

Original Research Article

Effect of Nitrogen Rates on Growth, Carbon Assimilation and Quality of Water Spinach (*Ipomea aquatica*)

ABSTRACT

Aims: This study was conducted to investigate the impact of the nitrogen fertilization on the growth, leaf gas exchange and bio-metabolite accumulation in *Ipomea aquatica*.

Study design: *Ipomea aquatica* plants were exposed to four different rates of nitrogen (0, 30, 60 and 90 N kg/ha) using NPK green fertilizer as a nitrogen source using Complete Randomize Design (CRD).

Place and Duration of Study: Department of Biology, Faculty of Science, Universiti Putra Malaysia between September to November 2016.

Methodology: There were four nitrogen rates were applied (0, 30, 60 and 90 N Kg/ha) using NPK green fertilizer as a nitrogen source. The growth data collections were conducted once a week after the application of the treatments for the plant growth parameter. The total chlorophyll content in the leaves was measured using a SPAD chlorophyll meter. The leaf gas exchange was determined using a LI-6400XT portable photosynthesis system. Total phenolics and flavonoid was determined using Folin-Ciocalteu reagent.

Results: It was found that the growth parameters which are plant height, leaf numbers, branches numbers, total biomass and chlorophyll content recorded the highest measurement at 90 kg N/ha and the lowest at 0 kg N/ha. As for the leaf gas exchange, the positive effect of nitrogen fertilization on kangkung was shown by the increased in photosynthesis rate (A) and stomatal conductance (gs) where the highest measurement recorded at 90 kg N/ha, while the lowest at 0 kg N/ha. However, the water use efficiency (WUE) decreased as the nitrogen rates increased. At lower rates of nitrogen fertilization (30 kg N/ha) produced the highest production of secondary metabolites, where the total phenolics and flavonoids production were enhanced compared to other nitrogen treatments.

Conclusion: In conclusion, as the nitrogen rates increased, the growth and leaf gas exchange properties was enhanced however the production of total phenolics and flavonoids was reduced and get the highest accumulation at 30 kg N/ha.

Keywords: [Nitrogen, *Ipomea aquatica*, growth, leaf gas exchange, biometabolites production]

1. INTRODUCTION

In Malaysia, agriculture sector has contributed about 8.5% to Gross Domestic Products (GDP). About 39% of the contributions originated from the production of food crops, fruits and vegetables. It is estimated that there are about 44, 000 hectares of the total area in Malaysia was used for vegetable cultivation [1]. According to Department of Agriculture Malaysia in 2011, *Ipomea Aquatica* is one out of ten types of vegetables that consumed the highest area for vegetable production. This plant is among the most consumed vegetable in Asia. This is because of it's low price compared to other types of vegetable. Kangkung air or it's scientific name, *Ipomea aquatica*, is a widely known leafy vegetable especially in Asian country. The plant is also commonly known with different local names, such as water spinach, swamp cabbage, or water convolvulus. From it's scientific classification, kangkung has been classified under a family of Convolvulaceae [2]. According to [3], Convolvulaceae family consist of primarily 1650 of tropical species. Moreover, the genus of kangkung which is *Ipomea* has about 500 to 600 different species and it has been the most number of containing species in Convolvulaceae family [4]. This species of family can nicely grown at almost anywhere at the higher or even the lower land. *Ipomea aquatica* is one of the species that is cultivated on the higher land. Besides easy to be grown, kangkung often be the favorable plant to be cultivated because it does not take long time to mature and harvest. It can easily adapt towards it's grow environment and usually unsusceptible to disease. Almost all parts of kangkung plant are edible [3].

32 According to Susila et al. [5], nitrogen is the primary nutrient that involve in producing a high yield of
33 vegetables. Nitrogen is one of the macro-nutrients that is very crucial especially for a plant to have a
34 proper growth and development [6] such as that required in constructing the matter of the plant cell
35 and tissue [7]. The amount of nitrogen in the soil could be insufficient for the plant to grow. Therefore,
36 the source of nitrogen for plant especially in agriculture field often be found in the form of a fertilizer.
37 Both organic and inorganic nitrogen fertilizer are widely used in agriculture especially in cultivating
38 green crops to keep the source of nutrients for plant being supply [6]. Practically, an appropriate and
39 suitable amount of nitrogen to be given to plant will affect it's crop yield. Nitrogen also is very
40 important especially to promote the growth of the plant leaf [8]. Nitrogen is a crucial element not only
41 to promote the growth and plant development, also increase yield and quality in vegetable crops.
42 Increasing level of nitrogen resulting in a number of leaves, leaf length and plant body [8]. Nitrogen
43 also enhanced the size of fruits and vegetables where at optimum application of N will resulting in a
44 better size. The metabolic process which stimulated by N by enhance the vegetative and also the
45 reproductive growth in plant. Besides, high plant biomass can be obtained when there is high N
46 accumulated in shoot, along with the increasing of root growth in plant if there is sufficient amount of
47 N supply [9]. However, the lack of N in plant would caused the reduced in plant development and
48 eventually will lower the crop yield.

49

50 Nitrogen had been proven to have a strong relationship with photosynthesis process in plant.
51 Increasing N level lead to higher N content in leaf. N also enhance the leaf chlorophyll and CO₂
52 assimilation which increase in the Rubisco activity [10]. Therefore, increase in rate of photosynthesis
53 photosynthesis is the most vital biochemical process in plants [11]. According to [12,13], the
54 photosynthesis rate (A) depends on the growth development of the plant's leaf. The leaf development
55 includes the increase in leaf area, leaf thickness, the surface volume of mesophyll cells, and leaf
56 chloroplast. The photosynthesis rate will be increased as the leaf development also increased [14].
57 Nitrogen is an element that has a significance role in photosynthesis which involve in the opening of
58 the stomata. The stomatal vent will decrease following the nitrogen deficit which then will decrease
59 the transpiration rate [15].

