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3 **Biochemical characteristics and Nutritional**
4 **profile of the stem bark extracts from the red**
5 **variety of *Byttneria catalpifolia*, an edible wild**
6 **plant growing in the Western part of Côte**
7 **d'Ivoire.**

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9
10 **ABSTRACT**

11 **Aims:** This study evaluates the biochemical characteristics and nutritional profile of stem bark extracts of the red variety of *Byttneria catalpifolia*, an edible wild plant used as a stem vegetable in the western part of Côte d'Ivoire.

Study design: Dried bark powder and mucilage extracted from the fresh bark of the red variety of *B. catalpifolia* were used to evaluate biochemical composition, minerals and nutritional profile.

Place and Duration of Study: Department of Food Science and Technology (UFR-STA), University Nangui Abrogoua, between January 2015 and December 2017

Methodology: The study was carried out on the mucilage extracted from the fresh bark and the dried bark powder of *B. catalpifolia*. Then, the biochemical composition and the nutritional profile were determined.

Results: The proximate analysis revealed high rates of ashes (11.51 %), crude fibre (50.33 %), reducing sugar (26.37 %), total sugar (44.88 %), caloric energy (135.18 Kcal /100g dw) in Bark powder while the amount of protein (6.01 %) and carbohydrate (30.95 %) were moderate and that of fat was low. Mucilage showed a content of ashes (3.82 %), total sugar (24.84 %), carbohydrate (87.79 %) and caloric energy (336.19 Kcal /100g dw) whereas the rate of crude fibre (0.5 %), reducing sugar (1.80 %) and fat (0.6 %) found to be low and that protein (7.29 %) was moderate. This study indicated that the both samples contained the amino acids and organic acids. The results also showed the both samples appeared to be good sources of minerals such as potassium, calcium and magnesium.

Conclusion: The bark extracts of the red variety of *B. catalpifolia* contain appreciable amounts of nutrients. Their nutritional profile by SAIN and LIM method showed that they belong to the group of foods recommended for health.

12
13 *Keywords: Byttneria catalpifolia, mucilage, nutritional profile, vegetable, stem bark*

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15 **1. INTRODUCTION**

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17 Many plants are consumed as vegetables [1]. The term vegetable corresponds to whole or
18 parts of edible plants consumed, cooked as a component of a dish or raw as a salad.
19 Vegetables include leaves, fruits, seeds, stems, roots, flowers, bulbs, tubers and mushrooms
20 [2] [3]. The vegetables contribute significantly to the nutrition of populations by providing
21 nutrients [4]. Thus, wild edible vegetables are beneficial for marginal populations, especially
22 in developing countries where people have very few resources.

23 Some wild liana plants are edible. Lianas are woody climbing plants that have been
24 extensively studied in the tropics [5] [6]. They constitute an important component of tropical
25 forests. With about 32 % of stems and 35 % of woody species diversity [7], they are
26 generally more abundant and of great diversity in tropical than temperate forests [8] [9].
27 Otherwise, the studies concerning the lianas were carried out on geographical, taxonomical
28 and ecological aspects [10]. Among the liana species, *B. catalpifolia* is a perennial plant that
29 is widely distributed in the tropics of Africa, Asia and America. It belongs to the Family
30 Sterculiaceae and it is the most abundant of the genus *Byttneria*. Based on the colour of the
31 bark, there are two varieties of *B. catalpifolia*. The white variety is characterized by the white
32 colour of the bark when the skin is scraped while the red variety, by a purple colour of the
33 bark. It is an edible wild plant whose bark is used to make a sticky sauce that is well
34 appreciated by the populations of Western Côte d'Ivoire [11] [12]. However, information on
35 the nutritional composition of this plant species is scarce [12]. Note that, assessing the
36 nutritional composition of wild edible plants is important for determining their nutritional
37 significance [13]. The considerable consumption of wild plant species by local people
38 motivated us to carry out the present proximate and nutrients analysis. Despite the use of *B.*
39 *catalpifolia* for several generations as a stem vegetable, to the best of our knowledge, there
40 are no scientific data on the nutritional composition of the the stem bark and the mucilage
41 extracted from the fresh bark of the red variety of *B. catalpifolia*. The mucilage of some
42 plants has been studied by scientists and found to possess biologically active principles.
43 Therefore, the present study was designed to evaluate the biochemical composition and
44 nutritional profile of the dried stem bark powder and the mucilage extracted from fresh bark
45 of the red variety of *B. catalpifolia* and provides consumers with the most appropriate mode
46 of consumption which would give them health benefits.

47 48 **2. MATERIALS AND METHODS**

49 50 **2.1 Sample collection and preparation**

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52 The stem samples of the red variety of *B. catalpifolia* were collected from the western part of
53 Côte d'Ivoire (7°24'45" North latitude and 7°33'13" West longitude). These samples were
54 immediately transported to the laboratory. After removing the epidermis with a kitchen knife,
55 the barks were removed from the stems and chopped. The bark sample was divided into two
56 parts. Part of sample was used for the extraction of mucilage. To do this, 20 g of fresh stem
57 bark were ground in 200 mL of demineralized water using a crusher (Moulinex). The
58 homogenate was centrifuged at 4000 rpm for 10 min. The pellet was ground again in 200 mL
59 and centrifuged under the same conditions as those previously described [14]. The two
60 viscous supernatants were mixed and kept in the freezer for physicochemical analysis. The
61 other part was dried in an oven at 65 °C for 72 h. The dried samples were ground using a
62 crusher (Moulinex) and the resulting sifted (Φ 2 mm) powder was stored in sealed plastic
63 boxes for biochemical analysis.

64 65 **2.2 Proximate analysis**

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67 The AOAC's standard methods [15] were used to obtain the contents in moisture, ash, crude
68 fibre, crude protein, fat and carbohydrate of bark extracts of *B. catalpifolia*. All these
69 analyses were done in triplicates. A difference of weight before and after drying a sample of
70 10 g in an oven (Mettler, Germany) at 105 °C until constant weight allowed to quantify the
71 moisture content. The ash fraction was expressed as the weight of the residue in percentage
72 obtained from a dried sample (5 g) incinerated in a muffle furnace (Pyrolabo, France) at 550
73 °C for 12 h while, the crude fibre content, was obtained from the loss in weight of dried
74 residue after a digestion for fat-free samples with 1.25 % each of sulfuric acid and sodium

75 hydroxide solutions. Through a digestion apparatus applying the macrokjeldahl nitrogen
76 assay method, the crude protein content ($N \times 6.25$) was estimated. A Soxhlet extraction with
77 hexane as a solvent was used for the fat content. Soluble sugars were extracted with 80%
78 neutral aqueous ethanol. Using the ethanolic extract, the reducing sugar content was
79 determined according to the Bernfeld method [16], while the total soluble sugar content was
80 measured through the phenol-sulfuric acid method as described by Dubois et al. [17]. The
81 following formulas allowed to calculate carbohydrate and calorific values [18]:

82 Carbohydrates (%) = $100 - (\% \text{ moisture} + \% \text{ proteins} + \% \text{ lipids} + \% \text{ ash} + \% \text{ fiber})$

83 Calorific value (kcal /100g) = $(\% \text{ proteins} \times 2.44) + (\% \text{ carbohydrates} \times 3.57) + (\% \text{ lipids} \times$
84 $8.37)$

85 The results of ash, crude fibre, crude protein, fat and carbohydrate contents were expressed
86 based on dry weight of samples.

