

3
4 **Effect of Nitrogen Rates on Growth and Quality of**
5 **Water Spinach (*Ipomea aquatica*)**
6

7 **ABSTRACT**
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Aims: The study was conducted to investigate the impact of nitrogen fertilization on growth, leaf gas exchange and bio-metabolite accumulation in *Ipomea aquatica*.

Treatment and experimental design: *Ipomea aquatica* plants were exposed to four different rates of nitrogen (0, 30, 60 and 90 N kg/ha) using Urea (46% N) as a nitrogen source. The experiment was laid out in Complete Randomize Design (CRD).

Place and Duration of Study: Department of Biology, Faculty of Science, Universiti Putra Malaysia between September to November 2016.

Methodology: Four nitrogen rates were applied (0, 30, 60 and 90 N Kg/ha) using Urea as a nitrogen source. The growth data collections were conducted once a week after the application of the treatments for the plant growth parameters. The total chlorophyll content in the leaves was measured using a Soil Plant Analytical Device (SPAD-502) chlorophyll meter. The leaf gas exchange was determined using a LI-6400XT portable photosynthesis system. Total phenolics and flavonoid were determined using Folin-Ciocalteu reagent.

Results: It was found that the highest measurements of growth parameters namely plant height, leaf numbers, branches numbers, total biomass and chlorophyll content were observed at 90 kg N/ha and the lowest at 0 kg N/ha. As for the leaf gas exchange, the positive effect of nitrogen fertilization on kangkung was shown by the increased in photosynthesis rate (A) and stomatal conductance (gs) where the highest measurement recorded at 90 kg N/ha, and the lowest at 0 kg N/ha. However, the water use efficiency (WUE) decreased as the nitrogen rates increased. At lower rates of nitrogen fertilization (30 kg N/ha) produced the highest production of secondary metabolites, where the total phenolics and flavonoids production were enhanced compared to other nitrogen treatments.

Conclusion: In conclusion, as the nitrogen rates increased, the growth and leaf gas exchange properties was enhanced however the production of secondary metabolites was decreased in *I. aquatica*

9
10 *Keywords:* [Nitrogen, *Ipomea aquatica*, growth, leaf gas exchange, biometabolites production]

11
12 **1. INTRODUCTION**
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14 In Malaysia, agricultural sector contributed about 8.5% to Gross Domestic Products (GDP). About
15 39% of the contributions originated from the production of food crops, fruits, and vegetables. It is
16 estimated that about 44, 000 hectares of the total area in Malaysia were used for vegetable cultivation
17 [1]. According to Department of Agriculture Malaysia in 2011, *Ipomea Aquatica* is one out of ten types
18 of vegetables that occupied the largest area for vegetable production. This plant is among the most
19 consumed vegetable in Asia. This is because of its low price compared to other types of vegetable.
20 Kangkung air or it's scientific name, *Ipomea aquatica*, is a widely known leafy vegetable, especially in
21 the Asian country. The plant is also commonly known by different local names, such as water spinach,
22 swamp cabbage, or water convolvulus. From its scientific classification, kangkung has been classified
23 under the family Convolvulaceae [2]. According to [3], Convolvulaceae family consists of primarily 1650
24 of tropical species. Moreover, the genus of kangkung which is *Ipomea* has about 500 to 600 different
25 species and it has been the most number of containing species in Convolvulaceae family [4]. This
26 species of the family can nicely be grown at almost anywhere at the higher or lower altitudes. *Ipomea*
27 *aquatica* is one of the species that is cultivated on the higher land. Besides easy to be grown,
28 Kangkung cultivation is favored due to its quick maturity period and it does not take long time to
29 mature and harvest. It can easily adapt towards it's growing environment and usually unsusceptible to
30 disease. Almost all parts of kangkung plant are edible [3].
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32 According to Susila et al. [5], nitrogen is the primary nutrient that involved in producing a high yield of
33 vegetables. Nitrogen is one of the macro-nutrients that is very crucial especially for a plant to have a
34 proper growth and development [6] such as that required in constructing the matter of the plant cell
35 and tissue [7]. The amount of nitrogen in the soil could be insufficient for the plant to grow. Therefore,
36 the source of nitrogen for plant especially in agriculture field is often found in the form of a fertilizer.
37 Both organic and inorganic nitrogen fertilizer is widely used in agriculture especially in cultivating
38 green crops to keep the source of nutrients for the plant being for supplied [6]. Practically, an
39 appropriate and suitable amount of nitrogen to be given to plant will affect its crop yield. Nitrogen is
40 also very important especially to promote the growth of the plant leaf [8]. Nitrogen is a crucial element
41 not only to promote the growth and plant development, also increase yield and quality in vegetable
42 crops. Increasing level of nitrogen resulting in a number of leaves, leaf length and plant body [8].
43 Nitrogen also enhancing the size of fruits and vegetables where at an optimum application of N will
44 result in a better size. The metabolic process which stimulated by N by enhance the vegetative and
45 also the reproductive growth in the plant. Besides, high plant biomass can be obtained when there is
46 high N accumulated in a shoot, along with the increasing of root growth in a plant if there is sufficient
47 amount of N supply [9]. However, the lack of N in a plant would cause the reduced in plant
48 development and eventually will lower the crop yield. Plants can take up nitrogen (N) either as
49 inorganic ions (NH_4^+ or NO_3^-), or as organic N. In leafy vegetables, high uptake of NO_3^- can cause
50 serious health problems to the consumers [7]. Nitrate has been attributed to have negative effects to
51 human health. Toxicity of nitrate to human can be manifested by headaches, syncope, vertigo and
52 discoloration that manifest in fingers or lips [6].

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54 Nitrogen has been proven to have a strong relationship with photosynthesis process in the plant.
55 Increasing N level leads to higher N content in leaves. N also enhances the leaf chlorophyll and CO_2
56 assimilation which increase in the Rubisco activity [10]. Therefore, increase in the rate of
57 photosynthesis is the most vital biochemical process in plants [11]. According to [12,13], rate of
58 photosynthesis (A) depends on the growth and development of the plant's leaf. The leaf development
59 includes the increase in leaf area, leaf thickness, the surface volume of mesophyll cells, and leaf
60 chloroplast. The photosynthesis rate will be increased as the leaf development also increased [14].
61 Nitrogen is an element that has a significance role in photosynthesis which involves in the opening of
62 the stomata. The stomatal vent will decrease following the nitrogen deficit which then will decrease
63 the transpiration rate [15].