60

61 Secondary metabolites such as phenolic in plant usually associated with the plant survival and health
62 benefits for those who consume the plant. Low nitrogen level in plant has been reported to have more
63 secondary metabolites compare to plant that has high N level [16]. Application of more N level
64 resulting in decrease of phenolic concentrations based on carbon/nutrients balance (CNB) hypothesis
65 [17]. Flavanoids also a secondary metabolite which is widely distributed with different functions in
66 plants. The biological functions of flavonoids include defense against UV-B radiation, pathogen
67 infection, nodulation and pollen fertility [18]. A study done by [8] on leaf mustard where the total
68 phenolics concentration was observed to be decreased as the level increased. It is well known that
69 nitrogen application can directly affect the morphological growth and yield of this plant, however, little
70 work has been carried out to look on the impact of nitrogen of the leaf gas exchange properties and
71 previous work have not comprehensively considered the production of secondary metabolites of *I.*
72 *aquatica* under nitrogen fertilization. there was no study about the production of secondary
73 metabolites and leaf gas exchange properties under different nitrogen rates. The main aim of the
74 research was to investigate the effect of nitrogen fertilization on the growth, leaf gas exchange and
75 production of secondary metabolites of *I. aquatica* and to determine the best nitrogen rates for growth
76 and development of *I. aquatica*. This research will provide the important information for vegetable
77 growers that involved in cultivation of vegetables in Malaysia.

78

79 2. MATERIAL AND METHODS

80

81 2.1. Plant material and maintenance

82

83 The experiment was conducted at Department of Biology, Universiti Putra Malaysia, Serdang (UPM),
84 Selangor where the area of the experiment was located at the site where the direct sunlight is always
85 sufficient and available. These seeds were allowed to germinate for two weeks. After all the seeds
86 has been germinated, all the plants then were transplanted into the polybags filled with a mixture of
87 topsoil, organic matter and sand with the ratio of 3:2:1. The nitrogen sources used were from the NPK
88 green fertilizer (15:15:15). The polybags were arranged according to Completely Randomized Design
89 (CRD) with five replications. There were four nitrogen rates were applied (0, 30, 60 and 90 Kg N/ha)
90 with overall 160 of *I. aquatica* plants were used. The growth data collections were conducted once a
91 week for four weeks after the application of the treatments for the plant growth parameter. Whereas

92 the destructive analysis and leaf gas exchange of the experiment was conducted at the end of the
93 experiment.

94

95 **2.2. Plant height, leaf and branch numbers**

96

97 As for plant height, it was measured starting from the stem that was at the soil surface up until the
98 highest shoot grow or at tip using measuring tape. The leaf and branches number were counted
99 manually per plant basis

100

101 **2.3 Plant total dry weight measurement**

102

103 The plants were first removed from the soil carefully and the dirt from the soil were washed with tap
104 water. After that, the shoot and the root parts were separated. All the plants were dried in an oven for
105 48 hours at temperature of 60°C until constant weight reached.

106

107 **2.4 Total Chlorophyll content**

108

109 SPAD chlorophyll meter was used to measure the total chlorophyll content of the leaves. Three
110 readings were taken at three spot on a leaf of each plant and the average readings were recorded.
111 Time interval between 9.00 a.m and 12.00 p.m was used to measure the chlorophyll content.

112

113 **2.5 Leaf gas exchange measurement**

114

115 The leaf gas exchange measurement was obtained after week 4 the treatment was given. The result
116 then was obtained by using the Portable Photosynthesis System machine (LICOR 6400 XT). The
117 IRGA was firstly warm up for at least 30 minutes before the leaf gas exchange was collected with
118 Zero IRGA mode. The optimal condition was set to 400 $\mu\text{mol mol}^{-1}$ carbon dioxide (CO_2), 30 °C
119 cuvette temperature, 60% relative humidity with air flow rate set at 500 $\text{cm}^3 \text{min}^{-1}$, and 800 $\mu\text{molm}^{-2}\text{s}^{-1}$
120 of cuvette condition of photosynthetic photon flux density (PPFD). The time for the measurement were
121 done at the morning of a day. The measurement of photosynthesis rate were taken from the first
122 kangkung leaves starting from the plant apex. The data then were recorded and stored in a console of
123 the system and analyse with Photosyn Assistant Software. The photosynthesis (A), transpiration rate
124 (E), stomata conductance (gs) and water use efficiency (WUE) data was recorded during the
125 measurement.

126

127 **2.7 Total phenolics and flavonoids quantification**

128

129 The methods used for extraction and quantification of total phenolics and flavonoids contents followed
130 that described in Ibrahim et al. [19]. A fixed amount of ground tissue samples (0.1 g) was extracted
131 with 80% ethanol (10 mL) on an orbital shaker for 120 min at 50 °C. The mixture was subsequently
132 filtered (Whatman™ No.1), and the filtrate was used for the quantification of total phenolics and total
133 flavonoids. Folin–Ciocalteu reagent (diluted 10-fold) was used to determine total phenolics content of
134 the leaf samples. The sample extract at 200 μL was mixed with Folin–Ciocalteu reagent (1.5 mL) and
135 allowed to stand at 22 °C for 5 min before adding NaNO_3 solution (1.5 mL, 60 g L^{-1}). After two hours
136 at 22 °C, absorbance was measured at 725 nm. The results were expressed as mg g^{-1} gallic acid
137 equivalent (mg GAE g^{-1} dry sample). For total flavonoids determination, samples (1 mL) were mixed
138 with NaNO_3 (0.3 mL) in a test tube covered with aluminium foil, and left for 5 min. Then 10% AlCl_3 (0.3
139 mL) was added followed by addition of 1 M NaOH (2 mL). The absorbance was measured at 510 nm
140 using a spectrophotometer with rutin as a standard (results expressed as mg/g rutin dry sample).

141

142 **2.8 Statistical analysis**

143

144 Data were analysed using the analysis of variance procedure in SAS version 17. Means separation
145 between treatments was performed using Duncan multiple range test and the standard error of
146 differences between means was calculated with the assumption that data were normally distributed
147 and equally replicated.

