ABSTRACT

Objectives: To investigate the effect of different mixtures from nano casein phosphopeptides (CPP), amorphous calcium phosphate (ACP), probiotic Lactobacillus rhamnosus B-445 (L. rhamnosus) and casein glycomacropeptide (GMP) against Streptococcus mutans (S. mutans) growth and its adhesion.

Methods: CPP was prepared from the tryptic digest of bovine casein and GMP by the action of chymosin on casein solution. Four mixtures namely: 1)Nano CPP ;2)Nano CPP+ ACP ;3)Nano CPP+ ACP + L. rhamnosus; and 4)Nano CPP+ ACP+ GMP were prepared and tested for its inhibitory activity against S. mutans growth and its adhesion to saliva-treated glass surfaces in comparison with a commercial product (GC MI paste plus) and chlorhexidine (0.2%) as a positive control. The particle size and zeta potential of nano CPP and its complex with ACP were evaluated. Furthermore, the viability of L. rhamnosus in its mixture was determined during two weeks of storage at pH 6.8 and 8 respectively.

Results: revealed that Nano CPP had an average particle size (7.75 nm) and zeta potential (-8.43 mV) lower than that of CPP+ACP mixture. Probiotic containing mixture exhibited inhibitory activity slightly less than the positive control at pH 6.8. All tested mixtures reduced the adhesion of S. mutans to saliva-treated glass surfaces and the highest was that containing probiotic and GMP. L. rhamnosus showed acceptable stability in CPP+ACP mixture during storage period. Conclusions: All these findings suggest the use of probiotic, CPP+AMP mixture as a dental anticariogenic and remineralizing agents.

Keywords: Probiotics, Oral health, Casein phosphopeptides, Casein glycomacropeptide, Adhesion, Inhibitory activity, S. mutans.

1. INTRODUCTION

Dental caries is one of the most prevalent and expensive human disease of bacterial origin to treat all over the world [1]. It is well documented that the progress of a caries lesion is known as a disproportion in the demineralization and remineralization cycle [2]. Demineralization arises from the action of the organic acids that result from the fermentation of dietary sugar by Streptococcus mutans (S. mutans), the dental plaque odontopathogenic bacteria [3]. Dental plaque is a matrix-enclosed microbial community that develops on the tooth surface, whereas a large number of bacteria are involved and there is fierce competition for nutrients and attachment sites [4]. Several strategies have been proposed to prevent the incidence of dental caries based on the mediation of the growth and activity of odontopathogenic bacteria and/or increased tooth remineralization.
Recent reviews have reported the use of probiotic strains for the prevention of oral diseases, including caries [5,6,7]. Probiotics are administered to maintain or restore the natural saprophytic micro-flora against a pathogen invasion, which is central to the development of the major oral diseases (caries and periodontal disease). The beneficial action of probiotics is relied on their interaction with other bacteria in the plaque and hampers growth by pathogens through production of hydrogen peroxide, bacteriocins, and organic acids [8]. The effect of probiotics on dental caries and its related risk factors has been evaluated in several experimental studies, using different strains; *L. rhamnosus GG*, *L. casei*, *L. reuteri*, *L. plantarum*, *L. brevis* CD2, *Bifidobacterium* spp [5]. These previous studies have demonstrated the capacity of probiotic to reduce *S. mutans* counts in saliva and/or plaque in short-term but randomized clinical trials on this topic were insufficient to provide scientific clinical evidence [5].

In the late 1950s Dairy products have been considered as effective food in preventing dental caries [9]. Phosphorus rich peptides (CPP) which is isolated from tryptic digest of bovine milk casein (αs1-casein, β-casein and αs2-casein), could stabilize calcium ions at wide range of pH (5.0-9.0) in a direct proportional relationship [1,10]. Amorphous calcium phosphate (ACP) is the initial solid phase that precipitates from a highly supersaturated calcium phosphate solution. In vivo, ACP is a good osteoconductivity, biodegradability and bioactivity but without cytotoxicity. However, unstabilized ACP undergoes rapid transformation to crystalline phases in the mouth where it may attribute to dental calculus [11]. Therefore, there is a need to stabilize ACP to retain its favorable properties. CPP was found bind to nanoclusters of ACP in supersaturated solutions, preventing the growth of ACP to the critical size required for phase transformation and precipitation. CPP–ACP nanoclusters work effectively as a remineralizing agent at wide range of pH [13]. Furthermore, CPP–ACP has the ability to inhibit cariogenic streptococci to the tooth surface inducing the formation of noncariogenic plaque [13]. Thus, the anticariogenic potency of CPP-ACP may act to promote its incorporation into food products and dental products as a novel tool in the fight against dental caries.