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65 Secondary metabolites such as phenolic compounds in plants are usually associated with survival of
66 the plant and health benefits for those who consume the plant. Low nitrogen level in the plant has
67 been reported to have more secondary metabolites compare to plant that has high N level [16].
68 Application of more N level resulting in a decrease of phenolic concentrations based on
69 carbon/nutrients balance (CNB) hypothesis [17]. Flavanoids, also a secondary metabolite, are widely
70 distributed with different functions in plants. The biological functions of flavonoids include defense
71 against UV-B radiation, pathogen infection, nodulation and pollen fertility [18]. A study was done by [8]
72 on leaf mustard where the total phenolics concentration was observed to be decreased as the level of
73 N increased. It is well known that nitrogen application can directly affect the morphological growth and
74 yield of this plant, however, little work has been carried out to look on the impact of nitrogen of the leaf
75 gas exchange properties and previous work have not comprehensively considered the production of
76 secondary metabolites of *I. aquatica* under nitrogen fertilization. The main aim of the research was to
77 investigate the effect of nitrogen fertilization on the growth, leaf gas exchange and production of
78 secondary metabolites of *I. aquatica* and to determine the best nitrogen rates for growth and
79 development of *I. aquatica*. This research will provide the important information for vegetable growers
80 that involved in the cultivation of vegetables in Malaysia.

81 82 **2. MATERIAL AND METHODS**

83 84 **2.1. Plant material and maintenance**

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86 The experiment was conducted at the Department of Biology, Universiti Putra Malaysia, Serdang
87 (UPM), Selangor The seeds were pre-germinated in the nursery for two weeks after which they were
88 transplanted into the polybags filled with a mixture of topsoil, organic matter and sand in the ratio of
89 3:2:1 respectively. All the plants were irrigated using overhead mist irrigation given four times a day or
90 when necessary. Each irrigation session lasted for 7 min. The nitrogen sources used was single
91 fertilizer Urea (46% N). The polybags were arranged in Completely Randomized Design (CRD) with

92 five replications. There were four nitrogen rates were applied (0, 30, 60 and 90 Kg N/ha). The
93 fertilization with nitrogen levels were split into three applications, given at 5, 15 and 25 days after
94 treatments and each phase was about 33.3% of total nitrogen fertilizer. The growth data collections
95 were conducted once a week for four weeks after the application of the treatments. While the
96 destructive analysis and leaf gas exchange of the experiment was conducted at the end of the
97 experiment.

98 99 **2.2. Plant height, number of leaves and branches**

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101 Plant height, was measured from the ground level to the tip of the highest growing point using
102 measuring tape. The leaf and branches number were counted manually per plant and the mean
103 recorded

104 105 **2.3 Plant total dry weight measurement**

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107 The plants were first uprooted carefully and were washed with tap water. After that, the shoot and the
108 root parts were separated. All the plants were dried in an oven for 48 hours at temperature of 60°C
109 until constant weight reached. Then, the plant total dry weight was measured by using electronic
110 digital scale.

111 112 **2.4 Total Chlorophyll content**

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114 SPAD-502 chlorophyll meter was used to measure the total chlorophyll content of the leaves. Three
115 readings were taken at three spot on a leaf of each plant and the average readings were recorded.
116 Time interval between 9.00 a.m and 12.00 p.m was used to measure the chlorophyll content.

117 118 **2.5 Leaf gas exchange measurement**

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120 The leaf gas exchange measurement was obtained after week 4 the treatment was given. The result
121 then was obtained by using the Portable Photosynthesis System machine (LICOR 6400 XT). The
122 IRGA was firstly warm up for at least 30 minutes before the leaf gas exchange was collected with
123 Zero IRGA mode. The optimal condition was set to 400 $\mu\text{mol mol}^{-1}$ carbon dioxide (CO_2), 30 °C
124 cuvette temperature, 60% relative humidity with air flow rate set at 500 $\text{cm}^3 \text{min}^{-1}$, and 800 $\mu\text{molm}^{-2}\text{s}^{-1}$
125 of cuvette condition of photosynthetic photon flux density (PPFD). The time for the measurement were
126 done at the morning of a day. The measurement of photosynthesis rate was taken from the first
127 kangkung leaves starting from the plant apex. The data then were recorded and stored in a console of
128 the system and analyse with Photosyn Assistant Software. The photosynthesis (A), transpiration rate
129 (E), stomata conductance (gs) and water use efficiency (WUE) data was recorded during the
130 measurement.

131 132 **2.7 Total phenolics and flavonoids quantification**

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134 The methods used for extraction and quantification of total phenolics and flavonoids contents followed
135 that described in Ibrahim et al. [19]. A fixed amount of ground tissue samples (0.1 g) was extracted
136 with 80% ethanol (10 mL) on an orbital shaker for 120 min at 50 °C. The mixture was subsequently
137 filtered (Whatman™ No.1), and the filtrate was used for the quantification of total phenolics and total
138 flavonoids. Folin–Ciocalteu reagent (diluted 10-fold) was used to determine total phenolics content of
139 the leaf samples. The sample extract at 200 μL was mixed with Folin–Ciocalteu reagent (1.5 mL) and
140 allowed to stand at 22 °C for 5 min before adding NaNO_3 solution (1.5 mL, 60 g L^{-1}). After two hours
141 at 22 °C, absorbance was measured at 725 nm. The results were expressed as mg g^{-1} gallic acid
142 equivalent (mg GAE g^{-1} dry sample). For total flavonoids determination, samples (1 mL) were mixed
143 with NaNO_3 (0.3 mL) in a test tube covered with aluminium foil, and left for 5 min. Then 10% AlCl_3 (0.3
144 mL) was added followed by addition of 1 M NaOH (2 mL). The absorbance was measured at 510 nm
145 using a spectrophotometer with rutin as a standard (results expressed as mg/g rutin dry sample).