148

149

150 3. RESULTS AND DISCUSSION

151

152 3.1 Plant height

153

154 Figure 1 shows the plant height of *I. aquatica* as influenced by differing nitrogen treatments. As can
 155 be seen, the plant height of kangkung were mostly affected by different rates of nitrogen treatment in
 156 all week of measurement ($P \leq 0.05$). In view of the result obtained, as nitrogen levels increased from 0
 157 to 90 kg N/ha the plant height was enhanced in all weeks of measurement. In 4 weeks after treatment
 158 (4 WAT), plant at 0 kg/ha have the average of 31.02 cm compared to 32.17 cm by 30 kg/ha, 35.61 cm
 159 by 60 kg/ha and 37.24 cm in 90 kg/ha. Clearly, as expected, applying higher rates of nitrogen levels
 160 would enhance the plant height of *I. aquatica*. The positive effects on plant height cause by the
 161 increase of nitrogen rates application may be due to the natural role of nitrogen on vegetative growth
 162 performance of plants [6]. The increase in plant height under nitrogen fertilization might be due to
 163 well-developed stem under high nitrogen fertilization that resulted in taller plant [20]. Besides that,
 164 increase in plant height might be associated with the increased of number and length of the
 165 internodes by nitrogen [21]. The result obtained agreed with the previous work carried out by [4] and
 166 [6] where the increment of nitrogen fertilization rates applied towards *I. aquatica* had significantly
 167 increased the plant height at end of the harvesting period. It can be concluded, that in the present
 168 study, that high application of nitrogen has shown to enhance the height of *I. aquatica*.

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

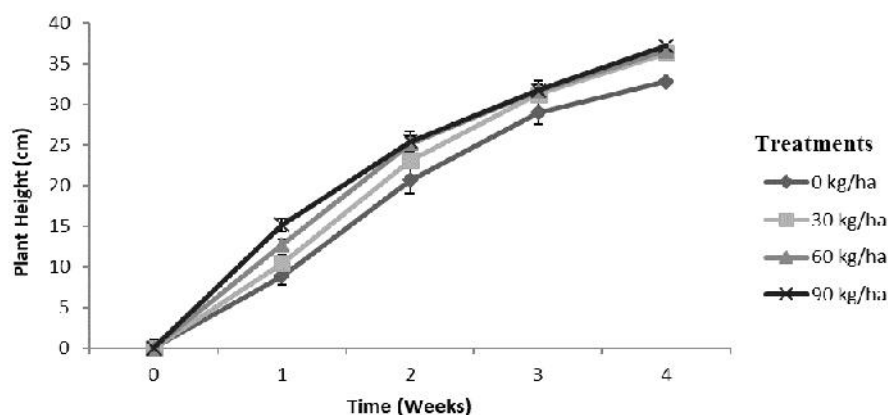
201

202

203

204

205



186 **Fig 1. The impact of different nitrogen rates on the height of *Ipomea aquatica*. Mean**
 187 **with the same letter indicates that all of the groups were not significantly different**
 188 **according to Duncan multiple range test ($P \geq 0.05$) N=10.**

190 3.2 Leaves numbers

191

192

193

194

195

196

197

198

199

200

201

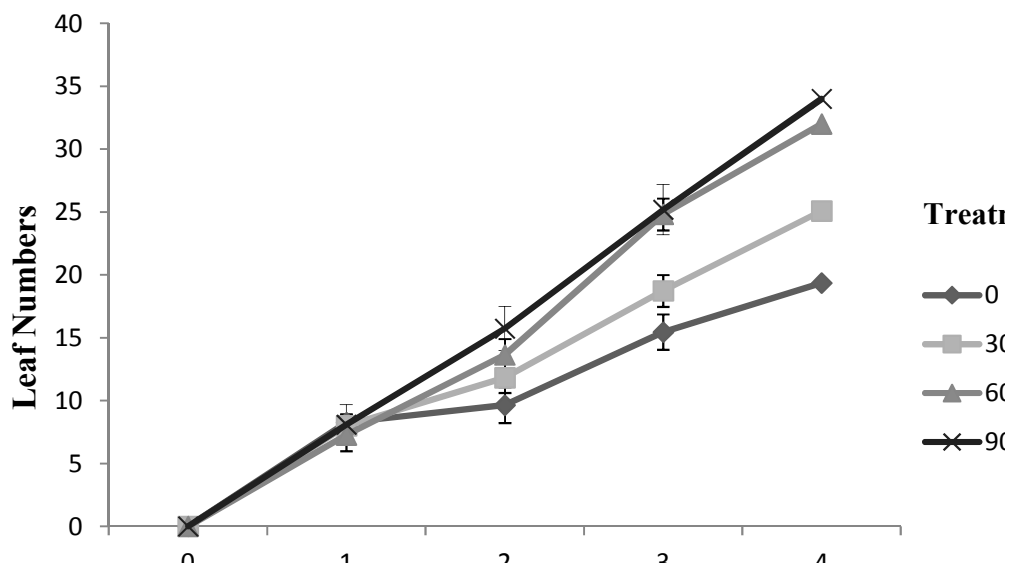
202

203

204

205

The variation of leaf numbers with different nitrogen fertilization in *I. aquatica* is depicted in Figure 2. Generally, leaf number of *I. aquatica* plant was found to be influenced by the different rates of nitrogen treatments (0, 30, 60 and 90 kg/ha; $P \leq 0.05$). Based on Figure 2, it shows that there were significant effects of nitrogen fertilization rates on the number of leaves in every week of measurements. Overall at 90 N kg/ha as the highest treatments of nitrogen applied, lead to the drastic production in number of leaves from 1 to 4 WAT. An increase in number of leafage in plants indicate better plant growth and development. Eventually, the plant production also will increase. Similar trends were observed in [6] and [20] where they found that as the rate of nitrogen increases the *I. aquatica* leaf numbers were also enhances. The increase in leaf number in *I. aquatica* might be due to increase in internodes number with high application of nitrogen [21]. The high application of nitrogen usually would reduce the apical dominance and stimulated the development of lateral buds that eventually increase the production of plant leaf and simultaneously enhanced the leaf numbers [22].



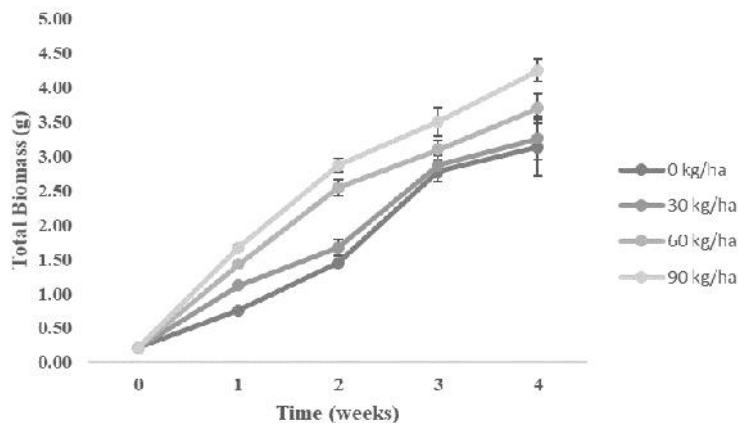
206
207
208
209
210
211

Fig 2. The impact of different nitrogen rates on the leaves numbers of *Ipomea aquatica*. Mean with the same letter indicates that all of the groups were not significantly different according to Duncan multiple range test ($P \geq 0.05$) $N=10$.

