

# **Incidence of Melodogyne Nematofauna in Some Selected Vegetable Crops in Nukkai Irrigation Field Jalingo, Taraba State, Nigeria.**

## **ABSTRACT**

A study to determine the incidence of root-knot nematode (*Melodogyne* spp) in some selected vegetables was conducted in Nukkai Irrigation Field of Jalingo, Nigeria. Samples of Okra (*Hibiscus esculentus*), Spinach (*Amaranthus* spp) and Sorrel (*Hibiscus sabdariffa*) were collected at 2-weeks old and at maturity (flowering stage). Their roots were cut off and nematodes (*Melodogyne* spp) were extracted using the Baermann method and identified using the female perineal pattern manual. The results showed that *Melodogyne* spp are incident in the study area as 248 (34.44%) stands out of the 720 stands studied were found to be infested by root-knot nematode. The results also showed that two *Melodogyne* spp, *M. javanica* and *M. incognita* were discovered, and had a total number of 535 individuals. *M. javanica* (345) were predominantly higher than *M. incognita* (190). There was no significant difference ( $\chi^2_{=0.05}$ ) between the number of root-knot nematode extracted from vegetables from the three different plots (plots A, B and C), but there is a significant difference ( $\chi^2_{=0.05}$ ) between the number of *M. javanica* recovered and *M. incognita*.

*Keywords:* *Amaranthus* , *Hibiscus* , Jalingo, *Melodogyne*, *Nigeria*, Nukkai.

## **INTRODUCTION**

Nematodes are soil dwelling organisms that constitute one of the largest animal phyla in the world, with over half a million species known. It has been estimated that four out of every five living animals on this planet are nematodes [1]. Nematodes can live as obligate parasites of plants and animals and can also alternate a parasitic life, with a free living life style or can be strictly free [2]. Despite the diversity in their life style and habitat, all nematodes are morphologically, anatomically and developmentally similar [3].

Root-knot nematodes (*Meloidogyne* spp) are minute worm-like animals that are very common in the soil and can invade roots of plants. They have a wide host range, and cause problems in many annual and perennial crops; they occur throughout the world infecting all major crops and causing substantial reduction in yield and quality [4]. There are four important species of the genus; *Meloidogyne arenaria*, *Meloidogyne hapla*, *Meloidogyne incognita* and *Meloidogyne javanica* which are considered as

32 the most economically important species responsible for causing more damage to agricultural produce [5,  
33 6].

34 Vegetables are much more important commodities needed in high demand due to their nutritive  
35 value for balanced nutrition in many areas of the world but because of root-knot nematode (*Meloidogyne*  
36 spp), it is increasingly very difficult and sometimes impossible to grow important vegetable in the tropics  
37 and semi tropical countries [2]. In Africa, the yield of vegetables is relatively lower, for which there are  
38 many constrains including prevalence of disease caused by different pathogens [7]. Among various  
39 pathogens responsible for the low yield, root-knot nematodes are of considerable economic importance  
40 [8] and can cause an annual loss of 22% in the tropics [9]. In addition, these parasites also interact with  
41 other disease causing organisms to produce a disease complex [10]; break down resistance against other  
42 pathogens and reduce plant tolerance to environmental stress [11,12].

43 It is against this background that this study was designed to determine the incidence of root-knot  
44 nematodes of some vegetables cultivated in Nukkai Irrigation Fields of Jalingo, Taraba State.

## 45 **MATERIALS AND METHODS**

### 46 *Study Area*

47 Nukkai Irrigation Field is an irrigation farming field in Jalingo, Taraba State, which has an area of  
48 about 20 hectares. Jalingo is located at latitudes 8<sup>0</sup>47' to 9<sup>0</sup> 01'N and longitude 11<sup>0</sup> 09' to 11<sup>0</sup>30'E. Nukkai  
49 Irrigation Field is located immediately after the Nukkai Bridge south of the Jalingo-Wukari main road in the  
50 city of Jalingo (Figure 1).