146 147 **2.8 Statistical analysis**

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149 Data were analysed using the analysis of variance procedure in SAS version 17. Means separation
150 between treatments was performed using Duncan multiple range test and the standard error of

151 differences between means was calculated with the assumption that data were normally distributed
152 and equally replicated.

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155 3. RESULTS AND DISCUSSION

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157 3.1 Plant height

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159 Figure 1 shows the plant height of *I. aquatica* as influenced by differing nitrogen treatments. The plant
160 height of kangkung was mostly affected by different rates of nitrogen treatment in all weeks of
161 measurement ($P \leq 0.05$). In view of the result obtained, as nitrogen levels increased from 0 to 90 kg
162 N/ha the plant height was enhanced in all weeks of measurement. At four weeks after treatment (4
163 WAT), plant at 0 kg/ha have the average height of 31.02 cm compared to 32.17 cm by 30 kg/ha,
164 35.61 cm by 60 kg/ha and 37.24 cm in 90 kg/ha. Clearly, as expected, applying higher rates of
165 nitrogen levels would enhance the plant height of *I. aquatica*. The positive effects on plant height
166 caused by the increase of nitrogen rates application may be due to the natural role of nitrogen on
167 vegetative growth performance of plants [6]. The increase in plant height under nitrogen fertilization
168 might be due to well-developed primary growth under high nitrogen fertilization that resulted in taller
169 plant [20]. Besides that, increase in plant height might be associated with the increased of number
170 and length of the internodes by nitrogen [21]. The result obtained agreed with the previous work
171 carried out by [4] and [6] where the increment of nitrogen fertilization rates applied to *I. aquatica*
172 significantly increased the plant height at end of the harvesting period.

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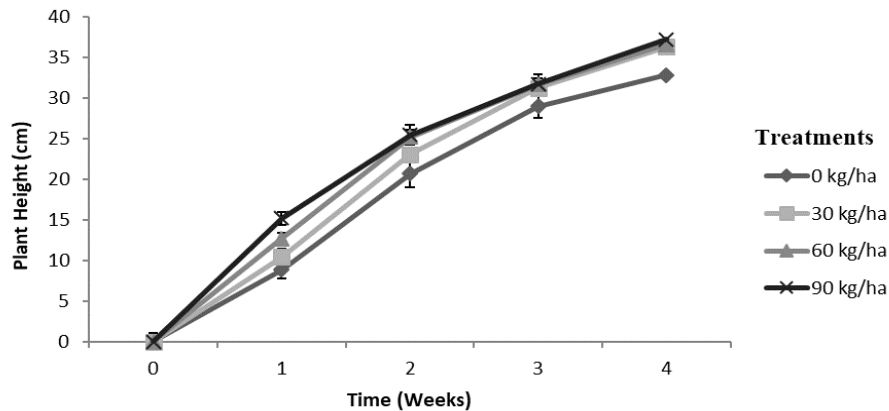
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189 **Fig 1. The impact of different nitrogen rates on the height of *Ipomea aquatica*. N =10. Bars**
190 **represent standard error of differences between means (SEM).**

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192 3.2 Leaves numbers

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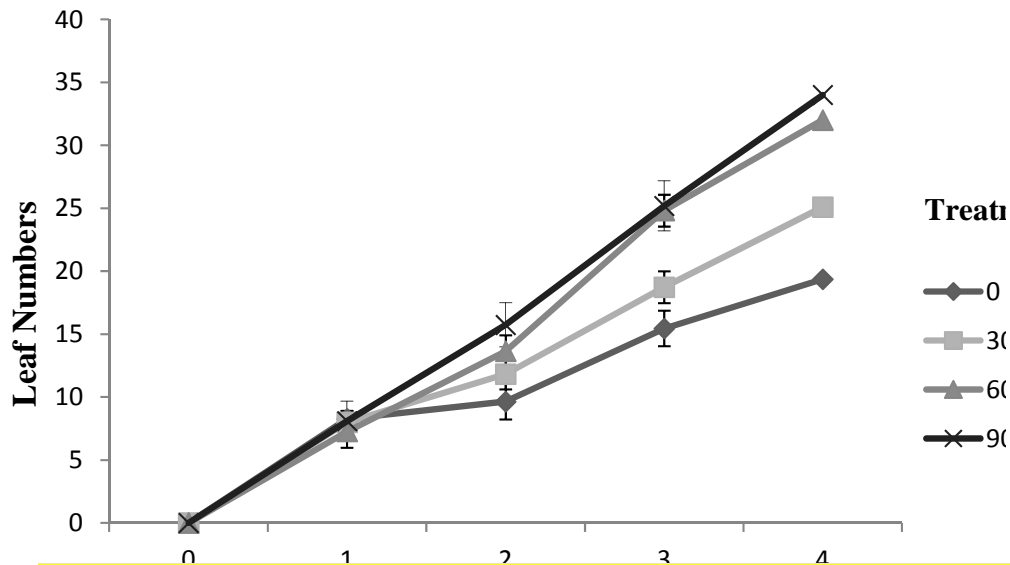
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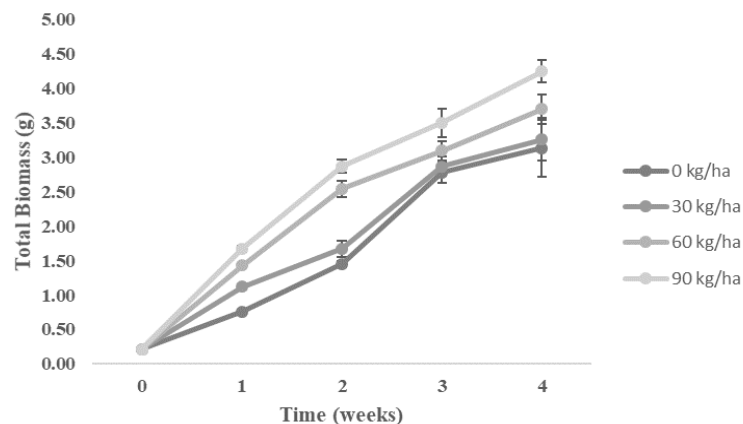
194 The variation of leaf numbers with different nitrogen fertilization is in *I. aquatica* is depicted in Figure 2.
195 Generally, leaf number of *I. aquatica* was found to be influenced by the different rates of nitrogen
196 treatments (0, 30, 60 and 90 kg/ha; $P \leq 0.05$). Based on Figure 2, it shows that there were significant
197 effects of nitrogen fertilization rates on the number of leaves in every week of measurements. Overall
198 at 90 N kg/ha as the highest treatments of nitrogen applied, lead to the drastic production in the
199 number of leaves from 1 to 4 WAT. An increase in number of leaf age in plants indicates better plant
200 growth and development. Eventually, the plant production also will increase. Similar trends were
201 observed in [6] and [20] where they found that as the rate of nitrogen increases the *I. aquatica* leaf
202 numbers were also enhances. The increase in leaf number in *I. aquatica* might be due to increase in
203 internodes number with the high application of nitrogen [21]. The high application of nitrogen usually
204 would reduce the apical dominance and stimulated the development of lateral buds that eventually
205 increase the production of plant leaf and simultaneously enhanced the leaf numbers [22].