3.3 Plant Total dry biomass

212
213
214
215
216
217
218
219
220
221
222
223
224
225
226
227
228
229

Nitrogen application had significantly influenced on the total plant dry weight of *I. aquatica* plant as shown in Figure 3. The graph pattern show increased in production in total biomass with higher application of nitrogen fertilization rates. At end of the treatments, It was observed that the highest total biomass of kangkung was obtained in 90 kg N/ha, followed by 60 kg N/ha and 30 N kg N/ha that recorded at 3.7g and 3.26g respectively. The lowest total biomass was recorded in control treatment 0 kg N/ha that just recorded 3.13g. The increase of total plant biomass with increasing nitrogen levels can be explained by the increase in plant sink strength with increasing nitrogen levels. As nitrogen uptake increased, more of accumulation of dry biomass will be expected due to increase in plant sink strength that can accommodate initiation of new plant sink There were no significant different occurred in between 0 and 30 N kg/ha treatment ($p \geq 0.05$). The result of the present study was in agreement with the research conducted by [23] where, they found that the dry weight of shoot increased with the increase of nitrogen supplied in *Ipomea aquatica*. This justify that high availability of nitrogen was important in increasing the dry biomass of *I. aquatica* that was observed in the present study [24,25].



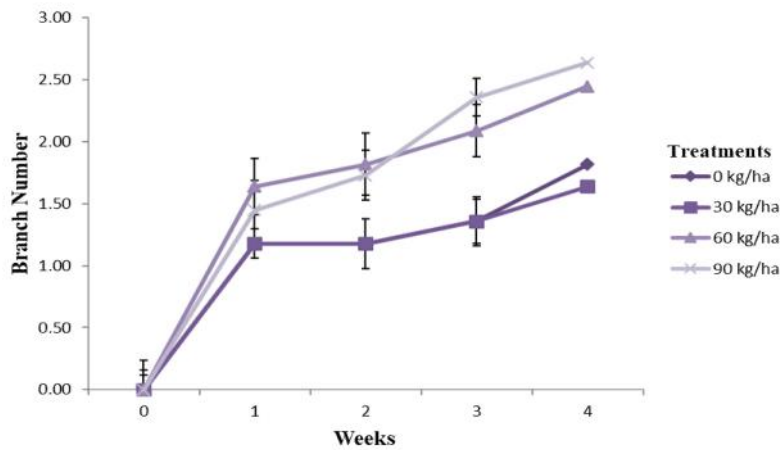
230
231

232
233
234
235
236
237
238
239
240
241
242
243
244
245
246
247
248
249
250
251

Fig. 3. The impact of different nitrogen rates on total biomass of *Ipomea aquatica*. Mean with the same letter indicates that all of the groups were not significantly different according to Duncan multiple range test ($p \geq 0.05$) N=10.

3.4 Number of branches

The Figure 4 below shows the branches number of kangkung plant as affected by nitrogen treatments in all four weeks of treatment. As the higher rate of nitrogen treatments, the branching of plants was enhanced. At the first 2 weeks after the treatments were applied, the number of branches at 60 N kg/ha were higher than plants that were applied with 90 N kg/ha. But then, at week 3 and 4, the opposite results were obtained where the highest number of branches occurred at 90 N kg/ha. The study was in agreement with findings by Nashrin et al. [6] on *Ipomea aquatica*, where the highest branching was obtained under highest nitrogen fertilization. Also, Osman and Abo Hassan [26], observed increased branching of Mangrove as nitrogen rate was increased to 100 kg N/ha. The increased in branching of the plant under high nitrogen fertilization might be due to increase in apical branches with higher nitrogen fertilization. This was due to enhanced vegetative growth under high nitrogen fertilization that enhanced the branching abilities of the plant [27].



252
253
254
255
256
257
258
259
260
261
262
263
264
265
266
267
268
269
270
271
272
273
274
275
276

Fig. 4. The impact of different nitrogen rates on the brancg number of *Ipomea aquatica*. Mean with the same letter indicates that all of the groups were not significantly different according to Duncan multiple range test ($p \geq 0.05$) N=10.

3.5 Total Chlorophyll Content

Figure 5 showed the impact of nitrogen fertilization on total chlorophyll content (TCC) of *I. aquatica* in 4 weeks of treatments. There were significant differences was observed for TCC in every weeks of measurement. The chlorophyll content increased after week 1 and reached it's maximum WAT content at week 3 as shown in Figure 5. In 1 WAT to 4 WAT, As the rate increased from 0 to 90 kg/ha, The TCC was steadily enhanced with the increasing nitrogen rates. In 2 -4 WAT there were no significant difference observed between 60 and 90 kg/ha in TCC. The study was in agreement with findings of According to Bojović and Marković [28] where higher application of nitrogen increased the TCC in wheat, where establishes a linear relationship between the rates of nitrogen and the chlorophyll content in plants. The plant that has been treated under high N level will resulted in higher chlorophyll content where this might be due to the immediate absorbance of nitrogen in plant [29]. Since N is important for the structural element of chlorophyll and protein molecules, low N level will affect the formation of chloroplasts and the accumulation of chlorophyll in the plant [22]. Furthermore, as the plant age increased or getting mature, the N level tend to decrease and get mobilized to other part of the plant [29]. It can be concluded that in the present study, the higher rates of nitrogen application have increases the TCC in *I. aquatica*.