### 51 *Sample Collection*

52 All samples were collected using the method adopted by Anwar and McKenry [13]. Samples of  
53 Okra (*Hibiscus esculentus*), Spinach (*Amaranthus* spp) and Sorrel (*Hibiscus sabdariffa*) were randomly  
54 collected at different growth stages, i.e. seedling stage and matured stage. For each vegetable species,  
55 seedling was collected at random when they were aged 2-weeks old and the matured vegetables were  
56 also collected immediately they started flowering. All samples collected were washed and cleaned of soil  
57 debris. The roots were cut off and kept in the fridge in a plastic container.

### 58 *Extraction of Nematodes*

59 Nematodes were extracted from roots of plants using the Baermann funnel extraction method as  
60 described by Baermann [14]. In the set-up, a funnel was placed in a stand and filled with water until it  
61 reached up to 1cm below the rim. A rubber tube was fixed at the mouth of the funnel whose end was  
62 tightened using a clip. It was ensured that formation of air bubbles was avoided. The clip ensured it was  
63 well closed and the rubber tube did not leak. The sieves with the sample were hung in the funnel so that

64 the sample was totally submerged, without touching the bottom of the funnel. Nematodes crawled out of  
65 the sieve into the water and settled. After a period of 16-72 hours, the nematodes suspension was  
66 trapped by opening the squeezer clip, regularly tapping and adding water to increase nematodes vitality.

#### 67 *Identification and Counting of Nematodes*

68 Nematodes were identified using the female perineal pattern method manual or guide by [15].  
69 During identification, nematodes were counted directly under microscope using the counting dish.

#### 70 *Statistical analysis*

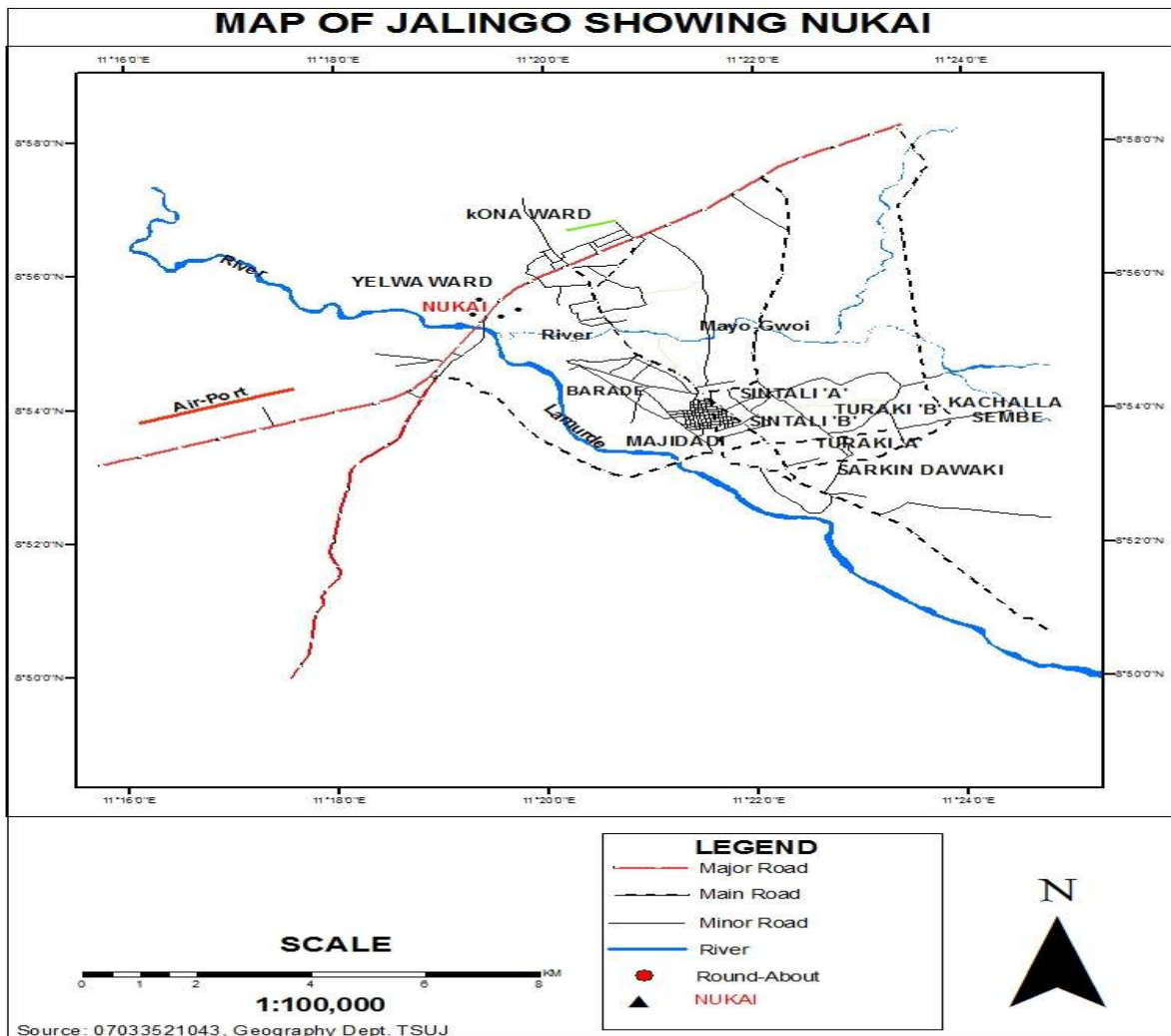
71 The collected data was analyzed using descriptive statistics of means, percentages and chi-  
72 square.

