Smith CD. Absorbable versus nonabsorbable
403 mesh repair of congenital diaphragmatic hernias in a growing animal model. *J Laparoendosc Adv Surg*
404 *Tech A*. 2011;21(5):449-54.
405

406 25. Böhm G, Binnebösel M, Krähling E, Schumpelick V, Steinau G, Stanzel S et al. Influence of the
407 elasticity module of synthetic and natural polymeric tissue substitutes on the mobility of the diaphragm
408 and healing process in a rabbit model. *J Biomater Appl*. 2011;25(8):771-93.
409

410 26. Böhm G, Steinau G, Krähling E, Schumpelick V, Hermanns-Sachweh B, Stanzel S et al. Is
411 biocompatibility affected by constant shear stress?--comparison of three commercially available meshes
412 in a rabbit model. *J Biomater Appl*. 2011;25(7):721-41
413
414

415 27. Urita Y, Komuro H, Chen G, Shinya M, Saihara R, Kaneko M. Evaluation of diaphragmatic hernia
416 repair using PLGA mesh-collagen sponge hybrid scaffold: an experimental study in a rat model. *Pediatr*
417 *Surg Int*. 2006;24(9): 1041-1045.
418

419 28. Sandoval JA, Lou D, Engum SA, Fisher LM, Bouchard CM, Davis MM et al. The whole truth:
420 comparative analysis of diaphragmatic hernia repair using 4-ply vs 8-ply small intestinal submucosa in a
421 growing animal model. *J Pediatr Surg*. 2006;41(3): 518-523.
422

423 29 Suzuki K, Takahashi T, Itou Y, Asai K, Shimota H, Kazui T. Reconstruction of diaphragm using
424 autologous fascia lata: an experimental study in dogs. *Ann Thorac Surg*. 2002;74(1): 209-212.
425

- 426 30. Upadhyaya M, Orford JE, Smith N, Barker A, Gollow I. Incorporation of Integra in tissue defects: a
427 pilot study in the rat model. *Pediatr Surg Int.* 2007;23(7):669-73.
428
429
- 430 31. Steinau G, Dreuw B, Schleef J, Lawong G, Schumpelick V. Short-term absorbable material for
431 diaphragmatic replacement. *Pediatr Surg Int.* 2000;16(1-2):19-22.
432
- 433 32. Lantis JC 2nd, Gallivan EK, Hekier R, Connolly R, Schwaitzberg SD, Crombleholme T. A comparison
434 of collagen and PTFE patch repair in a rabbit model of congenital diaphragmatic hernia. *J Invest Surg.*
435 2000;13(6): 319-325.
436
- 437 33. Kimber CP, Dunkley MP, Haddock G, Robertson L, Carey FA, Cuschieri A. Patch incorporation in
438 diaphragmatic hernia. *J Pediatr Surg.* 2000;35(1):120-3.
439
- 440 34. Dalla Vecchia, Engum S, Kogon B, Jensen E, Davis M, Grosfeld J. Evaluation of small intestine
441 submucosa and acellular dermis as diaphragmatic prostheses. *J Pediatr Surg.* 1999;34(1): 167-171.
442
- 443 35. Lally KP, Cheu HW, Vazquez WD. Prosthetic diaphragm reconstruction in the growing animal. *J*
444 *Pediatr Surg.* 1993;28(1):45-7.
445
- 446 36. Neufeld, M., Graham A. Levels of evidence available for techniques in antireflux surgery. *Dis*
447 *Esophagus.* 2007;20(2): 161-7.
448
- 449 37. Targarona EM, Bendahan G, Balague C, Garriga J, Trias M. Mesh in the hiatus: a controversial issue.
450 *Arch Surg.* 2004;139(12): 1286-96; discussion 1296.
451
- 452 38. Smith, G. Mesh repairs in hiatal surgery. The case for mesh repairs in hiatal surgery. *Ann R Coll Surg*
453 *Engl.* 2007;89(5): 481-3.
454

455 39. Kelty, C. J., Falk, G. L. Mesh repairs in hiatal surgery. The case against mesh repairs in hiatal
456 surgery. *Ann R Coll Surg Engl.* 2007;89(5): 479-81.

457

458 40. Frantzides CT, Madan AK, Carlson MA, Stavropoulos GP. A prospective, randomized trial of
459 laparoscopic polytetrafluoroethylene (PTFE) patch repair vs simple cruroplasty for large hiatal hernia.
460 *Arch Surg.* 2002;137(6): 649-52.

461

462 41. Antoniou SA, Antoniou GA, Koch OO, Pointner R, Granderath FA. Lower recurrence rates after mesh-
463 reinforced versus simple hiatal hernia repair: a meta-analysis of randomized trials. *Surg Laparosc Endosc*
464 *Percutan Tech.* 2012;22(6):498-502.

465

466 42. Carlson, M. A., Frantzides, C. T. Complications and results of primary minimally invasive antireflux
467 procedures: a review of 10,735 reported cases. *J Am Coll Surg.* 2001;193(4): 428-39.

468

469 43. Tatum RP, Shalhub S, Oelschlager BK, Pellegrini CA. Complications of PTFE mesh at the
470 diaphragmatic hiatus. *J Gastrointest Surg.* 2008;12(5): 953-7.

471

472 44. Oelschlager BK, Pellegrini CA, Hunter J, Soper N, Brunt M, Sheppard B et al. Biologic prosthesis
473 reduces recurrence after laparoscopic paraesophageal hernia repair: a multicenter, prospective,
474 randomized trial. *Ann Surg.* 2006;244(4):481-90.

475

476 45. Oelschlager BK, Pellegrini CA, Hunter JG, Brunt ML, Soper NJ, Sheppard BC et al. Biologic
477 prosthesis to prevent recurrence after laparoscopic paraesophageal hernia repair: long-term follow-up
478 from a multicenter, prospective, randomized trial. *J Am Coll Surg.* 2011;213(4):461-8.

479

480 46. Halpin V, Meyers BF, Luttmann D. Laparoscopic paraesophageal hiatal hernia repair using
481 prosthetics in a canine model. *Surg Endosc.* 2002;16(Suppl 1): S977

482

- 483 47. Frantzides CT, Welle SN. Cardiac tamponade as a life-threatening complication in hernia repair.
484 Surgery. 2012 Jul;152(1):133-5.
485
- 486 48. Brain, R. H., Maynard, J. Fascia lata graft repair of esophageal hiatal hernia. Am J Surg.
487 1968;115(4): 488-501.
488
- 489 49. Zehetner J, Lipham JC, Ayazi S, Oezcelik A, Abate E, Chen W et al. A simplified technique for
490 intrathoracic stomach repair: laparoscopic fundoplication with Vicryl mesh and BioGlue crural
491 reinforcement. Surg Endosc. 24(3): 675-9.
492
- 493 50. Powell BS, Wandrey D, Voeller GR. A technique for placement of a bioabsorbable prosthesis with
494 fibrin glue fixation for reinforcement of the crural closure during hiatal hernia repair. Hernia.
495 2013;17(1):81-4