277
 278
 279
 280
 281
 282
 283
 284
 285
 286
 287
 288
 289
 290
 291
 292
 293
 294
 295
 296
 297
 298
 299
 300
 301
 302
 303
 304
 305
 306
 307
 308
 309
 310
 311
 312
 313
 314
 315
 316
 317
 318

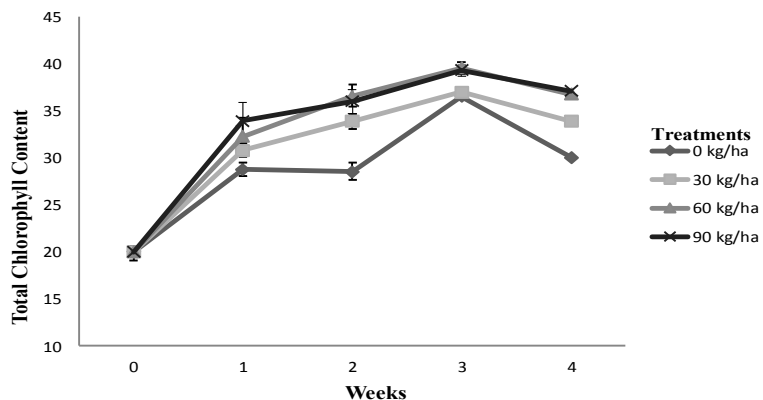
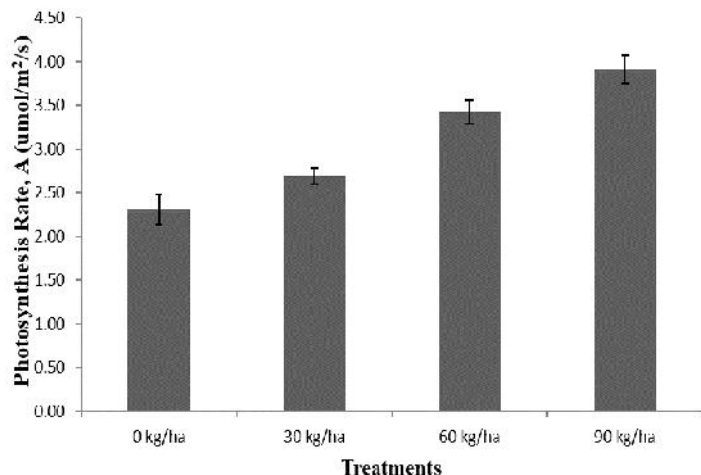


Fig. 5. The impact of different nitrogen rates on the total chlorophyll content of *Ipomea aquatica*. Mean with the same letter indicates that all of the groups were not significantly different according to Duncan multiple range test ($P \geq 0.05$) $N=10$.

3.6 Photosynthesis rate

The photosynthesis rate of *I. aquatica* was affected by four different nitrogen treatments. It is clearly observed that from the graph pattern, as the nitrogen rate fertilization become higher ($0 > 90$ kg/ha), the rate of photosynthesis also enhances (Figure 6). The highest A was observed in 90 kg/ha nitrogen, followed by 60 and 30 kg/ha, with the means of 3.91, 3.42, and 2.69 $\mu\text{mol}/\text{m}^2/\text{s}$ respectively. The lowest A was observed in 0 kg/ha where it just recorded 2.31 $\mu\text{mol}/\text{m}^2/\text{s}$. The increase in A under high nitrogen level might be due to increases in leaf area that correspondingly enhanced photosynthetic activity per plant [30]. The result was also in agreement with Boussadia et al. [31] where higher nitrogen content have shown to enhanced the photosynthesis rate in olive plants. The nitrogen and photosynthesis activity is linked together because of the Calvin Cycle protein which represent the nitrogen in leaf [32]. At lower N level, the rate of photosynthesis was low. This might be due to the greater resistance and low biochemical of chloroplast [33]. According to Makino et al. [34] the increase in rate of nitrogen leads to a greater N allocation to Rubisco. Rubisco is the primary CO_2 for enzyme fixation where the amount of this enzyme can drastically affect the photosynthesis rate. Besides, high N is needed in Rubisco protein due to the low rate of catalysis in Rubisco. It can be concluded that, enhanced application of nitrogen would enhance rubisco production that enhanced the net photosynthesis of *Ipomea aquatica* that was observed in the present study.



319

320 **Fig. 6. The impact of different nitrogen rates on the photosynthesis rate of *Ipomea***
 321 ***aquatica*. Mean with the same letter indicates that all of the groups were not**
 322 **significantly different according to Duncan multiple range test ($P \geq 0.05$) N=10.**
 323

324

325

326

3.7 Stomatal conductance

327

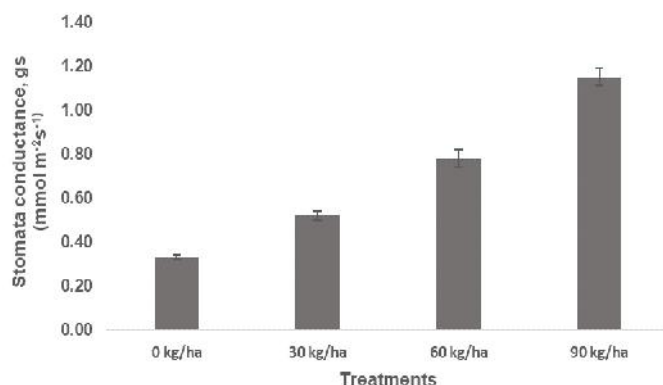
328 Stomatal conductance can be defined as the rate of carbon dioxide uptake and the water loss through
 329 stomatal leaves [35]. Based on Figure 7 below, it is distinctly observed that different rates of nitrogen
 330 had greatly affected the measurement of stomatal conductance. The higher the treatment
 331 concentrations (0,30,60,90 kg/ha), the rate of stomatal conductance have shown to increased. The
 332 stomatal conductance measurement was the highest at 90 N kg/ha ($1.15 \text{ mmol m}^{-2} \text{ s}^{-1}$), while the
 333 lowest rate of stomatal conductance was measured at 0 kg/ha nitrogen treatment that recorded 0.33
 334 $\text{mmol m}^{-2} \text{ s}^{-1}$. The present result was in agreement with the findings of [36], where they found that the
 335 increase in photosynthesis rate and stomatal conductance are correlated to increase in nitrogen
 336 application to the plants. Despite nitrogen, the size of the leaf can be important for certain plant
 337 species as it helps for greater conductance through the high number opening of the stomata [37]. This
 338 indicate that stomata conductance was enhanced with high levels of nitrogen applied to *I. aquatica*.

