

Biological characteristics of indigenous *Chrysoperla carnea*

(Neuroptera: Chrysopidae) fed on a natural and an alternative prey.

Abstract

Studying of prey suitability for indigenous insect predators is very important factor for their mass rearing in the future. We tested the effects of two different prey species on the preimaginal stages parameters and adult bionomics of the indigenous predator, *Chrysoperla carnea* under laboratory conditions. These prey species are the aphid; *Aphis fabae* as a natural prey and *Ephestia kuehniella* (Zeller) as a factitious prey. The results showed that prey species had a significant effect on preimaginal development times, survival and fecundity. In contrast, adult longevity, egg hatchability and egg duration were not significantly affected by prey species. Eggs of *E. kuehniella* led to high survival rates, short development times for the preimaginal stages and high fecundity. These results would be helpful for mass rearing of *C. carnea* as an indigenous predator in Saudi Arabia and help in designing integrated pest management programs involving its use as a biocontrol agent of aphids on various crops.

Key words: bionomics, lacewings, *Ephestia* eggs, *Aphis fabae*.

Introduction

The green lacewings, *Chrysoperla carnea* (Stephens, 1836) (Neuroptera: Chrysopidae) is a cosmopolitan polyphagous predator, commonly found in agricultural systems. It is considered as an effective generalist predator of aphids, thrips, coccids, mealy bugs and mites (1, 2, 3, 4). Moreover, *C. carnea* have a compatibility with different environmental conditions, food diversity, its high searching, its ability to predate about 80 species of pests (5, 6, 7). *C. carnea* has been widely used for biological control of aphids and other insect pests because of its polyphagous habits and compatibility with selected chemical insecticides, microbial agents and amenability to mass rearing (8, 9). It has been mass-reared and marketed commercially in the world specially in North America and Europe (5, 6).

C. carnea is not a single species; rather, the “*carnea*-group” is a complex of several cryptic species. Cryptic *Chrysoperla* species can only be identified by analyzing duetting courtship songs from live, sexually receptive males and females. Therefore, references to *C.*

carnea are qualified by sensu lato or sensu strict (10). *C. carnea* was recorded in different regions of Saudi Arabia and it was molecularly identified with different mitochondrial genes (11, 12).

Indigenous species of natural enemies eventually emerged as the key biocontrol agents (13). Consequently, local populations of beneficial arthropods may require a period of adaptation to respond maximally to a newly invasive aphid, although this natural process can be impeded or delayed by inherent characteristics of the cropping system and cultural practices, especially the use of broad-spectrum insecticides (14).

The black bean aphid, *Aphis fabae* (Scopoli, 1763) (Hemiptera: Aphididae), is a polyphagous cosmopolitan pest (15). It is considered as the most important pests of different crops throughout the world such as broad bean and sugar beet in addition to other crops such as cowpea (16). Eggs of the Mediterranean flour moth, *Ephestia kuehniella* (Zeller, 1879) (Lepidoptera: Pyralidae) are considered as one of the most factitious prey for mass production coccinellids species because they ensure rapid growth and development, high survival rates and high fecundity (17). It is very important to identify alternative high quality prey for the successful development of pest management programs that utilize *C. carnea* as a biocontrol agent (18).

The present study aimed to evaluate the developmental and reproductive performance of *C. carnea* when preying on *A. fabae* as a natural prey in comparison to alternative diet, eggs of *E. kuehniella* as a factitious prey under controlled laboratory conditions.

Materials and methods

***C. carnea* samples**

Specimens of the predator, *C. carnea* were obtained from Taif [21° 25' 42" N and 40° 29' 41" E], Saudi Arabia during June 2016 in an area cultivated with clover plants. Each collected male and female pair was maintained in a plastic container (10 cm diameter × 8 cm height) with a hole of 4 cm diameter in the lid and covered by gauze. These adults were fed on an artificial diet (19).

Prey species

E. kuehniella eggs were UV sterilized and maintained from a Chrysopa mass production laboratory (Faculty of Agriculture, Cairo University, Egypt). The bean aphid, *A. fabae* nymphs used in this experiments were collected from broad beans (*Vicia faba* L.) plants and reared in the laboratory on *V. faba*.