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88 **2.3 Mineral analysis**

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90 According to AOAC principles [19], a scanning electron microscope (SEM) with different
91 pressures (SEM FEG Zeiss Supra 40 VP) allowed to analyse the minerals after wet-ashing.
92 An X-ray detector (Oxford Instruments) related to an energy diffusion spectrometry (EDS)
93 microanalyzer platform (Inca Cool Dry, without liquid nitrogen) is associated to the SEM.
94 Analysis was performed applying evenly to a primed platform with double-sided adhesive
95 carbon, 10 mg of the sample ash residue. The measurement of the chemical elements
96 content was done by measuring the transition energy of the electrons from electronic clouds
97 of the K, L and M series of atoms of the sample.

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99 **2.4 Determination of Amino acids**

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101 The amino acids of the bark extracts (powder and mucilage) of the red variety *B. catalpifolia*
102 were determined by reversed-phase high performance liquid chromatography (PTC column
103 RP-18, 220 mm long, 2.1 mm internal diameter) equipped with a pre-column (SHIMADZU
104 SPD 20A) according to the method described by AOAC [20]. The samples were hydrolysed
105 under vacuum at 150 °C for 60 min in a Pico-Tag station (Waters, Milford, MA, USA) in the
106 presence of 6 % HCl at 1 % phenol. They were then taken up in ultra-pure water and derived
107 automatically thanks to a self-derivator-analyzer-420A (SHIMADZU SPD 20A). The amino
108 acid derivatives obtained in the form of phenyl isothiocyanates (PITC) were separated under
109 elution gradient (7-36 %) using buffer A (45 M sodium acetate at pH 5.9) and buffer B (30 %
110 sodium acetate 105 mM, pH 4.6; 70 % acetonitrile) at a flow rate of 1.5 mL /min. The
111 detection was set at 254 nm and the runtime was 31 min. The acquisition and exploitation of
112 the results was performed using the Model 600 Data Analysis System software (SHIMADZU
113 SPD 20A).

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115 **2.5 Determination of Organic acids**

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117 About 1 g of sample served to extract organic acids using 50 mL of 80 % methanol saturated
118 with NaCl. The organic acids contained in the methanolic extract were determined according
119 to the method of Karadeniz [21] using a HPLC system (Shimadzu Corporation, Japan)
120 equipped with a pump (Shimadzu LC-6A Liquid Chromatograph), a UV detector (Shimadzu

121 SPD-6A UV Spectrophotometric detector) and an integrator (Shimadzu CR 6A
122 Chromatopac). The analysis was performed in isocratic mode using ion exclusion column
123 (ICSep ICE ORH-801, 40 cm x 5 µm, Interchrom, France) that was maintained at 35 °C
124 thanks to a Meta Therm™ furnace (Interchrom, France). Standard solutions were prepared
125 at different concentrations with bidistilled water. Sample (20 µL) was injected and the elution
126 flow rate was maintained at 0.6 mL / min using a mobile phase consisting of sulphuric acid
127 (0.004 N). The detection was set at 210 nm and the runtime was 35 min. The levels of the
128 organic acids in the samples were obtained by comparing the retention times of the eluted
129 compounds with the retention times of the reference solutions. The analysis was carried out
130 in triplicate.

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132 **2.6 Determination of B vitamins**

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134 The determination of B vitamins was performed by the method of Morales et al. [22]. About 5
135 g of sample (mucilage / powder) served to extract B vitamins using 20 mL of methanol (80
136 %). The standards were prepared by dissolving 0.01 g of each standard in methanol (80 %).
137 The methanolic extract was analysed using a HPLC system (SHIMADZU SPD 20A)
138 equipped with a UV detector (PAD) and a C18 ODS column (250 x 4.6 from Cluzeau
139 France) in isocratic mode. A 10 µL of extract or standard was injected. The analysis was
140 carried out at a flow rate of 1.5 mL / min using a mobile phase consisting of a mixture of
141 acetonitrile (55 mL), tetrahydrofuran (37 mL) and water (8 mL) monitored at room
142 temperature. The compounds were detected at a wavelength of 325 nm.

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144 **2.7 Nutritional profile of extracts of the red variety of *B. catalpifolia* according** 145 **to the SAIN and LIM method**

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147 The nutritional profile of dried bark powder and mucilage extracted from the fresh bark of *B.*
148 *catalpifolia* was calculated using the nutrient profiling system proposed by AFSSA and based
149 on two scores, the SAIN score (score of nutritional adequacy of individual foods) and the LIM
150 score (score of nutrients to be limited) [23] [24]. The number of nutrients taken into account
151 to calculate the SAIN score is variable and generally based on the RDA (recommended
152 dietary allowance) of 5, 16 or 23 qualifying nutrients per 100 kcal of food. The LIM score is
153 based on the average percentage of excess of sodium, saturated fatty acids (SFA) and
154 added sugars per 100 g of food [25] [26]. The choice of nutrients to be taken into account is
155 based on the most prevalent nutritional problems of the target population. Thus, the
156 nutritional profile of *B. catalpifolia* extracts was calculated using proteins, crude fibre,
157 calcium, vitamin C and iron. Regarding the LIM score, only sodium was used, because *B.*
158 *catalpifolia* extracts do not contain saturated fatty acids or added sugars. Nutrient contents
159 were expressed on the basis of 100 g of dry matter [24]. The SAIN and LIM results obtained
160 were projected on a four-quadrant graph that positions food according to their composition in
161 qualifying and disqualifying nutrients [23].