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209 **Fig 2. The impact of different nitrogen rates on the leaves numbers of *Ipomea aquatica*. N =10.**
210 **Bars represent standard error of differences between means (SEM).**
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212 3.3 Plant Total dry weight

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214 Nitrogen application significantly influenced on the total plant dry weight of *I. aquatica* plant as shown
215 in Figure 3. The graph pattern shows increased in production in total biomass with the higher
216 application of nitrogen fertilization rates. At end of the treatments, It was observed that the highest
217 total biomass of kangkung was obtained in 90 kg N/ha, followed by 60 kg N/ha and 30 N kg N/ha that
218 recorded at 3.7g and 3.26g respectively. The lowest total biomass was recorded in control treatment 0
219 kg N/ha that just recorded 3.13g. The increase of total plant biomass with increasing nitrogen levels
220 can be explained by the increase in plant sink strength with increasing nitrogen levels. As nitrogen
221 uptake increased, more of accumulation of dry biomass will be expected due to increase in plant sink
222 strength that can accommodate initiation of new plant sink There were no significant different
223 occurred in between 0 and 30 N kg/ha treatment ($p \geq 0.05$). The result of the present study was in
224 agreement with the research conducted by [23] where, they found that the dry weight of shoot
225 increased with the increase of nitrogen supplied in *I. aquatica*. This justifies that high availability of
226 nitrogen was important in increasing the dry biomass of *I. aquatica* that was observed in the present
227 study [24,25].
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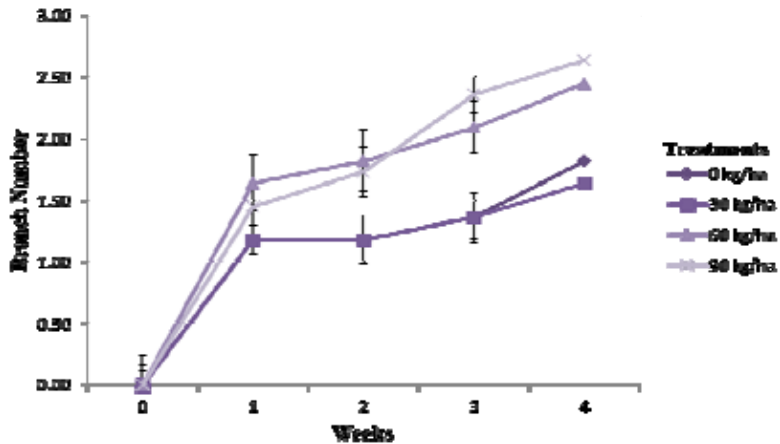


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244 **Fig. 3. The impact of different nitrogen rates on total biomass of *Ipomea aquatica*. N =10.**
245 **Bars represent standard error of differences between means (SEM).**
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248 3.4 Number of branches

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Figure 4 below shows the branches number of kangkung plant as affected by nitrogen treatments in all four weeks of treatment. As the higher rate of nitrogen treatments, the branching of plants was enhanced. At the first 2 weeks after the treatments were applied, the number of branches at 60 N kg/ha was higher than plants that were applied with 90 N kg/ha. But then, at week 3 and 4, the opposite results were obtained where the highest number of branches occurred at 90 N kg/ha. The study was in agreement with findings by Nashrin et al. [6] on *I. aquatica*, where the highest branching was obtained under highest nitrogen fertilization. Also, Osman and Abo Hassan [26], observed increased branching of Mangrove as nitrogen rate was increased to 100 kg N/ha. The increased in branching of the plant under high nitrogen fertilization might be due to increase in apical branches with higher nitrogen fertilization. This was due to enhanced vegetative growth under high nitrogen fertilization that enhanced the branching abilities of the plant [27].



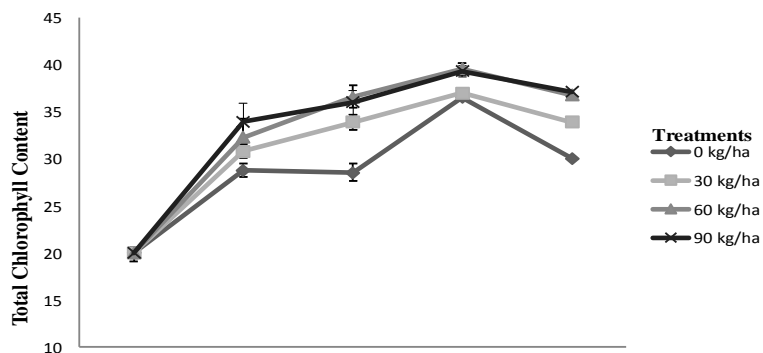
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Fig. 4. The impact of different nitrogen rates on the branch number of *Ipomea aquatica*. N =10. Bars represent standard error of differences between means (SEM).