Experimental design

The experiments were carried out in the laboratory at 26 ± 2 °C, 16 L: 8 D photoperiods and $65\pm 10\%$ humidity. Each treatment was started with 22 newly hatched larvae as replicates. The newly hatched larvae were transferred individually with a fine hair brush into a plastic petri dishes (5.5 cm diameter and 1.5 cm depth). A piece of filter paper was placed at the bottom of the petri dishes, and a few drops of water were added to maintain humidity (20). Larvae were fed with sufficient numbers of *A. fabae* instars or *E. kuehniella* eggs until the larvae pupated. The experiments were controlled daily to observe the development periods of eggs, first larvae, second larvae, third larvae and pupae. Females and males fed throughout their larval development on the tested prey species were transferred in pairs on the same day of emergence and maintained in plastic containers as described above. The adult stage was observed daily to investigate the total longevity of males and females. The number of eggs laid by each female was recorded daily.

Thirteen randomly chosen eggs laid by each female were collected in plastic Petri dishes under the same controlled conditions to record the hatching rates and egg duration.

Statistics

The t-test was used to compare all characters for male and female *C. carnea* between both prey. The analyses were conducted using SPSS version 23 (21).

Results

Effect of prey species on preimaginal development time and survival.

The effect of feeding on **two different prey** by *C. carnea* on its development time is shown in Table 1. The results indicated that **the duration** of both male and female larval development in *C. carnea* was significantly affected by species of prey tested. The fastest total larval development was obtained on the *E. kuehniella* eggs (13.6 days for females and 13.88 days for males) while these periods on *A. fabae* were 15.13 days for females and 14.75 days for males .

The pupal duration of both males and females were not significantly different when larvae were reared on *E. kuehniella* and *A. fabae*. It was ranged from 10.1 to 10.63 days, **respectively** (Table 1).

The presented results in Table (2) indicated that **the** total duration (from larva to adult) in *C. carnea* females was significantly affected by species of prey tested (25.75 and 23.70

days on *A. fabae* and *E. kuehniella* eggs, respectively) but it was not significantly different for males (25.37 and 24.0 days on *A. fabae* and *E. kuehniella* eggs, respectively).

The higher survival rate of pre-imaginal was recorded when *C. carnea* larvae was feeding on *E. kuehniella* eggs (81.8%) while it was 72.7% with feeding on *A. fabae*.

Effect of prey species on adult longevity and fecundity.

Feeding of different prey to larvae of *C. carnea* (Table 2), significantly affected their fecundity. The mean fecundity per female of *C. carnea* was 373.75 eggs when the larvae was fed on *A. fabae*, whereas it was 481.75 eggs per female when larvae fed on *E. kuehniella* eggs.

There was no significant variation in adult longevity of the same sex due to feeding on different preys although it was higher with feeding on *E. kuehniella* eggs. Female longevity was longer (31.0 and 32.5 days on *A. fabae* and *E. kuehniella* eggs, respectively) than male longevity (19.63 and 21.13 days on *A. fabae* and *E. kuehniella* eggs, respectively) on the same prey (Table 2).

Effect of prey species on egg duration and hatchability.

The incubation periods of eggs of *C. carnea* feeding on different prey were 3.43 and 3.4 days on *A. fabae* and *E. kuehniella* eggs, respectively (Table 3). There was no significant difference between both treatments. Also, no significant difference was recorded for the percentage of eggs hatched with feeding larvae on *A. fabae* (83.12%) and *E. kuehniella* eggs (81.25%) (Table 3).

Discussion

Our experiments indicated that the total larval development of *C. carnea* was shorter on *E. kuehniella* eggs than on *A. fabae*. A previous study stated that the development of *C. carnea* was faster on *E. kuehniella* eggs than on different two aphid species (14). Larva diets of *Chrysoperla rufilabris* (Burmeister, 1839) that contain proteins can promote fast growth and completion of the larval and pupal period (22). Other species of chrysopidae have the same character such as *Dichochrysa tacta* (Navás, 1921) larvae which develops faster on *E. kuehniella* than that of three different aphid species (20). Moreover, various insect predators belong to other orders achieved faster development when *E. kuehniella* eggs as food for their preimaginal stages such as *Hippodamia variegata* (Goeze, 1777) (Coocinellidae: Coleoptera) (23) and two species of genus *Orius* (Heteroptera, Anthocoridae) (24).