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$$SAIN = \frac{\frac{Prot}{65} + \frac{Fib}{30} + \frac{Ca}{900} + \frac{Vit.C}{110} + \frac{Fe}{12.5} \times 100}{Energy} \times 100$$

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$$LIM = \frac{\frac{Na}{3153} + \frac{SFA}{22} + \frac{Added\ sugar}{50}}{3} \times 100$$

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166 Prot : proteins, Fib : fiber, Ca : calcium, Vit. C : vitamin C, Fe : iron, Na : sodium, SFA :
167 saturated fatty acids

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169 2.8 Statistical analysis

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171 All analyses were performed in triplicates. Results were reported as means \pm SD. Means of
172 proximate composition, amino acid and organic acid contents of the dried bark powder and
173 mucilage of *B. catalpifolia* were separated according to the Student's t-test ($P \leq 0.05$) while
174 means of mineral values were analysed according to the Duncan test ($P \leq 0.05$) post
175 ANOVA one way, with the help of JMP® Pro software (version 12, SAS Institute Inc., Cary,
176 NC, 2007).
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178 3. RESULTS

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180 3.1 Proximate composition

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182 Results of the composition of dried stem bark powder and mucilage extracted from the fresh
183 bark of the red variety of *B. catalpifolia* are presented in Table 1. The pH value of dried stem
184 bark powder (5.95 ± 0.02) and mucilage (5.95 ± 0.01) was similar. The dry matter content
185 observed in fresh bark (45.03 ± 0.34 %) was significantly ($p \leq 0.05$) higher than that obtained
186 in the mucilage (6.21 ± 1.15 %). The ash contents obtained in dried stem bark powder and
187 mucilage were 11.51 ± 0.45 % and 3.82 ± 0.62 % of dry weight (dw), respectively. There
188 was meaningful difference ($p \leq 0.05$) between ash contents of dried stem bark powder and
189 mucilage. Our results showed that protein content (7.29 ± 1.01 % dw) and caloric energy
190 (336.19 ± 1.18 kcal /100 g dw) were significantly ($p \leq 0.05$) higher in mucilage than those
191 observed in dried stem bark powder, with values of 6.01 ± 0.20 % dw and 135.18 ± 2.26 kcal
192 /100 g dw, respectively. Crude fibres observed in the dried stem bark (50.33 ± 0.22 % dw)
193 was largely higher than that obtained in the mucilage (0.50 ± 0.01 % dw). Furthermore, this
194 study revealed the presence of parameters such as fat, reducing sugar, total soluble sugar,
195 vitamin B9 and vitamin B2 in the both samples.
196

197 **Table 1: Proximate composition of the dried stem bark and mucilage from the fresh**
198 **bark of the red variety of *B. catalpifolia***

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Parameters	Dried bark powder	Mucilage
pH	5.95 ± 0.02^a	5.95 ± 0.01^a
Dry matter* (%)	45.03 ± 0.34^b	6.21 ± 1.15^a
Ashes (% dw)	11.51 ± 0.45^b	3.82 ± 0.62^a
Fat (% dw)	1.20 ± 0.01^b	0.60 ± 0.01^a
Proteins (% dw)	6.01 ± 0.20^a	7.29 ± 1.01^a
Carbohydrates (% dw)	30.95 ± 0.51^a	87.79 ± 1.01^b
Crude fibres (% dw)	50.33 ± 0.22^b	0.50 ± 0.01^a
Reducing sugar (mg /100 g dw)	26.37 ± 0.05^b	1.80 ± 0.08^a
Total sugar (mg /100 g dw)	44.88 ± 0.16^b	24.84 ± 0.16^a
Vitamin B2 (mg /100 g dw)	694.83 ± 32.23^b	102.18 ± 0.61^a
Vitamin B9 (mg /100 g dw)	31.97 ± 1.37^b	2.09 ± 0.01^a
Caloric Energy (Kcal /100 g dw)	135.18 ± 2.26^a	336.19 ± 1.18^b

200 All analyses were performed in triplicates and the values in the table are the mean \pm standard
201 deviation. On the same line, the means followed by a similar letter are not significantly different ($p \leq$
202 0.05) according to the Student's test; dry weight: dw; *Dry matter of the fresh bark of the red variety of
203 *B. catalpifolia*

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3.2 Amino acid contents

The amino acid compositions of the dried stem bark powder and mucilage from the fresh bark of the red variety of *B. catalpifolia* are illustrated in Table 2. These results indicated that proline, valine, methionine, arginine, glycine, glutamic acid, tyrosine, threonine, lysine and cysteine were found in the both analysed samples. The amino acid contents of dried stem bark powder ranged from 0.11 ± 0.02 mg /100 g dw (threonine) to 3.70 ± 0.01 mg /100 g dw (arginine), while those of mucilage varied from 0.02 ± 0.01 mg /100 g dw (arginine) to 7.57 ± 0.31 mg /100 g dw (cysteine). This study revealed the presence of essential amino acids such as valine, methionine, threonine and lysine in both samples. Their levels ranged from 0.11 ± 0.02 mg /100 g dw (threonine) to 1.64 ± 0.01 mg /100 g dw (methionine) for dried stem bark powder, whereas those of mucilage varied from 1.09 ± 0.01 mg /100 g dw (lysine) to 4.77 ± 0.03 mg /100 g dw (methionine). The statistical analysis showed that the essential amino acids contents of dried stem bark powder were lower meaningfully ($p \leq 0.05$) than those observed in mucilage. Besides, proline, arginine, glutamic acid and glycine (non-essential amino acids) contents in dried bark powder were found also to be lower significantly ($p \leq 0.05$) than those obtained in mucilage, excepted for tyrosine and cysteine contents.

Table 2: Amino acid contents of the dried stem bark powder and mucilage from the fresh bark of the red variety of *B. catalpifolia*

Amino acids (mg /100g dw)	Dried stem bark powder	Mucilage
Proline	0.34 ± 0.11^a	2.33 ± 0.04^b
Valine	0.81 ± 0.28^a	1.11 ± 0.06^a
Methionine	1.64 ± 0.01^a	4.77 ± 0.03^b
Arginine	3.70 ± 0.01^a	7.57 ± 0.31^b
Glycine	0.60 ± 0.01^a	0.81 ± 0.06^b
Glutamic acid	0.43 ± 0.03^a	1.06 ± 0.29^b
Threonine	0.11 ± 0.02^a	2.28 ± 0.07^b
Tyrosine	0.15 ± 0.02^b	0.07 ± 0.00^a
Cysteine	0.43 ± 0.13^b	0.02 ± 0.01^a
Lysine	0.17 ± 0.02^a	1.09 ± 0.01^b

227 *All analyses were performed in triplicates and the values in the table are the mean \pm standard*
228 *deviation. On the same line, the means followed by a similar letter are not significantly different ($p \leq$*
229 *0.05) according to the Student's test; dry weight: dw*