Casein glycomacropeptide (GMP) is a blend of glycosylated and non-glycosylated peptides of the same peptide chain which originates from the influence of chymosin on κ-casein [14]. Because of the great antimicrobial effects of GMP including inhibitory adhesion of oral Actinomyces and Streptococci to surfaces, it was suggesting to be as inhibitor against dental caries [15]. Moreover, GMP exhibited a superior tooth protection against erosion than CPP at pH 3 while CPP exhibited better performance at pH 4.5 [16].

Previous studies reported that one of aforementioned agent could prevent dental caries; however, there is no reference has been cited on the use of combined agents. Hence, the aim of this study was to assess the effect of different combinations of probiotics, CPP-ACP, and GMP as anti-cariogenic or/and remineralizing agents against *S. mutans* growth and its adhesion on saliva-coated glass surfaces simulating the pellicle-covered enamel surfaces.

### 2. MATERIAL AND METHODS

#### 1.1. Separation of casein phosphopeptide (CPP).

Bovine casein solution (1%) in phosphate buffer (pH 8 0.1M) was hydrolyzed with trypsin (1:100, enzyme: protein ratio) at 45°C/5 hrs. Then, pH was adjusted to 4.6 and finally centrifuged at 1500 g/15 min. Calcium chloride (0.2%) was added to the clear supernatant followed by ethanol to precipitate CPP and then lyophilized [17].

#### 1.2. Determination of the purity of CPP by HPLC.

HPLC analyses of CPP were done using high purity water (Millipore, Bedford, MA, USA), methanol and acetonitrile (Redel and Fluka, Germany), also utilizing Trifluoroacetic acid (TFA; Merck, Dermo, Germany). Aliquot of a filtrate containing (0.1 g) of the lyophilized CPP with 10 mL of 0.1% (V/V) TFA was injected into the HPLC system. Chromatographic analysis was carried out using HPLC Agilent 1100 integrated system equipped with a G1313A automated injector, a G1311A pump and G1315B multi wavelength diode-array detector (DAD) operated at 220 nm. The separation was carried out at 20°C and flow rate of 0.5 mL/min. using linear gradient from 0.1% (V/V) TFA acid to a solution containing 0.1% and 80% acetonitrile (V/V). The chromatographic data was analyzed using Agilent Chemstation.

#### 1.3. Measurement of the phosphorus content of CPP.

This was conducted by following the standard method [AOAC, 1990] [18] and using phosphomolybdenic acid.

#### 2. Preparation of casein glycomicropeptide (GMP)
Bovine casein was dissolved at pH 8 to make 1% solution with the aid of 0.1% NaOH, then adjusted to pH 6.6 and finally, 0.2% CaCl2 was added. Chymosin (Ch. Hansen Lab, Denmark) was added at the ratio of 0.3g/l casein solution and incubated at 40ºC until the formation of a coagulum. The reaction mixture was heated to 70ºC to inactivate the enzyme, coagulum was broken and the curd was separated by filtration, then finally freeze dried [14].

3. *Lactobacillus rhamnosus* (*L. rhamnosus*) culture and enumeration

Probiotic *L. rhamnosus B-445* was provided in lyophilized form by the Northern Regional Research Laboratory, Illinois, USA. Bacterial activation was carried out through inoculation in MRS medium broth and anaerobic incubation (Gas Generating Kit Anaerobic System, Oxoid, UK) at 37°C for 48 hrs. Then cultured bacteria were centrifuged at 5000 rpm/20 min to obtain pure cell pellet. The number of the live bacteria cells in one gram of the previously obtained pellet was determined by colony counting method and the total live cell numbers per one gram of pellet were calculated using the following formula; Live cells (CFUs/g) = number of colonies x dilution factor [19].