In the present study, the highest survival rate was recorded when *C. carnea* larvae was fed on *E. kuehniella* eggs. Other investigation for *C. carnea* recorded that *A. fabae* was the least suitable prey, causing low survival rate and low fecundity (25). Other study showed that the juvenile survival of *C. carnea* was higher on *E. kuehniella* eggs than the other two different aphid species (14).

The current study recorded higher fecundity of *C. carnea* with larvae feeding on *E. kuehniella* eggs than that of *A. fabae*. Similar findings were recorded for *C. carnea* (26) whereas lower fecundity was achieved for the same predator with feeding on *E. kuehniella* eggs (14). Moreover, the number of eggs produced (336.4 per female) by *C. carnea* fed with *Rhopalosiphum maidis* (Fitch, 1856) (27), was near to that observed in the present study on *A. fabae*.

Generally, our experiments have shown that *E. kuehniella* eggs were more suitable than *A. fabae* for the development and reproduction of *C. carnea*, resulting in high preimaginal survival rates, short preimaginal development times and increased fecundity. Similar findings have been reported for *Chrysoperla sinica* (= *C. nipponensis* (Okamoto, 1914)) (26), *Chrysoperla carnea* (28) and *D. tecta* (22). These results would be helpful for optimizing the mass rearing of indigenous *C. carnea* and help in designing the integrated pest management programs involving its use as a biocontrol agent of aphids on various crops.

Table 1: Mean of preimaginal developmental stages time (\pm SE) (days) of *C. carnea* fed on *A. fabae* and *E. kuehniella* eggs at 26 ± 2 °C, 16:8 LD and $65\pm 10\%$ RH.

Prey species	Developmental time in days \pm SE					
	1 st larval instar	2 nd larval instar	3 rd larval instar	Total larva	Pupa	Total (larva-adult)
Females						
<i>A. fabae</i>	3.50 \pm 0.19	5.00 \pm 0.27	6.63 \pm 0.26	15.13 \pm 0.35	10.63 \pm 0.46	25.75 \pm 0.73
<i>E. kuehniella</i>	3.30 \pm 0.15	4.60 \pm 0.22	5.70 \pm 0.21	13.60 \pm 0.31	10.10 \pm 0.38	23.70 \pm 0.45
<i>T</i> values	0.833	1.164	2.762	3.290	0.889	2.506
<i>P</i>	0.417	0.262	0.014	0.005	0.387	0.023
Males						
<i>A. fabae</i>	3.37 \pm 0.18	4.88 \pm 0.30	6.50 \pm 0.19	14.75 \pm 0.31	10.63 \pm 0.42	25.37 \pm 0.50
<i>E. kuehniella</i> eggs	3.25 \pm 0.16	4.75 \pm 0.31	5.88 \pm 0.30	13.88 \pm 0.35	10.13 \pm 0.40	24.00 \pm 0.57
<i>T</i> values	0.509	0.290	1.784	1.861	0.864	1.823
<i>P</i>	0.619	0.776	0.096	0.048	0.402	0.090

Table 2: Adult bionomics of *C. carnea* fed on *A. fabae* and *E. kuehniella* eggs at 26 ± 2 °C, 16:8 LD and $65\pm 10\%$ RH.

Prey species	Male longevity (days \pm SE)	Female longevity (days \pm SE)	Fecundity (eggs/female \pm SE)
<i>A. fabae</i>	19.63 \pm 1.15	31.00 \pm 1.42	373.75 \pm 20.71
<i>E. kuehniella</i> eggs	21.13 \pm 0.90	32.50 \pm 1.60	481.75 \pm 29.13

<i>T</i> values	1.030	0.704	3.021
<i>P</i>	0.320	0.493	0.009

Table 3 : Effect of two different prey on progeny of *C. carnea* at 26±2° C, 16:8 LD and 65±10% RH.