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3.3 Mineral composition

The mineral composition of the dried bark powder and the mucilage extracted from the fresh bark of *B. catalpifolia* are shown in Table 3. The minerals detected were magnesium (Mg), phosphorus (P), potassium (K), calcium (Ca), sodium (Na), iron (Fe), copper (Cu), Zinc (Zn) and iodine (I). The amounts of these minerals in dried stem bark powder appeared significantly higher ($P \leq 0.05$) than those of mucilage. Ca ($414.17 \pm 27.97 - 1104.36 \pm 223.30$ mg /100 g dw) and K ($440.00 \pm 20.93 - 990.69 \pm 227.31$ mg /100 g dw) were the abundant minerals. The least concentrated macro-elements in the mucilage and stem bark powder was Na ($8.02 \pm 1.27 - 28.72 \pm 17.92$ mg / 100 g dw, respectively). The micro-elements observed in the dried stem bark powder were Fe, Cu, Zn and I. Cu and Zn were not detected in mucilage. Iodine (I) content of the mucilage (6.99 ± 1.10 μ g /100 g dw) was meaningfully lower ($p \leq 0.05$) than that obtained in dried stem bark powder (14.61 ± 1.10 μ g /100 g dw).

245 **Table 3: Mineral and mineral ratios of the dried stem bark and mucilage from the bark**
 246 **of the red variety of *B. catalpifolia***
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Minerals (*mg /100 g dw)	Dried bark powder	Mucilage	Pellet
Mg	381.56 ± 162.27 ^c	198.47 ± 16.88 ^b	129.24 ± 18.18 ^a
P	38.50 ± 9.46 ^c	35.09 ± 7.36 ^b	8.01 ± 1.57 ^a
K	990.69 ± 227.31 ^c	440.00 ± 20.93 ^b	368.40 ± 3.78 ^a
Ca	1104.36 ± 223.30 ^c	414.17 ± 27.97 ^a	543.79 ± 85.62 ^b
Na	28.72 ± 17.92 ^c	8.02 ± 1.27 ^a	15.24 ± 0.62 ^b
Fe	11.05 ± 5.07 ^c	5.42 ± 2.92 ^a	6.85 ± 0.1 ^b
Cu	0.49 ± 0.02 ^b	ND	0.21 ± 0.01 ^a
Zn	0.46 ± 0.01 ^b	ND	0.14 ± 0.00 ^a
I (µg /100 g dw)	14.61 ± 1.10 ^b	6.99 ± 1.10 ^a	13.55 ± 0.37 ^b
[Ca]/[P]	28.68	11.80	67.89
[Ca]/[Mg]	2.89	2.09	4.21

248 *All analyses were performed in triplicates and the values in the table are the mean ± standard*
 249 *deviation. On the same line, the means followed by a similar letter are not significantly different (p ≤*
 250 *0.05) according to the test of Duncan; dry weight: dw. Not detected: ND*

251 ** the unit does not apply to ratios.*

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253 **3.4 Organic acid contents**

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255 The organic acids were extracted from the powder of dried stem bark and mucilage of *B.*
 256 *catalpifolia*. They were identified and measured using HPLC system. Tannic, oxalic, citric,
 257 tartaric, sulfanilic, salicylic, adipic, fumaric and benzoic acids were observed. All of these
 258 organic acids were present in both samples. Table 4 clearly indicates that content organic
 259 acids contents of the dried bark powder was found to be lower (p ≤ 0.05) than those
 260 observed in the mucilage except tannic acid and citric acid. In both samples, levels of
 261 fumaric and benzoic acids did not vary significantly (p > 0.05). In addition, organic acids
 262 varied from 0.09 ± 0.02 mg /100 g dw to 51.35 ± 0.75 mg /100 g dw and from 0.11 ± 0.02 mg
 263 /100 g dw to 55.45 ± 0.12 mg /100 g dw for dry bark powder and mucilage, respectively.

265 **Table 4: Organics acid contents of the dried stem bark and mucilage extracted from**
 266 **the fresh bark of red variety of *B. catalpifolia***
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Organic acids (*mg /100 g dw)	Dried bark powder	Mucilage
Tannic acid	6.57 ± 0.03 ^b	4.64 ± 0.31 ^a
Oxalic acid	1.39 ± 0.36 ^a	5.28 ± 0.11 ^b
Citric acid	15.63 ± 0.22 ^b	13.39 ± 0.30 ^a
Tartric acid	51.35 ± 0.75 ^a	55.45 ± 0.12 ^b
Sulfanilic acid	0.15 ± 0.03 ^a	0.31 ± 0.01 ^b
Salicylic acid	0.32 ± 0.03 ^a	2.86 ± 0.23 ^b
Adipic acid	0.31 ± 0.02 ^a	0.50 ± 0.06 ^b
Fumaric acid	0.23 ± 0.07 ^a	0.25 ± 0.02 ^a
Benzoic acid	0.09 ± 0.02 ^a	0.11 ± 0.02 ^a
[oxalic acid] / [Ca]	0.001	0.061

268 *All analyses were performed in triplicates and the values in the table are the mean ± standard*
 269 *deviation. On the same line, the means followed by a similar letter are not significantly different (p ≤*
 270 *0.05) according to the Student's test; dry weight: dw.*

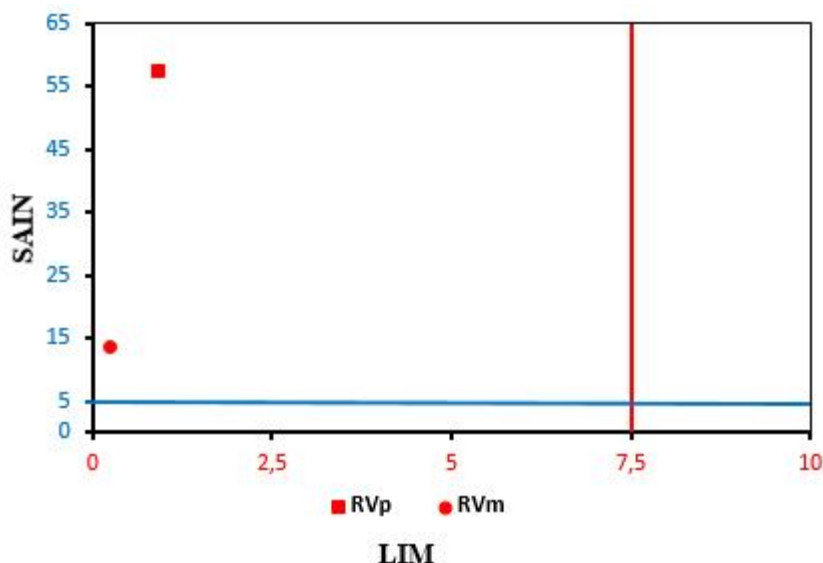
271 ** the unit does not apply to ratios.*