339

340

341

342



343

344

345

346 **Fig. 7. The impact of different nitrogen rates on the stomatal conductance of *Ipomea aquatica*.**
 347 **Mean with the same letter indicates that all of the groups were not significantly different**
 348 **according to Duncan multiple range test ($P \geq 0.05$) N=10.**
 349

350

351

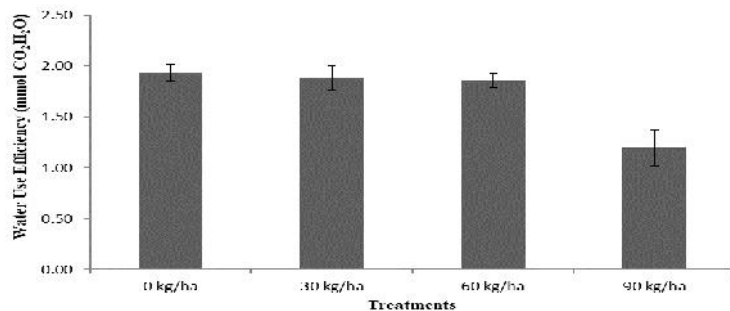
3.8 Water use efficiency (WUE)

352

353 Water use efficiency (WUE) was illustrated in Figure 8 as it was influenced by the nitrogen treatments
 354 ($P \leq 0.05$). Plant with the highest concentration of nitrogen (90 kg/ha) has the lowest measurement
 355 recorded in water use efficiency with the mean of $1.46 \mu\text{mol CO}_2/\text{H}_2\text{O}$ transpired. While the highest
 356 measurement in water use efficiency was recorded in plant that was applied with 90 Kg/Ha nitrogen
 357 with mean of $1.97 \mu\text{mol CO}_2/\text{H}_2\text{O}$ transpired. The current result was contradicting with the findings of
 358 Stewart [38] in cotton where the highest nitrogen application has shown to enhanced the WUE in the
 359 plant. The increased of WUE is usually, attributed to the increase of the transpiration rate and showed
 360 plant under water stress condition. The current result showed that higher application of nitrogen rates
 361 in *I. aquatica* can reduce the plant stress by having lower WUE. [22]. A similar result was obtained by
 362 Artur et al. [39] where the increase of N has reduced the WUE in Marandu grass that showed high
 363 application of nitrogen can reduce stress in *I. aquatica*.

364

365

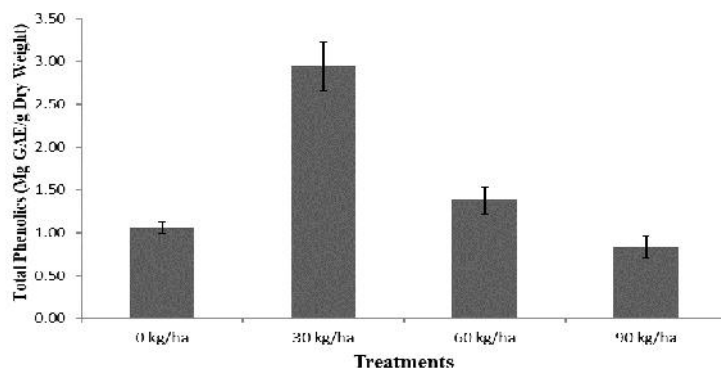


366
367
368
369
370
371
372
373
374
375
376
377
378
379
380
381
382
383
384
385
386
387
388

Fig. 8. The impact of different nitrogen rates on the water use efficiency of *Ipomea aquatica*. Mean with the same letter indicates that all of the groups were not significantly different according to Duncan multiple range test ($P \geq 0.05$) $N=10$.

3.9 Total phenolics

Total plant phenolics contents was influenced with nitrogen fertilization ($P \leq 0,05$; Figure 9). As levels of nitrogen enhanced, the total phenolics content was seemed to be reduced. Total phenolics was 203%, 41% and 13% higher in 30 kg/ha, 60 kg/ha and 0 kg/ha respectively compared to 90 kg/ha treatments. Previous study had showed that when the level of nitrogen decreased, the phenolic compound increased in Broccoli [40]. Another result obtained by Stewart et al. [41], also prove that the phenolic content increased as the plant faced deficiency in nitrogen level. The result obtained in this study suggested that at lower nitrogen fertilization i.e. 30 kg N/ha the production of total phenolics in *Ipomea aquatica* was enhanced. According [42], when a plant undergo N deficiency, the process of distributing carbon-based secondary compounds will increase, thus, decreasing the synthesis of nitrogen-based secondary compounds. Besides, Ibrahim et al. [19] stated that the increased in total phenolics production under low N level also might be due to the increase of total carbohydrate structural production that enhanced the production of carbon based secondary metabolites.



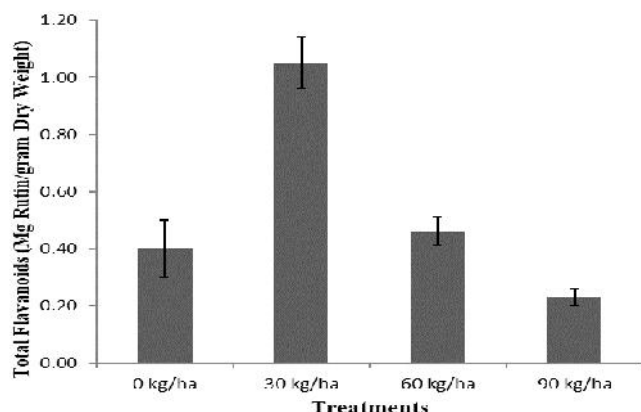
389
390
391
392
393
394
395
396
397
398
399
400
401

Fig. 9. The impact of different nitrogen rates on total phenolics of *Ipomea aquatica*. Mean with the same letter indicates that all of the groups were not significantly different according to Duncan multiple range test ($P \geq 0.05$) $N=4$.

3.10 Total flavanoids

The total flavanoids of *Ipomea aquatica* was observed to be affected by the different rates of nitrogen treatments (Figure 10; $P \leq 0.05$). The production of total flavanoids has the same trends with total phenolics production content where plants which applied with 30 N kg/ha treatments has the highest total flavanoids content (1.05 mg Rutin/g dry weight) compared to 90 kg/ha that only recorded 0.27

402 mg rutin/ g dry weight. The same observation was obtained by [43] (2012) in Yaupon where the
 403 flavonoid content reduces when applied with high N rate. According to [44] the flavonoids content in
 404 plant tissues can be increased when having lower nitrogen content in the plant tissues. The increases
 405 in synthesis of flavonoid at lower nitrogen level might be due to increases in phenylalanine availability
 406 that enhances the pheyllaline lyase activity that simultaneously enhanced the production of secondary
 407 metabolites [45]. It can be concluded in the present study, that under high nitrogen level the
 408 production of total phenolics and flavonoids was reduced in *I. aquatica*.
 409
 410



411
 412
 413
 414
 415
 416
 417
 418
 419

Fig.10.The impact of different nitrogen rates on total flavonoids of *Ipomea aquatica*. Mean with the same letter indicates that all of the groups were not significantly different according to Duncan multiple range test ($P \geq 0.05$) $N=4$.