Prey species	Hatchability%	Egg duration
<i>A. fabae</i>	83.12±2.49	3.43±0.09
<i>E. kuehniella</i> eggs	81.25±2.95	3.40±0.09
<i>T</i> values	0.486	0.258
<i>P</i>	0.635	0.798

References

- 1-Hesami S, Farahi S, Gheibi M. Effect of different host plants of normal wheat aphid (*Sitobion avenae*) on the feeding and longevity of green lacewing (*Chrysoperla carnea*), International Conference on Asia Agriculture and Animal IPCBEE. 2011; vol. 13, IACSIT Press, Singapore.
- 2-Singh NN, Manoj K. Potentiality of *Chrysoperla carnea* in suppression of mustard aphid population. Indian Journal of Entomology. 2000; 62: 323-326.
- 3-Zaki FN, Gesraha MA. Production of the green lacewing, *Chrysoperla carnea* (Steph.) (Neuroptera: Chrysopidae) reared on semi-artificial diet based on algae, *Chlorella vulgaris*. Journal of Applied Entomology. 2001; 125: 97-98.
- 4-McEwen PK, New TRR, Whittington A. Lacewings in the crop management. 2001; Cambridge University Press.
- 5-Daane KM, Yokota GY, Zheng Y, Hagen KS. Inundative release of common green lacewings (Neuroptera: Chrysopidae) to suppress *Erythroneura variabilis* and *E.elegantula* (Homoptera; Cicadellidae) in vineyards. Environmental Entomology. 1996; 25: 1224-1234.
- 6-Tauber MJ, Tauber CA, Daane KM, Hagen KS. Commercialization of predators: recent lessons from green lacewings (Neuroptera: Chrysopidae: Chrysoperla). **American Entomologist**. 2000; 46: 26–38.

- 7-Jokar M, Zarabi M. Investigation effect three diets on life table parameters *Chrysoperla carnea* (Steph.) (Neuroptera: Chrysopidae) under Laboratory Conditions. Egyptian Academic Journal of Biological Science. 2012; 5(1): 107-114.
- 8-Uddin J, Holliday NJ, Mackay PA. Rearing lacewings, *Chrysoperla carnea* and *Chrysopa oculata* (Neuroptera: Chrysopidae), on prepupae of alfalfa leafcutting bee, *Megachile rotundata* (Hymenoptera: Megachilidae). *Proceedings of the Entomological Society of Manitoba*. 2005; 61: 11-19.
- 9-Principi MM, Canard M. Feeding habits. In: Canard M, Semeria Y, New TR (Eds.). *Biology of Chrysopidae*. Dr Junk W. The Hague. 1984; pp. 76-92.
- 10-Aldrich JR, Zhang Q-H. Chemical ecology of Neuroptera. *Annual Review of Entomology*. 2016; 61: 197–218.
- 11-Sayed SM, Amer SAM. Molecular variability of *Chrysoperla* Steinman, 1964 (Neuroptera: Chrysopidae) inhabiting western Saudi Arabia. *The Pan-Pacific Entomologist*. 2015; 91(2): 101-107.
- 12-Alghamdi A. Genetic Diversity of *Chrysoperla* sp. at East of Red Sea Using Cytochrome Oxidase Subunit I (COI) Gene. *International Journal of Science and Research*. 2015; 4(3): 1639-1642.
- 13-Nechols JR, Harvey TL. Evaluation of a mechanical exclusion method to assess the impact of Russian wheat aphid natural enemies, pp. 270–279. In Quisenberry SS, Pears FB (eds.), *Response model for an introduced pest–The Russian wheat aphid*. 1998; Thomas Say Publications, Lanham,MD.
- 14-Colares F, Michaud JP, Bain CL, Torres JB. Indigenous aphid predators show high levels of preadaptation to a novel prey, *Melanaphis sacchari* (Hemiptera: Aphididae). *Journal of Economic Entomology*. 2015; 108(6): 2546-2555.
- 15-Blackman RL, Eastop VF. *Aphids on theWorld's Crop, an Identification and Information Guide*. 2000; John Wiley Ltd, London.
- 16-Völkl W, Stechmann DH. Parasitism of the black aphid (*Aphis fabae*) by *Lysiphlebus fabarum* (Hym., Aphidiidae): the influence of host plant and habitat. *Journal of Applied Entomology*. 1998; 122: 201–206.