3.5 Total Chlorophyll Content

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Figure 5 showed the impact of nitrogen fertilization on total chlorophyll content (TCC) of *I. aquatica* in 4 weeks of treatments. There were significant differences were observed for TCC in every week of measurement ($P \leq 0.05$). The chlorophyll content increased after week 1 and reached its maximum WAT content at week 3 as shown in Figure 5. In 1 WAT to 4 WAT, As the rate increased from 0 to 90 kg/ha, The TCC was steadily enhanced with the increasing nitrogen rates. In 2 -4 WAT there was no significant difference observed between 60 and 90 kg/ha in TCC. The study was in agreement with findings of According to Bojović and Marković [28] where the higher application of nitrogen increased the TCC in wheat, where establishes a linear relationship between the rates of nitrogen and the chlorophyll content in plants. The plant that has been treated with high N level will result in higher chlorophyll content where this might be due to the immediate absorbance of nitrogen in plant [29]. Since N is important for the structural element of chlorophyll and protein molecules, low N level will affect the formation of chloroplasts and the accumulation of chlorophyll in the plant [22]. Furthermore, as the plant age increased or getting mature, the N level tend to decrease and get mobilized to another part of the plant [29]. It can be concluded that in the present study, the higher rates of nitrogen application have increases the TCC in *I. aquatica*.

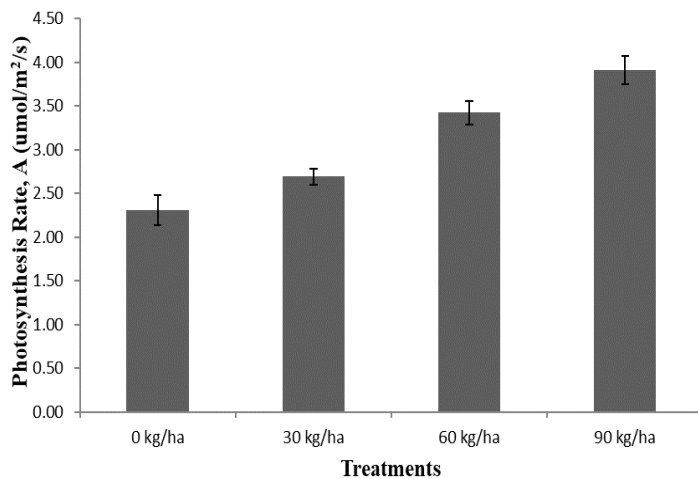


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Fig. 5. The impact of different nitrogen rates on the total chlorophyll content of *Ipomea aquatica*. N =10. Bars represent standard error of differences between means (SEM).

3.6 Photosynthesis rate (A)

The photosynthetic rate of *I. aquatica* was affected by four different nitrogen treatments. It was clearly observed that from the graph pattern, as the nitrogen rate become higher (0>90 kg/ha), the rate of photosynthesis also enhances (Figure 6). The highest A was observed in 90 kg/ha nitrogen, followed by 60 and 30 kg/ha, with the means of 3.91, 3.42, and 2.69 $\mu\text{mol}/\text{m}^2/\text{s}$ respectively. The lowest A was observed in 0 kg/ha where it just recorded 2.31 $\mu\text{mol}/\text{m}^2/\text{s}$. The increase in A under high nitrogen level might be due to increases in leaf area that correspondingly enhanced photosynthetic activity per plant [30]. The result was also in agreement with Boussadia et al. [31] where higher nitrogen content has shown to enhance the photosynthesis rate in olive plants. The nitrogen and photosynthesis activity is linked together because of the Calvin Cycle protein which represents the nitrogen in leaf [32]. At lower N level, the rate of photosynthesis was low. This might be due to the greater resistance and low biochemical of chloroplast [33]. According to Makino et al. [34], the increase in the rate of nitrogen leads to a greater N allocation to Rubisco. Rubisco is the primary CO_2 for enzyme fixation where the amount of this enzyme can drastically affect the photosynthesis rate. Besides, high N is needed in Rubisco protein due to the low rate of catalysis in Rubisco. It can be concluded that, enhanced application of nitrogen would enhance rubisco production that enhanced the net photosynthesis of *Ipomea aquatica* that was observed in the present study.



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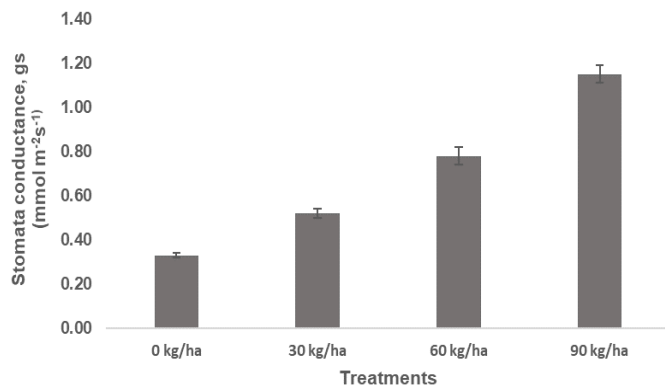
Fig. 6. The impact of different nitrogen rates on the photosynthesis rate of *Ipomea aquatica*. N =10. Bars represent standard error of differences between means (SEM).

3.7 Stomatal conductance (gs)

Based on Figure 7 below, it is distinctly observed that different rates of nitrogen had greatly affected the measurement of stomatal conductance. The higher the treatment concentrations (0,30,60,90

337 kg/ha), the rate of stomatal conductance have shown to increase. The stomatal conductance
338 measurement was 70%, 167% and 260% respectively higher in 30, 60 and 90 kg/ha Nitrogen
339 compared to the control that recorded only $0.33 \text{ mmol m}^{-2} \text{ s}^{-1}$. The present result was in agreement
340 with the findings of [35,36], where they found that the increase in photosynthesis rate and stomatal
341 conductance are correlated to increase in nitrogen application to the plants. Despite nitrogen, the size
342 of the leaf can be important for certain plant species as it helps for greater conductance through the
343 high number opening of the stomata [37]. This indicates that stomata conductance was enhanced
344 with high levels of nitrogen applied to *I. aquatica*.