73

74

75

76



77  
78 **Figure1:** The Map of Jalingo showing the study area (Nukkai irrigation field).

79 **RESULTS**

80 *Incidence of Infestation*

81 The incidence of infestation of plant by root knot nematodes is shown in Table 1. The result  
82 revealed that plant parasitic nematodes are incident in the field of study as they are found in all the  
83 studied vegetable. Out of the 120 stands from the three (3) plots sampled, Okra (*Hibiscus esculentus*)  
84 had the highest infestation of 60 (50.0%), followed by Spinach (*Amaranthus spp*) which had a total  
85 infestation of 52 with an incidence of 43.3% while the least infestation (30) was recorded in Sorrel  
86 (*Hibiscus sabdariffa*) with a total incidence of 25.0%. Chi-square test also revealed that there was a  
87 significant difference ( $p>0.05$ ) between the total number of infested and uninfested stands studied (Table

88 1). Across all plots, plot A showed highest infestation, with an incidence rate of 42.5%, followed by plot B  
89 (40.0%), while the least infestation was observed in plot C (35.8%).

90 The incidence of infestation of matured stage of the vegetable plants by root knot nematodes is  
91 presented in Table 2. Matured Spinach (*Amaranthus* spp) had the highest total infestation of 54 stands  
92 out of the 120 stands from the three (3) plots sampled with an incidence rate of 45.0%, followed by Okra  
93 (*Hibiscus esculentus*) which had a total infestation of 51 with an incidence of 42.5% while the least  
94 infestation of 38 was recorded in Sorrel (*Hibiscus sabdariffa*) with a total incidence of 31.7%. Chi-square  
95 test showed that there is a significant difference ( $p>0.05$ ) between the total number of infested and  
96 uninfested stands of all the vegetable plants (Table 2). The results also revealed that plot A had the  
97 highest infestation rate (40.0%) followed by plot B (39.2%) while the least infestation was recorded in plot  
98 C (35.8%), as shown in Table 2.