4. Preparation of different experimental mixtures

Different combinations from Nano (CPP), amorphous calcium phosphate (ACP), *L. rhamnosus* and (GMP) were prepared to form four groups as follows; Group (1): Nano CPP (5.0 g) was dissolved and made up to 100 ml with phosphate buffer at pH 6.8 and 8, Group (2): Nano CPP (2.0 g) and (0.375 g) nano ACP (<150 nm particle size , Sigma-Aldrich, USA) were dissolved and made up to 100 ml with phosphate buffer at pH 6.8 and 8, Group (3): Nano CPP (2.0 g) and nano ACP (0.375 g) were dissolved followed by inoculation of 10⁶ CFU/g of *L. rhamnosus* B-445, and then were made up to 100 ml phosphate buffer at pH 6.8 and 8, Group(4): nano CPP (2.0 g), nano ACP(0.375 g) and GMP (2.0 g) were dissolved and made up to 100 ml phosphate buffer at pH 6.8 and 8, Group(5): Commercial products (GC MI paste plus; Contains 0.2% (900ppm) Fluoride and Recaldent (cpp-acp)), Group (6) (Positive control): % 0.2 Chlorhexidine diacetate (Sigma-Aldrich, USA, Product Number C 6143).

5. Determination of paricle size and zeta potential

The size and zeta potential of the droplets in the prepared CPP containing particles were evaluated by Zetasizer var. 704 instrument (Malvern Instruments, Malvern, UK). Sample was diluted with MQ water before measurement its light scattering for a laser beam (633 nm) at an angle of 173 at 25°C over time intervals. The changes in laser beam scattering versus time was used to determine the particle size distribution and polydispersity index (PDI) were calculated from the particle size distribution. Zeta potential was measured in the same sample by electrophorsis and results were expressed as mV. and (PDI) [20].

6. Microbiological assessment

*S. mutans* was isolated from the dental plaque of high caries index patients by swabbing method according to Sanchez et al [21]. Selective media Mitis salivarius-bacitracin agar (MSB; BD Difco, Paris) was used to isolate and grow S. mutans (Dairy microbiological Lab., National Research Center, Giza, Egypt). A single colony of S. mutans was isolated and placed in a test tube containing 10 ml of Treptone Soya broth (TSB: Difco laboratories, Detroit, MI, USA) and incubated at a temperature 37°C for 24 hrs. The cultures were then diluted with fresh media until a concentration 10⁶ CFU/ml was obtained and used as working microbial solution [22].


The antimicrobial activity of eighty-four specimens of the previously prepared four experimental groups were divided into twenty-eight specimens at pH6.8 and other twenty-eight at pH 8 (n=7) as well the fifth commercial group (n=14) and the sixth positive group (n=14) was evaluated on the profile growth of *S. mutans*. Nutrient agar medium was poured into sterile petri dishes (15ml each) and 20μl of the formerly made microbial solution was spread over each agar plate and allowed to dry for 5 min. A sterile 6 mm cork-borer was used to cut six wells at equidistance in the plate and the first four wells, each was filled with 50μl of each four experimental groups, while the fifth well with 50μl of commercial product and the sixth well with 50μl of positive group. The agar plates were incubated at 37°C for 24 hrs and the inhibition zones around the wells were measured [23].

6.2. Survival profile of probiotic strain in the experimental Group (3).

The viability of *L. rhamnosus B-445* in its mixture at pH6.8 and pH8 of Group (3) was determined at different storage period (after 24hrs, 72hrs, and 2 weeks). Live bacterial cells in one ml of mixture were enumerated using colony counting method as aforementioned [19].
6.3. Adherence inhibition assay

The previously prepared microbial solution was poured onto anaerobic agar plates and incubated at 37°C for 18-20 hrs. The colonies were then harvested and dispersed into 30 ml Brain Heart Infusion (BHI, Oxoid) broth containing 5% (w/v) sucrose. The turbidity of the suspension was adjusted spectrophotometrically at 550 nm to an absorbance of about 0.144. At this absorbance, the concentration of cell is standardized to about 1 CFU/ml for use in the anti-adherence study. Also, whole saliva (WS) was collected from a single donor by expectoration after chewing a piece of rubber band, and then clarified by centrifugation (17,000 ×g, 30 min) [24].

6.3.1. Determination of the adherence affinities of S. mutans on the saliva-coated glass surfaces.

Saliva-coated glass culture tubes were prepared by treating its inner surface with 2 ml of clarified saliva for two min. Then the surface was rinsed with sterile distilled water and the rinse was used as control. After that, 2 ml of bacterial suspension was added to saliva-coated glass culture tubes incubated for 18-20 hrs at 37°C. The tube contents were removed and its turbidity (A) was measured spectrophotometrically at 550 nm. The emptied tube was then washed with 2 ml of distilled water, suspension was collected and its turbidity (B) was measured at 550 nm. The sum of A and B turbidity (Aub) corresponds to the amount of the total unbound cells. The concentration of the adherent S.mutans was determined by adding 2 ml of distilled water into the emptied tube, sonicated for 10 sec and the turbidity (Ab) of the suspension was measured at 550 nm [25].