4. CONCLUSION

420
 421
 422
 423
 424
 425
 426
 427
 428

In this work, four levels of nitrogen rates (0, 30, 60 and 90 kg/ha) was applied to *I. aquatica* to assess the growth, leaf gas exchange and production of secondary metabolites characteristics. It was found that as the nitrogen rates increased, the growth and leaf gas exchange properties of *I. aquatica* was enhanced. However, the production of phenolics and flavonoids of kangkung was reduced with high levels of nitrogen application as both total phenolics and flavonoid reached highest content at 30 kg N /ha. This work gives support that high nitrogen fertilization to *I. aquatica* can reduces the production of secondary metabolites although the growth parameters was enhanced with high nitrogen fertilization.

REFERENCES

429
 430
 431
 432
 433
 434
 435
 436
 437
 438
 439
 440
 441

1. Malaysia Department of Agriculture. Ringkasan Maklumat Perangkaan Agromakanan. www.moa.gov.my.2011
2. Ismail A, Marjan ZM and Foong CW. Total antioxidant activity and phenolic contents in selected vegetables. Food Chem. 2004;87: 581–6.
3. Edie EE and Ho BWC. Ipomea aquatica as a vegetable crop in Hong Kong. Econ. Bot. 1969; 23:32–6.
4. Nashrin S, Farooque A, Siddiqua M, Rahman M, and Khanam M. Effect of nitrogen and spacing on the growth and yield of gimakalmi, Ipomea reptans poir. J. Biol. Sci. 2002;76: 170-174.
5. Candlish I K, Gourley L and Lee HP. Dietary fiber and starch contents of some Southeast Asian vegetables. J. Agri. Food Chem. 1987; 35: 319–21.

- 442 6. Susila A, Prasetyo T, and Palada M. Optimum Fertilizer rate for kangkong (*Ipomoea reptans*
443 L.) Production in Ultisols Nanggung.2008;34:12-34.
- 444 7. Ayodele O, Alabi E, and Aluko M. Nitrogen fertilizer effect on growth, yield, and chemical
445 composition of hot pepper (rodo). *Int. Agri. Crop Sci.* 2015;8(5): 666-673.
- 446 8. Li J, Zhu Z, and Gerendas J. Effects of nitrogen and sulfur on total phenolics and antioxidant
447 activity in two genotypes of leaf mustard. *J. Plant Nutr.* 2008; 31: 1642-1655.
- 448 9. Mokhele B, Zhan X, Yang G, and Zhang X. Nitrogen assimilation in crop plants and its
449 affecting factors. *Canadian Journal of Plant Science.* 2011;17: 231-245.
- 450 10. Zaman MS, Hashem MA, Jahiruddin M, and Rahim MA. Effect of nitrogen for yield
451 maximization of garlic in old Brahmaputra flood plain soil. *Bangla. J. Agri. Res.*2011; 36(2):
452 357-367.
- 453 11. Mansour MMF. Nitrogen containing compounds and adaptation of plants to salinity stress.
454 *Biol. Plantarum.*2000; 43: 491–500.
- 455 12. Grusak MA, Della PD and Welch RM. Physiological process affecting the content and
456 distribution of phytonutrients in plants. *Nut. Rev.* 1999; 57(9): S27–S33.
- 457 13. Campostrini E, Yamanishi O, and Martinez E. Leaf Gas Exchange Characteristics of Four
458 Papaya Genotypes During Different Stages of Development. *Revista Brasileira de*
459 *Fruticultura.* 2001; 23(3): 522-525.
- 460 14. Hajiboland R and Beiramzadeh N. Growth, gas exchange and function of antioxidant defense
461 system in two contrasting rice genotypes under Zn and Fe deficiency and hypoxia. *Acta Biol.*
462 *Szegediensis* 2008; 52: 283–294.
- 463 15. Stiller V, Lafitte HR, Sperry JS. Hydraulic properties of rice and the response of gas exchange
464 to water stress. *Plant Physiol.* 2003; 132: 1698–1706.
- 465 16. Dingkuhn M; Cruz RT, O'Toole JC, Turner NC, Doerffling K. Responses of seven diverse rice
466 cultivars to water deficits. III. Accumulation of abscisic acid and proline in relation to leaf
467 water-potential and osmotic adjustment. *Field Crops Res.* 1991; 27: 103–117.
- 468 17. Dixon RA and Paiva NL. Stress-induced phenylpropanoid metabolism. *Plant Cell.* 1995; 7: 1
469 085–1097.
- 470 18. Due BM, Humphries D, Mai LTB, Dao AH, Co TM, Nga HH and Kim PT. Iron and vitamin C
471 content of commonly consumed foods in Vietnam. *Asia-Pacific Journal of Clinical Nutrition.*
472 1999; 8: 36–38.
- 473 19. Ibrahim MH, Jaafar HZE, Rahmant A, and Rahman Z. Effects of Nitrogen fertilization on
474 synthesis of primary and secondary metabolites in three varieties of kacang Fatimah (*Labisia*
475 *pumila blume*). *Int. J. Mol Sci.* .2011; 12(8): 5238-5254.
- 477 20. Taheri E, Soleymani A, and Javanmard HR. Effect of different levels of nitrogen on
478 morphological traits of two cultivars of rapeseed in isfahan region. *Int. J. Agri Crop Sci.*
479 2012;35: 1587-1590.
- 480 21. Amin MEM. Effect of different nitrogen sources on growth, yield and quality of fodder maize
481 (*Zea mays L.*). *J. Saudi Soc. Agri. Sci.* 2010; 67: 17-23.
- 482 22. Sarkar R, Jana J and Datta, S. Effect of cutting frequencies and nitrogen levels yield and
483 quality of water spinach (*Ipomoea reptans Poir.*). *Journal of Applied and Natural Science,*
484 2014;76: 545-551.
485
486
487