99 **Table 1:** Incidence of Plant Parasitic Nematodes at Seedling Stage in Nukkai Irrigation Field, Jalingo (2016)

Plant species	Plot A		Plot B		Plot C		Total	
	+ve	-ve	+ve	-ve	+ve	-ve	+ve	-ve
<i>Amaranthus</i> spp	19(47.5%)	21(52.5%)	13(32.5%)	27(67.5%)	20(50.0%)	20(50.0%)	52(43.3%)	68(55.7%)
<i>Hibiscus esculentus</i>	20(50.0%)	20(50.0%)	25(62.5%)	15(37.5%)	15(37.5%)	25(62.5%)	60(50.0%)	60(50.0%)
<i>Hibiscus sabdariffa</i>	12(30.0%)	28(70.0%)	10(25.0%)	30(75.0%)	08(20.0%)	32(80.0%)	30(25.0%)	90(75.0%)
Total	51(42.5%)	69(57.5%)	48(40.0%)	72(60.0%)	43(35.8%)	87(64.2%)	142(39.4%)*	218(60.6%)*

100 Note: Values with asterisk are statistically significant ( $\chi^2_{=0.05}$ )

101

102 **Table 2:** Incidence of Plant Parasitic Nematodes at Maturated Stage in Nukkai Irrigation Field, Jalingo (2016)

Plant species	Plot A		Plot B		Plot C		Total	
	+ve	-ve	+ve	-ve	+ve	-ve	+ve	-ve
<i>Amaranthus</i> spp	20 (50.0%)	20 (50.0%)	19 (47.5%)	21 (52.5%)	15 (37.5%)	25 (62.5%)	54 (45.0%)	66 (55.0%)
<i>Hibiscus esculentus</i>	18 (45.0%)	22 (55.0%)	16 (40.0%)	24 (60.0%)	17 (42.5%)	23 (57.5%)	51 (42.5%)	69 (57.5%)
<i>Hibiscus sabdariffa</i>	10 (25.0%)	30 (75.0%)	12 (30.0%)	28 (70.0%)	16 (40.0%)	24 (60.0%)	38 (31.7%)	82 (68.3%)
Total	48 (40.0%)	72 (60.0%)	47 (39.2%)	73 (68.8%)	48 (40.0%)	72 (60.0%)	143 (39.7%)	299 (60.3%)

103 Note: Values with asterisk are statistically significant ( $\chi^2_{=0.05}$ )

104 Table 3, showed the abundance of *Melodogyne* spp extracted throughout the study. Out of the  
 105 720 stands of vegetables studied across the three plots, 285 stands were found to be positively infested.  
 106 A total of 535 *Melodogyne* spp of nematode were extracted, out of which *M. incognita* recorded a total of  
 107 190 individuals across the three plots with the highest (78 individuals) in Plot B, while the least in Plot C  
 108 (51). *M. javanica* proved to be more abundant in the study area which had a total of 345 individuals  
 109 across the three Plots, with Plot A having the highest (115 individuals), followed by Plot C (127  
 110 individuals), and the least (103 individuals) was recorded in Plot B (Table 3). The chi-square analysis also  
 111 revealed that there was no significant difference ( $p>0.05$ ) between the total number of nematodes  
 112 extracted across the three plots (Plots A, B and C) but there was a significant difference ( $p>0.05$ )  
 113 between the total number of *M. incognita* and *M. javanica* extracted.

114 **Table 3:** Abundance of *Melodogyne* spp Collected During the Studies (2016)

Nematode spp	Plot A	Plot B	Plot C	Total
<i>Melodogyne incognita</i>	61	78	51	190*
<i>Melodogyne javanica</i>	115	103	127	345*
Total	176	181	178	535

115 Note: Values with asterisk are statistically significant ( $\chi^2_{=0.05}$ )

## 116 DISCUSSION

117 Vascular feeders have become sedentary endoparasites [16]. They usually damage their hosts  
 118 by redirecting large amount of energy and nutrients from normal cellular activities into their special  
 119 feeding sites and developmental activities [16]. They also alter tissues and then disrupt the vascular  
 120 system, which leads to prevention of water transportation and dissolved nutrients upward to the shoots by  
 121 the xylem and also the translocation of photosynthesized materials to other parts of the plants by the  
 122 phloem [17]. Roots severely galled by these nematodes could predispose the plants to root rots which  
 123 could subsequently lead to shorter life span of the plants [18]. Such galled tissues could become  
 124 succulent, poorly protected from invasion by other disease pathogens and nutrient rich food substance  
 125 that can easily be colonized by fungi [18]. Anwar and Mckenry [19] reported that damages by root-knot  
 126 nematodes could lead to poor growth, decline in quality and yield, and reduced resistance to stresses like  
 127 drought and disease.