6.3.2. Determination of the anti-adherence effect of different groups on S.mutans attached to saliva-treated glass surfaces.

Two ml of each four experimental groups and the fifth commercial group was added into previously treated saliva-coated culture tubes and left for two min. The inner surfaces were then rinsed with sterile distilled water and the procedure was repeated as above described. The percentage of adherence was calculated as follows:

\[ \text{Adherence} \% = \frac{\text{Ab}}{\text{Ab} + \text{Aub}} \times 100 \]

Where, Ab and Aub are the absorbance of bound and unbound cells at 550 nm respectively.

The percentage of adherence reduction of cells in the presence of experimental group or commercial group was calculated as follows:

\[ \text{Adherence reduction} \% = \frac{\text{Ab} - \text{Abt}}{\text{Ab}} \times 100 \]

Where Ab, Abt are the absorbance of bound cells without and with treated groups respectively at 550 nm.

7. Statistical analysis

Data were explored for normality using Kolmogorov-Smirnov and Shapiro-Wilk tests and showed parametric distribution. For parametric data; One-way ANOVA followed by tukey post hoc test was used for comparison among the groups in non-related samples. Repeated measure ANOVA and paired sample t-test was used for comparison among the related samples. The significance level was set at \( P \leq 0.05 \). Statistical analysis was performed with IBM® SPSS® Statistics Version 20 for Windows.

3. RESULTS

Regarding preparation and purity of Casein phosphopeptide (CPP); the phosphorus content of the prepared CPP was found to be 2.32%, and HPLC separation of the prepared CPP demonstrated the presence of major and minor peaks respectively (Figure 1).

For the Particle size of CPP nanoparticles in Group 1 was 7.75 nm ± 1.506 nm St Dev. and 0.561 for (PDI). While 90% of particles number in Group (2) with average size of 109.7 nm ± 25.66 nm St Dev and 10% of the total particle had an average size of 509.5 ± 122.4 nm StDev. In Group (4) 100% of particles number had an average size 199.6±156.9 nm and 0.471 for PDI (Figure 2). For the Zeta potential values of CPP, Group (1) followed normal distribution curve with -8.43 ±13.0 mV and Group (2) was revealed an average Zeta potential of -6.5 ± 8.73 mV. While low average -5.40 ±29.3 mV was observed in Group (4) (Figure 3).

In (Table 1), the antibacterial efficacy of CPP containing solutions at pH 8 against S. mutans growth showed a significant difference between (Group 6; positive control) and all other groups (\( P \leq 0.001 \)), while there were no inhibition zones in (Group 1), (Group 2), (Group 3), (Group 4) and (Group 5) at lower pH value of 6.8, inhibition zones value was
found in (Group 6) (16.00 ± 0.00) followed by (Group 3) (10.29 ± 3.55) (Ps.001), where no inhibition zones were found in (Group 1), (Group 2), (Group 4) and (Group 5) (0.00 ± 0.00) (Table 1).

For the survival profile of *L. rhamnosus* in its mixture of Group (3) at pH 8, the results revealed that the highest survival mean value was after 72 hours storage period (231.25x10^7 ± 13.15x10^7) followed by after 2 weeks (216.75x10^7 ± 11.35x10^7), then after 24 hours (179.00x10^7 ± 8.60x10^7), while the least mean value was in Base line (156.50x10^7 ± 22.34x10^7) (Table 2). On contrary, the probiotic survival in mixture at pH 6.8 showed highest mean values in Base line (129.50x10^7 ± 12.37x10^7) followed by after 24 hours (82.25x10^7 ± 9.03x10^7) and 72 hours (73.25x10^7 ± 10.90x10^7), while the least mean value was found after 2 weeks (73.00x10^7 ± 7.02x10^7) (Table 2).