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273 **3.5 Nutritional profile**

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275 The figure 1 shows the SAIN and LIM scores of *B. catalpifolia* extracts (Dried bark and
 276 mucilage). The SAIN score for the dried bark was 57.42 and its LIM score was 0.30. As for
 277 mucilage, the SAIN score was 13.75 and its LIM score recorded was 0.08. The extracts of
 278 this stem vegetable had a high SAIN (SAIN > 5) and a low LIM (LIM < 7.5).
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282 **Fig. 1. SAIN and LIM profile of powder and mucilage extracted from the bark of *B.***
 283 ***catalpifolia***
 284 **(RVp: Red Variety powder; RVm: Red Variety mucilage)**

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286 4. DISCUSSION

287

288 4.1 Proximate composition

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290 This work revealed that ash contents of the dried stem bark and mucilage from the red
 291 variety of *B. catalpifolia* differed meaningfully ($p \leq 0.05$). Note that, ash content is useful in
 292 assessing the quality grading of food materials and also gives an idea of the amount of their
 293 mineral composition [27]. This suggested that a significant part of the minerals was not
 294 extracted with the mucilage. The recorded values of ash were lower than that of the bark
 295 from *Bridelia thermifolia* (12.14 ± 0.63 %) and *Corchorus olitorius* (18.2 %) [28] [29]. By
 296 contrast, they were higher than that found by [30] in tubers of *Tacca leontopetaloides* ($0.80 \pm$
 297 0.01 %).

298 The fat contents of samples testified that the red variety of *B. catalpifolia* is poor in lipids.
 299 Indeed, fat contents of dried stem bark and mucilage of this plant species were low
 300 compared with those of the seeds of *Daniella oliveri* (7.09 %) and *Olax subscorpoidea* (3.66
 301 %) [31]. Our results were also found to be slightly higher than those reported by [32] for the
 302 seed of *Parkia filicoidea* (0.43 %).

303 Dietary fibre is an important component of food in human nutrition because it promotes
 304 intestinal mobility and reduces serum cholesterol, breast cancer and hypertension [33] [34].
 305 In fact, there is evidence that dietary fibre is likely to reduce the absorption rate of glucose
 306 and fat [35], leading to health benefit. Our results revealed that the crude fibre of stem bark
 307 was high while it was low in mucilage. Moreover, the crude fibre of the dried bark powder of
 308 *B. catalpifolia* was higher than that of *Melochia corchorifolia* (23.33 ± 2.89 %), a leafy

309 vegetable [36]. According to [37], the recommend limit for fibre intake is from 1.4 % to 3.5 %.
310 Based on this report, only the dried stem bark powder was good source of dietary fibre.
311 As concern reducing and total soluble sugars, their levels in the stem bark powder were
312 higher than those observed in mucilage, whereas the carbohydrates content of mucilage
313 was higher than that observed in dried bark powder. This showed that the mucilage could
314 contain most complex sugars. The level of carbohydrates of the mucilage from the fresh bark
315 of the red variety of *B. catalpifolia* was high compared with the values of 52.6 % in mucilage
316 of *Ziziphus mauritiana* [38].
317 As for vitamin B2 and B9, the result showed that the both samples were a good source of
318 the vitamin B9. This work revealed that the calculated caloric energy of the both samples
319 was low compared with the respective values of 1476.7 KJ / 100 g dw and 1513.5 KJ /100 g
320 dw in mushroom flours from *Ganoderma spp* and *Hebeloma mesophaeum* [39]. Therefore,
321 the consumption of bark and mucilage of the red variety of *B. catalpifolia* could be
322 recommended to people suffering from obesity.

323

324 4.2 Amino acid content

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326 Amino acids are among the main functionally essential compounds in a food. As a result, the
327 amino acid composition is a reliable indicator of the nutritional value of foods. The amino
328 acid contents found in the mucilage was higher than those of the bark. These low levels of
329 amino acids could be due to the effects of the processing (drying, grinding, sieving)
330 undergone by the bark to obtain the powder. [40] in their work, showed that the quality of a
331 dried product is often lower than the original food with an important impact on nutritional
332 value. Otherwise, amino acids contents of bark powder or mucilage of *B. catalpifolia* were
333 found to be very lower than those of 11 wild edible mushrooms (from 153.09 mg/100 g dw in
334 *F. hepatica* to 2267.32 mg /100 g dw in *B. edulis*) from northeastern Portugal [41]. Besides,
335 the essential amino acids contents of both samples studied were very low compared with the
336 respective values of 154.3 mg /100 g in *B. craspedius* and 5232.5 mg /100 g in *T.*
337 *microcarpus* [42]. This suggested that both samples were poor in essential and non-
338 essential amino acids. Moreover, levels of some of them were very lower than that
339 recommended by [43]. In addition to that, protein contents were relatively low in the mucilage
340 as well as in the dried bark powder and were not sufficient to meet protein requirements or
341 the balance of the amino acids. Thus, we recommend that consumers eat *B. catalpifolia* with
342 meat or fish to balance the diet in protein.

343

344 4.3 Mineral composition

345

346 The mineral composition indicated that the macro-elements such as Mg, P, K, and Ca had
347 relatively high contents in *B. catalpifolia* samples. These results showed a close agreement
348 with those of [44] [45], who reported that Ca and K levels were highest in vegetables. Ca is
349 an essential nutrient needed for many functions in human body. Ca content of the dried bark
350 of *B. catalpifolia* was higher than that of the leaves of *Urtica urens* (830 mg /100 g dw) [46],
351 *Melochia corchorifolia* (750.37 mg /100 g dw) [36] and that of mucilaginous vegetables such
352 as *Irvingia gabonensis* (452 mg /100 g dw) and *Beilschmedia manii* (104 mg /100 g dw) [47].
353 As the recommended daily allowance of Ca is 1200 mg, one serving of the dried bark
354 powder of *B. catalpifolia* per day would help to meet daily Ca requirements [48].
355 The level of K in the dried bark of the red variety of *B. catalpifolia* was higher than that of
356 mucilage and both had high contents in K as well as leaves of *Urera trinervis* (1.25 %) and
357 *Hippocratea myriantha* (1.29 %) [49]. K is involved in the acid-base balance and osmotic
358 regulation of body fluids. It also contributes to nerve and muscle excitability and
359 carbohydrate metabolism [50].