- 488 23. Phimmasan H, Kongvongxay S, Chhayty P and Preston T R. Water spinach (*Ipomoea*
489 *aquatica*) and Stylo 184 (*Stylosanthesguianensis* CIAT 184) as basal diets for growing rabbits.
490 *Livestock Res. for Rural Development*. 2004;16: 46–59.
491
- 492 24. Hare PD, Cress WA and Staden VJ. Proline synthesis and degradation: a model system for
493 elucidating stress-related signal transduction. *J. Exp. Bot.* 1999; 50: 413–34.
494
- 495 25. Prasad NK, Divakar S, Shivamurthy GR and Aradhya SM. Isolation of a free radical
496 scavenging antioxidant from water spinach (*Ipomoea aquatic* Forsk.). *J. Sci. Food Agri.* 2005;
497 85: 1 461–8.
498
- 499 26. Osman H, and Abu HA. Effect of NPK Fertilization on growth and dry matter accumulation in
500 mangrove [*Avicennia marina* (Forssk) vierh] Grown in Western Saudi Arabia. *Meteorol.*
501 *Environ. Arid Land Agri.*2010;56: 57-70.
502
- 503 27. Koo HM and Suhaila M. Flavonoid (Myricetin, quercetin, kaempferol, luteolin and apegenin)
504 content of edible tropical plants. *J. Agri. Food Chem.* 2011; 49: 3 106– 12.
505
- 506 28. Bojovic B and Markoviae A. Correlation Between Nitrogen and Chlorophyll Content In Wheat
507 (*Triticum aestivum* L.). *Kragujevac J. Sci.*2009; 3: 69-74.
508
- 509 29. Moreno N, Barrios A, Leal R, Franco A, Rodriguez A, and Hernandez L. Effect of Nitrogen
510 Deficiency and Toxicity in Two Varieties of Tomato (*Lycopersicum esculentum* L.). *Agri.*
511 *Sci.*2014; 5: 1361-1368.
512
- 513 30. Mansour MMF. Nitrogen containing compounds and adaptation of plants to salinity stress.
514 *Biologia Plantarum.* 2000; 43: 491–500.
515
- 516 31. Boussadia K, Steppe K, Labeke MC, Lemeur R and Braham M. Effects of Nitrogen deficiency
517 on Leaf Chlorophyll Fluorescence Parameters in Two Olive Tree Cultivars Meski' and
518 'Koroneiki'. *J. Plant Nutr.*2015;76; 2230-2246.
519
- 520 32. Baque MA, Karim MA, Hamid A, Tetsushi H. Effects of fertilizer potassium on growth, yield
521 nutrient uptake of wheat (*Triticum aestivum*) under water stress conditions. *South Pac.*
522 *Studies* 2006; 27: 29–35.
523
- 524 33. Grzebisz W, Gransee A, Szczepaniak W, Diatta J. The effects of potassium fertilization on
525 water-use efficiency in crop plants. *J. Plant Nutr. Soil Sci.* 2013: 176: 355–374.
526
- 527 34. Makino, A. Photosynthesis, Grain Yield, and Nitrogen Utilization in Rice and Wheat. *Plant*
528 *Physiology.* 2011; 155: 125-129.
529
- 530 35. Pankovica D, Plesnicar M, Arsenijevicâ MI, Petrovica N, Sakaci Z, and Kastori R. Effects Of
531 Nitrogen nutrition on photosynthesis in cd-treated sunflower plants. *Annals of Bot.*,2000; 86:
532 841-847.
533
- 534 36. Nori M, Bayat F, and Esmaeili A. Changes of vegetative growth indices and yield of garlic
535 (*Allium sativum* L.) in different sources and levels of nitrogen fertilizer. *Int. J. Agri. Crop*
536 *Sci.*2012;67; 1394-1400.
537
- 538 37. Schulze ED, Kelliher F, Korner C, Lloyd J, and Leuning R. Relationships among maximum
539 stomatal conductance, ecosystem surface conductance, carbon assimilation rate, and plant
540 nitrogen nutrition: a global ecology scaling exercise. *Ann. Rev. of Ecol. Systematics.*1994; 25:
541 629-660.
542
- 543 38. Stewart W. Balanced fertilization increases water use efficiency. Atlanta, Georgia: Potash &
544 Phosphate Institute (PPI) and Potash & Phosphate Institute of Canada (PPIC).2001.
545
- 546 39. Artur A, Garcez T, and Monteiro F. Water use efficiency of marandu palisadegrass as
547 affected by nitrogen and sulphur rates. *Artigo Cientifico.* 2014; 45(1): 10-17.

- 548
549
550
551
552
553
554
555
556
557
558
559
560
561
562
563
564
565
566
567
568
569
570
40. Naguib, AM. Enhancement of phenolics, flavonoids and glucosinolates of Broccoli (*Brassica oleracea*, var. *Italica*) as antioxidants in response to organic and bio-organic fertilizers. *Journal of the Saudi Society of Agricultural Sciences*.2013;78: 135-142.
 41. Stewart A, Chapman W, Jenkins G, Graham I, Martin T, and Crozier A. The effect of nitrogen and phosphorus deficiency on flavonol accumulation in plant tissues. *Plant Cell Environ*.2011; 24: 1189-1197.
 42. Orphanides A, Goulas V and Gekas V. Effect of drying method on the phenolic content and antioxidant capacity of spearmint. *Czech J. Food Sci*.2013; 31(5): 509-513.
 43. Palumbo, M., Putz, F., & Talcott, S. Nitrogen fertilizer and gender effects on the secondary metabolism of yaupon, a caffeine-containing north American holly. *Oecologica*.2006; 151(1): 1-9 .
 44. Meyer S, Cerovic ZG, Goulas Y, Montpied P, Demotes S, Bidet LPR, Moya I and Dreyer E. Relationship between assessed polyphenols and chlorophyll contents and leaf mass per area ratio in woody plants. *Plant Cell Environ*. 2006;29: 1338-1348.
 45. Margna U, Margna E and Vainjarv T. Influence of nitrogen nutrition on the utilization of L-phenylalanine for building flavonoids in buckwheat seedling tissue. *J. Plant Physiol*. 1989;134:697-702.