When comparing the mean % of cell adherence values for different experimental solutions and commercial one, the highest mean value of % of cell adherence was found in (Control) (33.35% ± 0.82) followed by (Group 2) (23.00% ± 0.82), (Group 1) (18.60% ± 1.63), (Commercial) (17.50% ± 0.82) and (Group 4) (15.00% ± 0.83) while least % of cell adherence was found in (Group 3) (14.80% ± 0.79) (Figure 4). Accordingly, the highest mean value of % of reduced adherence was found in (Group 3) (65.00% ± 2.06) followed by (Group 4) (64.00% ± 3.27), (Commercial) (57.00% ± 0.82), (Group 1) (55.70% ± 0.82) and (Group 2) (42.00% ± 1.63) while least % of reduced adherence was found in (Control) (33.35% ± 0.82).

### 4. DISCUSSION

Casein phosphopeptide (CPP) is a mixture of peptides containing clusters of phosphate groups and emerged from the action of proteolytic enzymes on the different casein fraction. In the present study, Trypsin was chosen to hydrolyze bovine casein because of its high specificity and its capability to release phosphopeptides from casein [17]. As it was reported in the literature that several phosphopeptides can be released from the different casein fraction and precipitated together by the applied procedure [17]. Therefore, in this study, the phosphorus content of the prepared CPP was found to be 2.32% and indicated that almost the phosphorus content was concentrated in the prepared CPP. Regarding HPLC separation of the prepared CPP (Figure 1), the results revealed the presence of major and minor peaks respectively indicating the high purity of the prepared CPP.

Solutions containing CPP nanoparticles were examined for their particle sizes and the results are presented in CPP (Figure 2). Previous study by Fitz [17] was demonstrated that CPP are small molecular weight peptides composed of 20-25 amino acid residues, and this latter observation explain the low molecular size of CPP particles in Group (1) (7.75 nm ± 1.506 nm) and high homogeneity of the tested CPP molecules (Figure 2. A). For Group (2) the recorded particle size of the major number (109.7 nm ± 25.66 nm) was less than in the commercial ACP (150 nm) which was incorporated into the CPP-ACP solution Group (2). The previous results revealed that the interaction of CPP with ACP could be attributed in a complex of nano size particles with small amount of larger aggregates that apparent in 10% of the total particle number in Group (2) (509.5 ± 122.4 nm) (Figure 2. B). Concerning Group (4) where GMP incorporated into CPP+ACP solution, a significant change in the particle size distribution pattern were observed in (Figure 2. C) where a broad shoulder in this distribution curve indicating the presence of large aggregates. Comparing results between Group (2) and Group (4) indicated that the addition of GMP resulted in significant increase in major number of particle size due to possible aggregation of the three components in the solution.

For Zeta potential measurement of CPP in Group (1) (-8.43 ±13.0 mV) and Group (2) (-6.5 ± 8.73 mV) (Figure 3. A, B), the results could be explained by Reynolds [10] who demonstrated that CPP to bind ACP forming clusters in metastable solution preventing their growth to the critical size required for their flocculation and precipitation. Furthermore, Uskoković et al [26] reported that the increase of HPO4groups to Ca ions could increase the negative zeta potential of the solution. While for Group 4, the addition of GMP to the CPP+ACP solution resulted in wide variation of the zeta potential of the solution with low average of -5.40 ±29.3 mV (Figure 3. C). This latter result could be related to the previously observation with variation in its particle size (Figure 2. C).

For antimicrobial activity of four experimental groups at pH 8.0, the results exhibited no inhibitory activity against *S. mutans* except the positive control which had an inhibitory zone of 14.43 ± 3.41 mm). While at lower pH value of 6.8, the results revealed that both Group 3 and Group 6 exhibited clear inhibitory zones against *S. mutans* (Table 1). The effect of pH on the growth and activity of probiotic bacteria has been documented by Rault et al [27] and Lambert [28] whereas most bacteria exhibit maximum activity at pH 6-7. Accordingly, the present results suggest that pH plays an important role in the inhibitory activity of the probiotic containing solution in Group 3. The previous observation is in agreement with Deepika et al [29] who demonstrated that increasing the pH during fermentation from 5 to 6 was found to change the surface composition of *L. rhamnosus* whereas it increased its hydrophobicity and decreased its adhesion to caco-2-cells. These latter results support the pH at 6.8 is best for the inclusion of probiotic *L. rhamnosus* in oral health preparations as a natural therapeutic mean to control oral pathogen *S. mutans* while pH 8.0 is inappropriate for this purpose.
Concerning the survival profile of *L. rhamnosus* in its mixture Group 3, the count of *L. rhamnosus* stored at pH 6.8 decreased rapidly after 24 hours losing about 37% of its original count and then slightly decreased to reach \((73.00 \times 10^7 \pm 7.02 \times 10^7)\) after 2 weeks and this indicates reasonable stability under the tested conditions (Table 2). While *L. rhamnosus* storage at pH 8.0 showed an increase in its viable count up to 72 hours and then decreased slightly. The observed increase in latter result may be attributed to possible disaggregation of cell aggregates due to the hydroxyl ion in the medium.