128 As revealed in this study, it is important to note that, there is an increase in number of infestation  
 129 in each vegetable crop as it ages. This probably signifies that as the plants aged and increases in size,  
 130 more spaces are created for more nematodes to infest and colonizes the vegetable crops. In all stages of  
 131 sampling, *Hibiscus esculentus* proved to be more susceptible to infestation, followed by *Amarantus* spp  
 132 while the least infested was *Hibiscus sabdariffa*. This probably might be due to selection of host by the  
 133 parasites which favored the attack or infestation of *Hibiscus esculentus* than the rest of the vegetables. It

134 could also be that, Sorrel (*H. sabdariffa*) had the least number of infested stands probably because of its  
135 sour nature which hinders the nematodes from infesting it more.

136 In terms of abundance, this study identified two species of root-knot nematodes from the study  
137 area, *Meloidogyne incognita* and *Meloidogyne javanica*. *M. javanica* appeared to be more abundant than  
138 *M. incognita* and this could probably be as a result of the nature of the soil in the study area, which might  
139 be more suitable for the survival of *M. javanica* than *M. incognita*. It could also be that *M. javanica* is more  
140 resistant to stress and other environmental challenges than *M. incognita* which would generally give *M.*  
141 *javanica* reproductive advantage over its cousin (*M. incognita*). This study is in conformity with the works  
142 of Haroon and Zylstru [3], Nagesh *et al.*, [20] and Kayami *et al.*, [9] who all reported that root-knot  
143 nematodes, *M. javanica* is abundantly available than the other species. The result of this study also  
144 agrees with the work of many researchers among which are Khan [21], Verdejo-Lucas *et al.* [22], Anwar  
145 and McKenry [13] and Kayami *et al.* [9] who all reported availability or incidence and abundance of root-  
146 knot nematode in different regions of the world.

## 147 REFERENCE

- 148 1. Platt HM. *The Phylogenetic Systematics of free living nematode*. The Ray Society, London, 1994;  
149 pp.5.
- 150 2. Babatola OO. A Complete Characterization of the Four Most Common Species of Root-Knot  
151 Nematode, *Meloidogyne* with pictorial key. IM Publication Raleigh, N.C.U.S.A, 2003; pp. 20-22.
- 152 3. Haroon SA and Zylstru C. Rapid Identification of Genetic Relationship of *Meloidogyne incognita*  
153 Population by Polymerase Chain Reaction RAP Markers. Egyptian Journal of Agronomatolog,  
154 2003; 32(4): 18-20.
- 155 4. Sasser JN. Economic Importance of *Meloidogyne* in Tropical Countries. In: Lamberti, F. and  
156 Taylor C.E. (Eds). *Root-Knot Nematodes (Meloidogyne spp): Systematics Biology and*  
157 *Control*. Academic Press, New York, 1987; pp 359–374.
- 158 5. Wilson JA. The Incidence of Root-Knot Nematodes *Meloidogyne arenaria*, *Meloidogyne*  
159 *incognita* and *Meloidogyne javanica* on Vegetables. Plant Disease Journal, 2012; 22: 12–18.
- 160 6. Babatola OO and Ogurnunmare OB. Interaction between *Meloidogyne incognita* and *Fusarium*  
161 *oxysporum* on Selected Bean Genotypes. Journal of Nematology, 2012; 45 (2): 20-24.
- 162 7. Ahmed R. Root-Knot of Potato in West Pakistan. Plant Disease Reporter, 2012; 57: 505-510.
- 163 8. Hassan SS and Nagesh M. Incidence of Root-Knot Nematode, *Meloidogyne incognita* and  
164 *Meloidogyne hapla* in Slovenia. Acta Agriculturae Slovenica, 2011; 451 – 474.
- 165 9. Kayami MZ, Mukhtar T and Hussain MA. Association of Root- Knot Nematodes (*Meloidogyne*  
166 spp) With Cucumber in the Pothowar Region of Punjab Province of Pakistan. International  
167 Journal of Biotechnology, 2012; 9 (1-2): 23 – 29.
- 168 10. Begun GP. Geographical Distribution and Infestation of Plant Parasitic Nematodes *Fusarium*  
169 *oxysporum* on selected Bean Genotypes. Journal of Nematology, 2012; 45(1): 231-233.