360 Ca and Mg are essential elements of human physiology and are particularly important in the
361 biological functions which are characteristics of the cardiovascular system [51]. The

362 absorption of Mg is closely related to Ca level and P level also influence Ca absorption. Mg
363 levels obtained in the bark and mucilage were greater than those reported by [52] and [53] in
364 *Artocarpus altilis* flours (90.63 - 92.7 mg /100 g) and cassava (36.58 - 37.71 mg /100 g),
365 respectively. Moreover, in the extracts of the bark of *B. catalpifolia*, the [Ca] / [Mg] ratio was
366 greater than 2. Note that, the absorption of Ca and Mg are interdependent, depending on
367 their ratio. Diet is considered good if the ratio between these two minerals in the diet is from
368 2 to 1 in favour of Ca [54].

369 P has more functions than any other mineral element in the body. It forms a complex with Ca
370 that gives stiffness to bones, teeth and muscles [55]. It acts as a cofactor for many enzymes
371 and activates several of the B complex vitamins. It also affects the production of the
372 adenosine triphosphate molecule, which is crucial for energy storage [56]. The P contents of
373 powder and mucilage of *B. catalpifolia* were higher than that of *O. gratissimum*, *T.*
374 *occidentalis*, and *V. amygdalina* (13.8, 13.1 and 15.08 mg /100 g dw, respectively) which are
375 leafy vegetables eaten in South West of Nigeria [57]. It should be noted that, food is also
376 considered "good" if the [Ca] / [P] ratio is greater than 1 and "mediocre" if less than 0.5 [54].
377 The [Ca] / [P] ratio of dried bark powder and mucilage was 28.68 and 11.80, respectively.
378 Thus, the levels of these minerals showed that the dried stem bark and mucilage of this plant
379 may be used to complement the required macro-element needed for proper growth and
380 development in human beings and other domesticated animals.

381 The high content of iodine (I) and other minerals in the pellet compared to the mucilage
382 showed that the use of mucilage to prepare sauce does not allow access to all the minerals
383 contained in this stem vegetable. The relative high concentration of iodine in dried stem bark
384 powder compared with other vegetables such as eggplant leaves (7.6 µg /100 g dw) and
385 melon leaves (6.13 µg /100 g dw) [58] indicated that the bark could be a good source of
386 iodine. Thus, its regular consumption would supply appreciable amount of iodine and would
387 benefit consumers, particularly those in the western of Côte d'Ivoire, where endemic goitre is
388 high (38.3 %) [59]. Indeed, iodine is essential to produce thyroid hormones, triiodothyronine
389 and thyroxine, so that its deficiency can lead to hypothyroidism, goitre, poor physical
390 development and mental disabilities [60].

391

392 4.4 Organic acid contents

393

394 Several organic acids were observed in *B. catalpifolia* extracts with relative important level.
395 Citric and tartaric acids had the highest levels in both samples. Citric acid contents of
396 samples were higher than that observed for *moringa* leaf (1.56 ± 0.45 mg /100 g dw) [61].
397 However, they were lower than that observed in kale (*Brassica oleraceae* L.var. acephala
398 DC.) leaf (2213 mg /100 g dw) [62]. As for tartaric acid contents of both samples, they were
399 found to be lower than that obtained by [61] in Ceylon Spinach (1200 ± 11.56 mg /100 g dw).
400 In contrast, tartaric acid levels were higher than that found in *moringa* leaf by [61], who
401 recorded the value of 0.45 ± 0.02 mg /100 g dw. Note that, the organic acid contents of
402 foods influence their flavour. In addition, they influence the pH of the stomach and the
403 stability and acceptability of foods [63]. The [oxalic acid] / [Ca] ratios of dried bark (0.001)
404 and mucilage (0.061) were less than 2.25, which is the threshold value not to be exceeded.
405 Indeed, oxalic acid interferes with Ca uptake. Thus, the [oxalic acid] / [Ca] ratios below 2.25
406 indicated that the amount of oxalic acid in both samples will not interfere with Ca [64].

407

408 4.5 Nutritional profile

409

410 The nutritional profile of extracts of the red variety of *B. catalpifolia*, following the SAIN and
411 LIM method showed that they belong to the group of foods recommended for health. These
412 results corroborate those of [65] in their work on the assessment of the nutritional profile of

413 foods for children under-five in Abidjan, Côte d'Ivoire. According to these authors, the
414 different vegetable-based soups such as okra, tomato and eggplant have a good nutritional
415 value because their SAIN > 5 and LIM < 7.5. [66] also showed that 80% of vegetables
416 belong to the food group with a strong SAIN and a low LIM and therefore recommended for
417 health. According to [67], non-starchy vegetables such as *B. catalpifolia* are rich in fibres that
418 improve intestinal transit by forming a mass, which leads to a more gradual absorption of
419 nutrients, thus avoiding constipation. They are also very poor in energy and therefore can be
420 consumed in relatively larger amounts to maintain a normal weight [68].

421

422 5. CONCLUSION

423

424 On the basis of the above results it can be concluded that the dried bark and the mucilage
425 extracted from the fresh bark of the red variety of *B. catalpifolia* contain appreciable amounts
426 of nutrients such as carbohydrates, Proteins, essential amino acids, vitamins and minerals.
427 Our findings indicated that the both samples were nutritionally important, since their
428 nutritional profile by SAIN and LIM method showed that they belong to the group of foods
429 recommended for health. The powder of the dried bark was very rich in iodine and other
430 nutrients, therefore the consumption of this powder would be more beneficial to nutritional
431 balance than the mucilage.

432

433

434

435 COMPETING INTERESTS

436

437 The authors declared that they have no conflict of interest.

438

439

440

441 CONSENT

442

443 It is not applicable.

444

445 ETHICAL APPROVAL

446

447 It is not applicable.

448

449 REFERENCES

450

- 451 1. Idris S, Iyaka YA, Ndamitso MM, Paiko YB. Nutritional Composition of the Leaves and
452 Stems of *Ocimum Gratissimum*. J Emerg Trends in Eng Appl Sci. 2011;2(5):801-805
- 453 2. Uzo JO. Tropical vegetable production. In: Food crops productions. Dotan publisher Ltd.
454 Ibadan. 1989;45-49.
- 455 3. Uwaegbule AC. Vegetables: Nutrition and utilization. In: Food crops production. Dotan
456 publishers Ltd. Ibadan. 1989;39-44.
- 457 4. Shad MA, Nawaz H, Rehman T, Ikram N. Determination of some biochemicals,
458 phytochemicals and antioxidant properties of different parts of *Cichorium intybus* L: a
459 comparative study. J Anim Sci. 2013;23:1060-1066.
- 460 5. Phillips OL, Vásquez Martínez R, Arroyo L, Baker TR, Killeen T, Lewis SL et al. Increasing
461 dominance of large lianas in Amazonian forests. Nature. 2002;418(6899):770-4.
- 462 6. Schnitzer SA, Bongers F. Increasing liana abundance and biomass in tropical forests:
463 emerging patterns and putative mechanisms. Ecol Lett. 2011;14:397-406