The anti-adherence activities of CPP containing solutions against *S. mutans* attached to saliva-treated glass surface at pH 6.8 showed that, all tested experimental solutions and the commercial product exhibited variable degrees of anti-adherence activities against attached *S. mutans* but with variable degrees. For Group1 the adherence of the pathogen was reduced by \((55.70\% \pm 0.82)\) which being close to the reduction percentage exhibited by the commercial product Group5 \((57.00\% \pm 0.82)\). Both the solutions that containing the probiotic Group 3 and GMP Group 4 exhibited high adherence reduction percentages of \((65.00\% \pm 2.06)\) followed by \((64.00\% \pm 3.27)\) respectively. This is in agreement with previous study conducted by Marttinen et al [30] who demonstrated that Lactobacilli could reduce streptococcal adhesion in particular on saliva-coated hydroxyapatite. Also, it has been reported in the literature by Neeser [15] that GMP reduces the adhesion of toxins which may explain the present results for Group (4).

![HPLC chromatogram of prepared CPP](image)

Fig. 1. HPLC chromatogram of prepared CPP
Fig. 2. Particle size distribution of (A) CPP solution (B) CPP+ACP solution and (C) CPP+ACP+GMP solution by number.
Table 1. Mean and standard deviation (means ± SD) values of the inhibition zones diameter (mm) were exhibited by different groups against *streptococcus mutans* growth at pH 8.0 and pH 6.8.

<table>
<thead>
<tr>
<th>Constituent in solution</th>
<th>pH 8.0</th>
<th>pH 6.8</th>
</tr>
</thead>
<tbody>
<tr>
<td>CPP (G1)</td>
<td>0.00</td>
<td>0.00</td>
</tr>
<tr>
<td>CPP+ACP (G2)</td>
<td>0.00</td>
<td>0.00</td>
</tr>
<tr>
<td>CPP+ACP+Probiotic (G3)</td>
<td>0.00</td>
<td>10.3±3.6&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>CPP+ACP+GMP (G4)</td>
<td>0.00</td>
<td>0.00</td>
</tr>
<tr>
<td>Commercial inhibitor (G5)</td>
<td>0.00</td>
<td>0.00</td>
</tr>
<tr>
<td>Chlorhexidine diacetate (G6)</td>
<td>14.4</td>
<td>16.0±2.3&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>ab</sup> Different letters in the same column indicate statistically significant difference at P < .05

Table 2. Mean Survival of *L. rhamnosus* (CFU/ml) in the solution containing CPP + ACP at different storage periods at pH 8.0 and pH 6.8

<table>
<thead>
<tr>
<th>Storage time</th>
<th>pH 8.0</th>
<th>pH 6.8</th>
</tr>
</thead>
<tbody>
<tr>
<td>Base line</td>
<td>156.50±22.34&lt;sup&gt;a&lt;/sup&gt;</td>
<td>129.50±12.37&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>24 h</td>
<td>179.00±8.60&lt;sup&gt;a&lt;/sup&gt;</td>
<td>82.25±9.03&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>72 h</td>
<td>231.25±13.15&lt;sup&gt;b&lt;/sup&gt;</td>
<td>73.25±10.90&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>2 wk</td>
<td>216.75±11.35&lt;sup&gt;b&lt;/sup&gt;</td>
<td>73.00±7.02&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>P value</sup><sup>• Values are given as (mean ± SD) x10<sup>7</sup></sup>

<sup>• Different letters in the same column indicate statistically significant difference</sup>

<sup>*; significant (P<.05)</sup>
4. CONCLUSION

The present *in vitro* study confirmed the anti-adhesive and antibacterial activities of the experimental solution containing mixture of probiotic *L. rhamnosus* in CPP-ACP against *S. mutans*. Moreover, *L. rhamnosus* showed high stability in this solution for a practical period of time study. Therefore, this mixture can be considered a promising ingredient in oral health preparations.

COMPETING INTERESTS

Authors have declared that no competing interests exist

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