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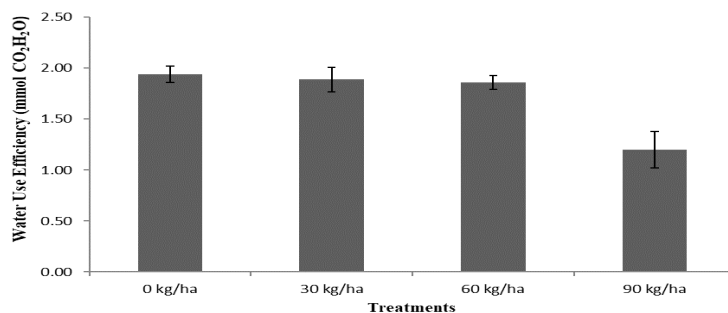
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Fig. 7. The impact of different nitrogen rates on the stomatal conductance of *Ipomea aquatica*. N =10. Bars represent standard error of differences between means (SEM).

3.8 Water use efficiency (WUE)

356 Water use efficiency (WUE) was illustrated in Figure 8 as it was influenced by the nitrogen treatments
357 ($P \leq 0.05$). Plant with the highest concentration of nitrogen (90 kg/ha) has the lowest measurement
358 recorded in water use efficiency with the mean of $1.46 \mu\text{mol CO}_2/\text{H}_2\text{O}$ transpired. While the highest
359 measurement in water use efficiency was recorded in the plant that was applied with 0 Kg/Ha nitrogen
360 with a mean of $1.97 \mu\text{mol CO}_2/\text{H}_2\text{O}$ transpired. The current result was contradicting with the findings
361 of Stewart [38] in cotton where the highest nitrogen application has shown to enhance the WUE in the
362 plant. The increased of WUE is usually, attributed to the increase of the transpiration rate and showed
363 plant under water stress condition. The current result showed that higher application of nitrogen rates
364 in *I. aquatica* can reduce the plant stress by having lower WUE. [22]. A similar result was obtained by
365 Artur et al. [39] where the increase of N has reduced the WUE in Marandu grass that showed a high
366 application of nitrogen can reduce stress in *I. aquatica*.

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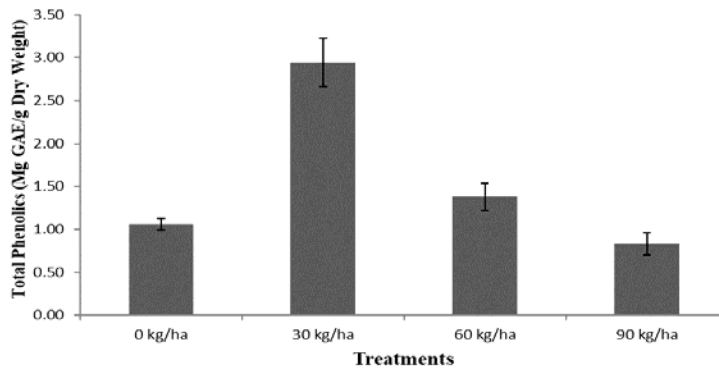
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Fig. 8. The impact of different nitrogen rates on the water use efficiency of *Ipomea aquatica*. N =10. Bars represent standard error of differences between means (SEM).

3.9 Total phenolics

Total plant phenolics contents were influenced by nitrogen fertilization ($P \leq 0,05$; Figure 9). As levels of nitrogen enhanced, the total phenolics content was seemed to be reduced. Total phenolics was 203%, 41% and 13% higher in 30 kg/ha, 60 kg/ha and 0 kg/ha respectively compared to 90 kg/ha treatments. The previous study had shown that when the level of nitrogen decreased, the phenolic compound increased in Broccoli [40]. Another result obtained by Stewart et al. [41], also prove that the phenolic content increased as the plant faced deficiency in nitrogen level. The result obtained in this study suggested that at lower nitrogen fertilization i.e. 30 kg N/ha the production of total phenolics in *Ipomea aquatica* was enhanced. According [42], when a plant undergoes N deficiency, the process of distributing carbon-based secondary compounds will increase, thus, decreasing the synthesis of nitrogen-based secondary compounds. Besides, Ibrahim et al. [19] stated that the increase in total phenolics production under low N level also might be due to the increase of total carbohydrate structural production that enhanced the production of carbon- based secondary metabolites.

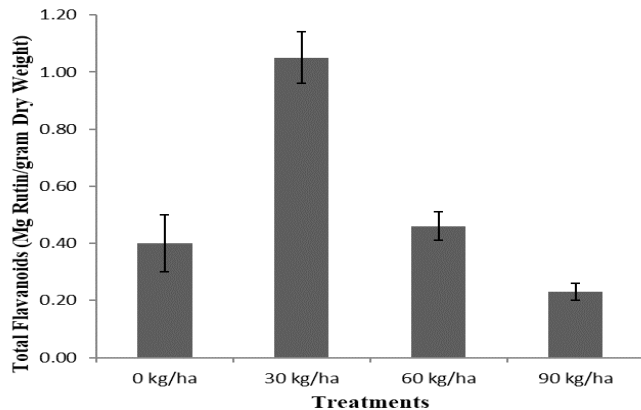


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Fig. 9. The impact of different nitrogen rates on total phenolics of *Ipomea aquatica*. N =10. Bars represent standard error of differences between means (SEM).

3.10 Total flavanoids

The total flavonoids of *Ipomea aquatica* were observed to be affected by the different rates of nitrogen treatments (Figure 10; $P \leq 0.05$). The production of total flavonoids has the same trends with total phenolics production content where plants which applied with 30 N kg/ha treatments has the highest total flavonoids content (1.05 mg Rutin/g dry weight) compared to 90 kg/ha that only recorded 0.27 mg rutin/ g dry weight. The same observation was obtained by [43] (2012) in Yaupon where the flavonoid content reduces when applied with high N rate. According to [44] the flavonoids content in plant tissues can be increased when having lower nitrogen content in the plant tissues. The increases in synthesis of flavonoid at lower nitrogen level might be due to increases in phenylalanine availability that enhances the phenylalanine lyase (PAL) activity that simultaneously enhanced the production of secondary metabolites [45]. It can be concluded in the present study, that under high nitrogen level the production of total phenolics and flavonoids was reduced in *I. aquatica*.