- 464 7. Schnitzer SA, Mangan SA, Dalling JW, Baldeck C, Hubbell SP, Ledo A et al. Liana
465 abundance, diversity, and distribution on Barro Colorado Island, Panama. PLoS One. 2012;
466 7(12):e52114.
- 467 8. Schnitzer SA, Bongers F. The ecology of lianas and their role in forests. Trends Ecol Evol.
468 2002;17:223-230
- 469 9. Schnitzer SA. A mechanistic explanation for global patterns of liana abundance and
470 distribution. Am Nat. 2005;166:262-276.
- 471 10. Malizia A, Paula I, Campanello, Villagra M, Ceballos S. Geographical, taxonomical and
472 ecological aspects of lianas in subtropical forests of Argentina. N. Parthasarathy (ed.),
473 Biodiversity of Lianas. Sustain Dev Biodivers. 2015;5:17-42
- 474 11. Dan CG, Yao K, Topka GA. Technical sheet of fresh bark of the “white variety” of
475 *Byttneria catalpifolia*, a wild plant consumed in the western part of Côte d’Ivoire. J Pharm Sci
476 Innov. 2017;6(3). DOI: 10.7897/2277-4572.06354
- 477 12. Tokpa GA, Dan CG, Gonnety TJ, Faulet MB, Kouassi KI, Bakayoko A et al. Nutritive
478 Value of Bark Extracts from the “White Variety” of *Byttneria catalpifolia*, a Wild Edible Plant,
479 Consumed as Stem Vegetable in Western of Côte d’Ivoire. Food Public Health.
480 2018;8(3):72-78. DOI: 10.5923/j.fph.20180803.03
- 481 13. Mamta P, Abidi AB, Singh S, Singh RP. Nutritional evaluation of leafy vegetable paratha.
482 J Hum Ecol. 2006;19(2):155-156
- 483 14. Woolfe ML, Chaplin MF, Otchere G. Studies on the mucilages extracted from okra fruits
484 (*Hibiscus esculentus* L.) and baobab leaves (*Adansonia digitata* L.). J Sci Food Agric.
485 1977;28 (6):519 – 529.
- 486 15. AOAC (Association of Official Analytical Chemistry). Official methods of analysis (15th
487 ed.). Washington, DC: Association of AOAC international; Vols. I and II. 16th ed; 1995.
- 488 16. Bernfeld P. Amylase α and β . Methods in enzymology 1.S. P. Colswick and N.O.K., Ed.
489 Academic Press Inc, New-York; 1955.
- 490 17. Dubois M, Gilles KA, Hamilton JK, Roben FA, Smith F. Colorimetric method for
491 determination of sugar and related substances. Analytical Chemistry. 1956;28:350-356.
- 492 18. FAO. Global network on integrated soil management for sustainable use of salt-affected
493 soils. FAO Land and Plant Nutrition Management Services, Rome, Italy; 2002.
- 494 19. AOAC (Association of Official Analytical Chemists). Official Methods of Analysis. AOAC,
495 Washington, 14th Edition. DC; 1990.
- 496 20. AOAC. Official Methods of Analysis, 18th edition Association of Official; 2005.
- 497 21. Karadeniz F. Main organic acid distribution of authentic citrus juices in Turkey. J Agric
498 For. 2004;28:267-271.
- 499 22. Morales P, Fernández-Ruiz V, Sánchez-Mata MC, Cámara M, Tardío J. Optimization
500 and application of FL-HPLC for folates analysis in 20 species of mediterranean Wild
501 vegetables. Food Anal Methods. 2015;8(2):302-311.
- 502 23. AFSSA. Définition de profils nutritionnels pour l’accès aux allégations nutritionnelles et
503 de santé: propositions et arguments [Setting of nutrient profiles for accessing nutrition and
504 health claims: proposals and arguments]. 2008. Accessed 20 June 2018.
505 Available: <http://www.afssa.fr/Documents/NUT-Ra-Profiles.pdf>.
- 506 24. Darmon N, Vieux F, Maillot M, Volatier JL, Martin A. Nutrient profiles discriminate
507 between foods according to their contribution to nutritionally adequate diets: a validation
508 study using linear programming and the SAIN, LIM system. Am J Clin Nutr. 2009;89:1227-
509 36.
- 510 25. Issa C, Salameh P, Batal M, Vieux F, Lairon D, Darmon N. The nutrient profile of
511 traditional Lebanese composite dishes: comparison with composite dishes consumed in
512 France. Int J Food Sci Nutr. 2009;60(S4):285-295.
- 513 26. El Hadrati B. Caracterisation des recettes de certains plats traditionnels a fort potentiel
514 nutritionnel de la region de Sidi Bouzid en Tunisie et evaluation de leur valeur nutritionnelle.
515 Master Nutrition, agro-valorisation, sécurité de l’aliment, Institut de recherche pour le
516 developpement, Université Montpellier 1; 2014.