- 170 11. Wheeler TA, Hake KD and Dever JK. Survey of *Meloidogyne incognita* and *Thielaviopsis basicola*:  
171 Their Impact on Cotton Fruiting and Producers Management Choices in Infested Fields. Journal  
172 of Nematology, 2000; 32(4): 576 – 583.
- 173 12. Barbosa DH, Viera SG, Souza RM and Silva CP. Survey of Root-Knot Nematode *Meloidogyne*  
174 spp. in Coffee plantations in the state Rio de Janeiro, Brazil. Nematologia Brasileira, 2004; 28(1):  
175 43 – 47.
- 176 13. Anwar SA and McKenry MV. Incidence and Population Density of Plant-Parasitic Nematodes  
177 Infecting Vegetable Crops and Associated Yield Loss in Punjab, Pakistan. Pakistan Journal of  
178 Zoology, 2012; 44(2): 327 – 333.
- 179 14. Baermann G. *Baermann method of root-knot nematode extraction. Methods and Techniques for*  
180 *Nematology*, North Carolina State University and state Agency for International Development,  
181 Raleigh, 1948; pp 18-19.
- 182 15. Taylor DP Netscher C. An Improved Technique for Preparing Perineal Patterns of *Meloidogyne*  
183 spp. Nematologica, 1974; 20(2): 107–270.
- 184 16. Anwar SA. Influence of *Meloidogyne incognita*, *Partrichodorus minor* and *Pratylenicus scribneri* in  
185 root-shoot growth and carbohydrate partitioning in tomato. Pakistan Journal of Zoology, 1995; 27:  
186 105 – 113.
- 187 17. Hajera H, Feroza N and Shahina F. Effect of Verm and Nematode Interaction on Some  
188 Biochemical Parameters of Sunflower. *Pakistan Journal of Nematology*, 2009; 27: 193-201.
- 189 18. Abawi GS and Chen J. Concomitant Pathogen and Pest Interactions. In: Plant and Nematode  
190 Interactions (eds. K. R. Barker, G. A. Pederson, and G. I. Windham), Agronomy Monograph 36.  
191 American Society of Agronomy, Madiston, WI,1988; pp 135 – 158.
- 192 19. Anwar SA and McKenry MV. Incidence and reproduction of *Meloidogyne incognita* on vegetable  
193 crop genotypes. Pakistan Journal of Zoology, 2010; 42: 135-141.
- 194 20. Nagesh M, Hussaini SS and Chidanandaswamy BS. Incidence of Root-Knot Nematode,  
195 *Meloidogyne incognita* on Gherkin (*Cucumis sativus*) and Yield Losses. Indian Journal of Plant  
196 Production, 2005; 33:309-311.
- 197 21. Khan RM. Distribution of *Radopholus similis* in India, its Spread in New Regions and Analysis of  
198 the Nematofauna of Banana Crop Pathosystem. Nematologia Mediterranea, 1999; 27(2):  
199 239-246.
- 200 22. Verdejo-Lucas S, Ornat C, Sorribas FJ and Stcheigel A. Species of Root-Knot Nematodes and  
201 Fungal Eggs Parasites Recovered From Vegetables in Almeria and Barcelona, Spain. Journal of  
202 Nematology, 2002; 34(4): 405 – 411.