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Fig.10.The impact of different nitrogen rates on total flavonoids of *Ipomea aquatica*. N =10. Bars represent standard error of differences between means (SEM).

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4. CONCLUSION

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In the present work It was found that as the nitrogen rates increased, the growth and leaf gas exchange properties of *I. aquatica* was enhanced. However, the production of phenolics and flavonoids of kangkung was reduced with high levels of nitrogen application as both total phenolics and flavonoid reached the highest content at 30 kg N /ha. This work gives support that high nitrogen fertilization to *I. aquatica* can reduce the production of secondary metabolites although the growth parameters were enhanced with high nitrogen fertilization.

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REFERENCES

430
431

1. Malaysia Department of Agriculture. Ringkasan Maklumat Perangkaan Agromakanan.www.moa.gov.my.2011

432
433

2. Ismail A, Marjan ZM and Foong CW. Total antioxidant activity and phenolic contents in selected vegetables. Food Chem. 2004;87: 581–6.

434
435

3. Edie EE and Ho BWC. Ipomoea aquatica as a vegetable crop in Hong Kong. Econ. Bot. 1969; 23:32–6.

436
437
438

4. Nashrin S, Farooque A, Siddiqua M, Rahman M, and Khanam M. Effect of nitrogen and spacing on the growth and yield of gimakalmi, Ipomea reptans poir. J. Biol. Sci. 2002;76: 170-174.

439
440

5. Candlish I K, Gourley L and Lee HP. Dietary fiber and starch contents of some Southeast Asian vegetables. J. Agri. Food Chem. 1987; 35: 319–21.

441
442

6. Susila A, Prasetyo T, and Palada M. Optimum Fertilizer rate for kangkong (Ipomoea reptans L.) Production in Ultisols Nanggung.2008;34:12-34.

443
444

7. Ayodele O, Alabi E, and Aluko M. Nitrogen fertilizer effect on growth, yield, and chemical composition of hot pepper (rodo). Int. Agri. Crop Sci. 2015;8(5): 666-673.

445
446

8. Li J, Zhu Z, and Gerendas J. Effects of nitrogen and sulfur on total phenolics and antioxidant activity in two genotypes of leaf mustard. J. Plant Nutr. 2008; 31: 1642-1655.

447
448

9. Mokhele B, Zhan X, Yang G, and Zhang X. Nitrogen assimilation in crop plants and its affecting factors. Canadian Journal of Plant Science. 2011;17: 231-245.

- 449 10. Zaman MS, Hashem MA, Jahiruddin M, and Rahim MA. Effect of nitrogen for yield
450 maximization of garlic in old Brahmaputra flood plain soil. *Bangla. J. Agri. Res.* 2011; 36(2):
451 357-367.
- 452 11. Mansour MMF. Nitrogen containing compounds and adaptation of plants to salinity stress.
453 *Biol. Plantarum.* 2000; 43: 491-500.
- 454 12. Grusak MA, Della PD and Welch RM. Physiological process affecting the content and
455 distribution of phytonutrients in plants. *Nut. Rev.* 1999; 57(9): S27-S33.
- 456 13. Campostrini E, Yamanishi O, and Martinez E. Leaf Gas Exchange Characteristics of Four
457 Papaya Genotypes During Different Stages of Development. *Revista Brasileira de*
458 *Fruticultura.* 2001; 23(3): 522-525.
- 459 14. Hajiboland R and Beiramzadeh N. Growth, gas exchange and function of antioxidant defense
460 system in two contrasting rice genotypes under Zn and Fe deficiency and hypoxia. *Acta Biol.*
461 *Szegediensis* 2008; 52: 283-294.
- 462 15. Stiller V, Lafitte HR, Sperry JS. Hydraulic properties of rice and the response of gas exchange
463 to water stress. *Plant Physiol.* 2003; 132: 1698-1706.
- 464 16. Dingkuhn M; Cruz RT, O'Toole JC, Turner NC, Doerffling K. Responses of seven diverse rice
465 cultivars to water deficits. III. Accumulation of abscisic acid and proline in relation to leaf
466 water-potential and osmotic adjustment. *Field Crops Res.* 1991; 27: 103-117.
- 467 17. Dixon RA and Paiva NL. Stress-induced phenylpropanoid metabolism. *Plant Cell.* 1995; 7: 1
468 085-1097.
- 469 18. Due BM, Humphries D, Mai LTB, Dao AH, Co TM, Nga HH and Kim PT. Iron and vitamin C
470 content of commonly consumed foods in Vietnam. *Asia-Pacific Journal of Clinical Nutrition.*
471 1999; 8: 36-38.
- 472
473 19. Ibrahim MH, Jaafar HZE, Rahmant A, and Rahman Z. Effects of Nitrogen fertilization on
474 synthesis of primary and secondary metabolites in three varieties of kacang Fatimah (*Labisia*
475 *pumila blume*). *Int. J. Mol Sci.* 2011; 12(8): 5238-5254.
- 476 20. Taheri E, Soleymani A, and Javanmard HR. Effect of different levels of nitrogen on
477 morphological traits of two cultivars of rapeseed in isfahan region. *Int. J. Agri Crop Sci.*
478 2012;35: 1587-1590.
- 479
480 21. Amin MEM. Effect of different nitrogen sources on growth, yield and quality of fodder maize
481 (*Zea mays L.*). *J. Saudi Soc. Agri. Sci.* 2010; 67: 17-23.
- 482
483 22. Sarkar R, Jana J and Datta, S. Effect of cutting frequencies and nitrogen levels yield and
484 quality of water spinach (*Ipomoea reptans Poir.*). *Journal of Applied and Natural Science,*
485 2014;76: 545-551.
- 486
487 23. Phimmasan H, Kongvongxay S, Chhayty P and Preston T R. Water spinach (*Ipomoea*
488 *aquatica*) and Stylo 184 (*Stylosanthesguianesis CIAT 184*) as basal diets for growing rabbits.
489 *Livestock Res. for Rural Development.* 2004;16: 46-59.
- 490
491 24. Hare PD, Cress WA and Staden VJ. Proline synthesis and degradation: a model system for
492 elucidating stress-related signal transduction. *J. Exp. Bot.* 1999; 50: 413-34.
- 493
494 25. Prasad NK, Divakar S, Shivamurthy GR and Aradhya SM. Isolation of a free radical
495 scavenging antioxidant from water spinach (*Ipomoea aquatic Forsk.*). *J. Sci. Food Agri.* 2005;
496 85: 1 461-8.
- 497