- 17-Specty O, Febvay G, Grenier S, Delobel B, Piotte C, Pageaux JF, Ferran A, Guillaud J. Nutritional plasticity of the predatory ladybeetle *Harmonia axyridis* (Coleoptera: Coccinellidae): comparison between natural and substitution prey. Archives of Insect Biochemistry and Physiology. 2003; 52: 81-91.
- 18-Takaloozadeh HM. Effect of different prey species on the biological parameters of *Chrysoperla carnea* (Neuroptera: Chrysopidae) in laboratory conditions. Journal Crop Protection. 2015; 4(1):11-18.
- 19-Hassanpour M, Maghami R, Rafiee-Dastjerdi H, Golizadeh A, Yazdanian M, Enkegaard A. Predation activity of *Chrysoperla carnea* (Neuroptera: Chrysopidae) upon *Aphis fabae* (Hemiptera: Aphididae): Effect of different hunger levels. Journal of Asia-Pacific Entomology. 2015; 18: 297–302.
- 20-Sayed SM, Alghamdi A. Suitability of four different prey species for *Dichochrysa tacta* (Neuroptera: Chrysopidae). Biocontrol science and technology. 2017; 27(2). 200-209.
- 21-SPSS. SPSS Base 23.0 for Windows User's Guide. 2015; Chicago, Illinois.
- 22-Cohen AC, Smith LK. A new concept in artificial diets for *Chrysoperla rufilabris*: the efficacy of solid diets. Biological Control. 1998; (13): 49–54.
- 23-Sayed SM, El Arnaouty SA. Effect of corn pollens, as supplemental food, on development and reproduction of the predatory species, *Hippodamia variegata* (Goeze) (Coleoptera: Coccinellidae). Egyptian Journal of Biological Pest Control. 2016; 26 (3): 457-461.
- 24-El-Husseini MM, Agamy EA, Sayed SMH. Comparative biological studies on immatures of *Orius albidipennis* and *Orius laevigatus* (Heteroptera, Anthocoridae) reared on two different preys. Egyptian Journal of Biological Pest Control. 2000; 10 (1&2), 81-88.
- 25-Osman MZ, Selman BJ. Suitability of different aphid species to the predator, *Chrysoperla carnea* (Stephens) (Neuroptera: Chrysopidae). University Journal of Zoology, Rajshahi University. 1993; 12: 101-105.
- 26-El-Arnaouty SA, Ferran A, Beyssat-Arnaouty V. Food consumption by *Chrysoperla carnea* (Stephens) and *Chrysoperla sinica* (Tjeder) of natural and substitute prey: determination of feeding efficiency (Insecta: Neuroptera: Chrysopidae). In: Pure and

Applied Research in Neuropterolog, Proceedings of the 5th International Symposium on Neuropterology, Cairo, May, 1994. (pp. 2-6).

27-El-Serafi HA, Abdel-Salam AH, Abdel-Baky NF. Effect of four aphid species on certain biological characteristics and life table parameters of *Chrysoperla carnea* Stephens and *Chrysopa septempunctata* Wesmael (Neuroptera: Chrysopidae) under laboratory conditions. Pakistan Journal of Biological Sciences. 2000; 3: 239–245.

28-El-Bolok MM, El-Arnaouty SA, Mohammed SM. Predation Capacity of *Chrysoperla carnea* (Stephens) (Neuroptera; Chrysopidae), *Orius albidipennis* (Reuter) (Hemiptera: Anthocoridae) and *Adalia bipunctata* (Linnaeus)(Coleoptera; Coccinellidae) on Two Prey Species. Egyptian Journal of Biological Pest Control. 2010; 20(2): 161-165.