- 498 26. Osman H, and Abu HA. Effect of NPK Fertilization on growth and dry matter accumulation in
499 mangrove [*Avicennia marina* (Forssk) vierh] Grown in Western Saudi Arabia. Meteorol.
500 Environ. Arid Land Agri.2010;56: 57-70.
501
- 502 27. Koo HM and Suhaila M. Flavonoid (Myricetin, quercetin, kaempferol, luteolin and apegenin)
503 content of edible tropical plants. J. Agri. Food Chem. 2011; 49: 3 106– 12.
504
- 505 28. Bojovic B and Markoviae A. Correlation Between Nitrogen and Chlorophyll Content In Wheat
506 (*Triticum aestivum* L.). Kragujevac J. Sci.2009; 3: 69-74.
507
- 508 29. Moreno N, Barrios A, Leal R, Franco A, Rodriguez A, and Hernandez L. Effect of Nitrogen
509 Deficiency and Toxicity in Two Varieties of Tomato (*Lycopersicum esculentum* L.). Agri.
510 Sci.2014; 5: 1361-1368.
511
- 512 30. Mansour MMF. Nitrogen containing compounds and adaptation of plants to salinity stress.
513 *Biologia Plantarum*. 2000; 43: 491–500.
514
- 515 31. Boussadia K, Steppe K, Labeke MC, Lemeur R and Braham M. Effects of Nitrogen deficiency
516 on Leaf Chlorophyll Fluorescence Parameters in Two Olive Tree Cultivars Meski' and
517 'Koroneiki'. J. Plant Nutr.2015;76; 2230-2246.
518
- 519 32. Baque MA, Karim MA, Hamid A, Tetsushi H. Effects of fertilizer potassium on growth, yield
520 nutrient uptake of wheat (*Triticum aestivum*) under water stress conditions. South Pac.
521 Studies 2006; 27: 29–35.
522
- 523 33. Grzebisz W, Gransee A, Szczepaniak W, Diatta J. The effects of potassium fertilization on
524 water-use efficiency in crop plants. J. Plant Nutr. Soil Sci. 2013: 176: 355–374.
525
- 526 34. Makino, A. Photosynthesis, Grain Yield, and Nitrogen Utilization in Rice and Wheat. *Plant*
527 *Physiology*. 2011; 155: 125-129.
528
- 529 35. Pankovica D, Plesnicar M, Arsenijevicâ MI, Petrovica N, Sakaci Z, and Kastori R. Effects Of
530 Nitrogen nutrition on photosynthesis in cd-treated sunflower plants. *Annals of Bot.*,2000; 86:
531 841-847.
532
- 533 36. Nori M, Bayat F, and Esmaeili A. Changes of vegetative growth indices and yield of garlic
534 (*Allium sativum* L.) in different sources and levels of nitrogen fertilizer. *Int. J. Agri. Crop*
535 *Sci.*2012;67; 1394-1400.
536
- 537 37. Schulze ED, Kelliher F, Korner C, Lloyd J, and Leuning R. Relationships among maximum
538 stomatal conductance, ecosystem surface conductance, carbon assimilation rate, and plant
539 nitrogen nutrition: a global ecology scaling exercise. *Ann. Rev. of Ecol. Systematics*.1994; 25:
540 629-660.
541
- 542 38. Stewart W. Balanced fertilization increases water use efficiency. Atlanta, Georgia: Potash &
543 Phosphate Institute (PPI) and Potash & Phosphate Institute of Canada (PPIC).2001.
544
- 545 39. Artur A, Garcez T, and Monteiro F. Water use efficiency of marandu palisadegrass as
546 affected by nitrogen and sulphur rates. *Artigo Cientifico*. 2014; 45(1): 10-17.
547
- 548 40. Naguib, AM. Enhancement of phenolics, flavonoids and glucosinolates of Broccoli (*Brassica*
549 *olaracea*, var. *Italica*) as antioxidants in response to organic and bio-organic fertilizers.
550 *Journal of the Saudi Society of Agricultural Sciences*.2013;78: 135-142.
551
- 552 41. Stewart A, Chapman W, Jenkins G, Graham I, Martin T, and Crozier A. The effect of nitrogen
553 and phosphorus deficiency on flavonol accumulation in plant tissues. *Plant Cell Environ.*2011;
554 24: 1189-1197.
555
- 556 42. Orphanides A, Goulas V and Gekas V. Effect of drying method on the phenolic content and
557 antioxidant capacity of spearmint. *Czech J. Food Sci.*2013; 31(5): 509-513.

558
559
560
561
562
563
564
565
566
567
568
569

43. Palumbo, M., Putz, F., & Talcott, S. Nitrogen fertilizer and gender effects on the secondary metabolism of yaupon, a caffeine-containing north American holly. *Oecologia*.2006; 151(1): 1-9 .
44. Meyer S, Cerovic ZG, Goulas Y, Montpied P, Demotes S, Bidet LPR, Moya I and Dreyer E. Relationship between assessed polyphenols and chlorophyll contents and leaf mass per area ratio in woody plants. *Plant Cell Environ*. 2006;29: 1338-1348.
45. Margna U, Margna E and Vainjarv T. Influence of nitrogen nutrition on the utilization of L-phenylalanine for building flavonoids in buckwheat seedling tissue. *J. Plant Physiol*. 1989;134:697-702.