- 517 27. Smart J. *Canavalia gladiata* (Jacq.) D.C. (Sword bean). Tropical Pulses. Longman Group
518 Ltd, London; 1996.
- 519 28. Adeboye GB, Ameen OM, Abass LT., Physicochemical properties of biodiesel produced
520 from *Jatropha curcas* oil and fossil diesel. J Microbiol Biotechnol Resour. 2004;1:12-16.
- 521 29. Saidou C. Propriétés physico-chimiques et fonctionnelles des gommés hydrocolloïdes
522 des écorces de *Triumfetta cordifolia* et *Bridelia thermifolia*. Université de Grenoble;
523 Université de Ngaoundéré. 2012;235 p.
- 524 30. Ndouyang CJ, Ejoh AR, Aboubakar FB, Njintang YN, Mohammadou BA, Mbofung CM.
525 Valeur nutritionnel de *Tacca leontopetaloides* (L.) Kuntze, tubercule non conventionnel. Rev
526 ind eng. 2009;3:24-32. French
- 527 31. Otori AA, Mann A. Determination of chemical composition, minerals and antinutritional
528 factors of two wild seeds from Nupeland, North Central Nigeria. Am J Chem Appl.
529 2014;1(1):20-26.
- 530 32. Oderinde RA, Ajayi IA, Taiwo VO, Agbedana EO. Dietary effects on growth, plasma lipid
531 and tissues of rats fed with non-conventional oil of *Telfairia occidentalis*. J Sci Food Agric.
532 2004;84:1715-1720.
- 533 33. Anhwange BA, Ajibola VO, Oniye SJ. Chemical studies of *Moringa oleifera* (Lam.) and
534 *Detarium microcapum* (Guill and Sperr). J Biol Sci. 2004;711-715.
- 535 34. Hassan LG, Umar KJ. Proximate and mineral composition of seeds and pulp of *Parkia*
536 *biglobosa* L. Nig J Basic Applied Sci. 2004;13:15-27.
- 537 35. Ekop AS, Ephraim PE, Ekpenyong EO. Chemical composition of African black snails
538 (*Archachatina marginata*) from three different habitats in Akwa, IbornState, Nigeria. Chem
539 Class J. 2004;1:123-126.
- 540 36. Umar KJ, Hassan LG, Dangoggo SM, Inuwa M, Almustapha MN. Nutritional content of
541 *Melochia corchorifolia* (Linn.) Leaves. Int J Biol Chem. 2007;1(4):250-255.
- 542 37. Dietary Guidelines for Americans. U.S. Department of Health and Human Services. U.S.
543 Department of Agriculture; 2005. Accessed 11 February 2018.
544 Available: www.healthierus.gov/dietaryguidelines
- 545 38. Thanatcha R, Pranee A. Extraction and characterization of mucilage in *Ziziphus*
546 *mauritiana* Lam. Int Food Res J. 2011;18:201-212.
- 547 39. Aremu MO, Basu SK, Gyar SD, Goyal A, Bhowmik PK, Datta Banik S. Proximate
548 Composition and Functional Properties of Mushroom Flours from *Ganoderma spp.*,
549 *Omphalotus olearius* (DC.) Sing. and *Hebeloma mesophaeum* (Pers.) Qué. Used in
550 Nasarawa State, Nigeria. Mal J Nutr. 2009;15(2):233-241.
- 551 40. Askari G, Eman-Djomeh Z, Mousari S. An investigation of the effects of drying methods
552 and conditions on drying characteristics and quality attributes of agricultural products during
553 hot air/ microwave-assisted dehydration. Drying Technol. 2009;27:831-841.
- 554 41. Ribeiro B, Andrade PB, Silva BM, Baptista P, Seabra RM, Valentão P. Comparative
555 study on free amino acid composition of wild edible mushroom species. J Agric Food Chem.
556 2008;56:10973–10979
- 557 42. Sun L, Liu Q, Bao C, Fan J. Comparison of Free Total Amino Acid Compositions and
558 Their Functional Classifications in 13 Wild Edible Mushrooms. Molecules. 2017;22(3):1-10
- 559 43. Adeyeye EI. Amino acid composition of three species of Nigerian fish: *Clarias anguillaris*,
560 *Oreochromis niloticus* and *Cynoglossus senegalensis*. Food Chem. 2009;113:43-6.
- 561 44. Oke LO. Chemical studies on commonly used leafy vegetables in Nigeria. J West Afr Sci
562 Ass. 1988;2:47- 49.
- 563 45. Omale J, Ugwu CE. Comparative studies on the protein and mineral composition of
564 some selected Nigerian vegetables. Afr J Food Sci. 2011;5(1):22 – 25.
- 565 46. Turan M, Kordali S, Zengi H, Dursun A, Sezen Y. Macro and micro mineral contents of
566 some edible leaves consumed in Eastern Anatolia. Soil Plant Sci. 2003;53:129-137.
- 567 47. Sahoré A, Kouame M, NemLin J. Physicochemical properties of some traditional
568 vegetables in Ivory Coast: seeds of *Beilschmiedia mannii* (Lauraceae), seeds of *Irvingia*

569 *gabonensis* (Irvingiaceae) and mushroom *Volvariella volvacea*. J Food Technol. 2011;9:57-
570 60.

571 48. National Research Council. Nutrient Requirements of Dairy Cattle, 6th revised ed.,
572 National Academic Press, Washington, D.C. 1989;138-147.

573 49. Andzouana M, Monbouli JB. Proximate, mineral and phytochemical analysis of the
574 leaves *H. myriantha* and *Urera trinervis*. Pak J Biol Sci. 2012;15(11):256-541.

575 50. Aganga AA, Aganga AO, Thema T, Obocheleng KO. Carcass analysis and meat
576 composition of the Donkey. Pak J Nutri. 2003;2(3):138 – 147.

577 51. Huang JH, Tsai LC, Chang YC, Cheng FC. High or low calcium intake increases
578 cardiovascular disease risks in older patients with type 2 diabetes. Cardiovasc Diabetol.
579 2014;13:120.

580 52. Appiah F, Oduro I, Ellis WO. Proximate and Mineral Composition of Artocarpus altilis
581 Pulp Flour as Affected by Fermentation. Pak J Nutr. 2011;10(7):653-657

582 53. Nassar NMA, Alves J, De Souza E. Nutritive value and stature of a cassava (Mandioca),
583 *Manihot esculenta* Crantz Hybrid. Gene Conserve. 2003;2:111-117.

584 54. Adeyeye EI, Aye PA. Chemical composition and the effect of salts on the food properties
585 of *Triticum durum* whole meal flour. Pak J Nutr. 2005;4:187-196.

586 55. Bolanle AO, Funmilola AS, Adedayo A. Proximate Analysis, Mineral Contents, Amino
587 Acid Composition, Anti-Nutrients and Phytochemical Screening of *Brachystegia eurycoma*
588 Harms and *Pipper Guineense* Schum and Thonn. Am J Food Nutr. 2014;2(1):11-17.

589 56. Knochel JP. Phosphorus. In: Modern Nutrition in Health and Disease, 10th edn (eds M.E.
590 Shils, M. Shike, A.C. Ross, B. Caballero and R.J. Cousins) pp. 211-222. Lippincott Williams
591 & Wilkins, Baltimore; 2006.

592 57. Sobowale SS, Olatidoye OP, Olorode OO, Akinlotan, JV. Nutritional potentials and
593 chemical value of some tropical leafy vegetables consumed in south west Nigeria. J Sci
594 Multidiscip Res. 2011;3:1-11

595 58. Taga I, Sameza M, Kayo A, Ngogang J. Évaluation de la teneur en iode des aliments et
596 du sol de certaines régions du Cameroun. Cahiers d'études et de recherches francophones /
597 Santé. 2004;14(1):11-5. French

598 59. Kouamé P, Ouattara H, Bellis G. La carence en iode chez les enfants de glanle, un
599 village de l'Ouest de la Côte d'Ivoire : Une urgence silencieuse. Rev Ivoir Sci Technol.
600 2015;26:245 – 253. French

601 60. Suskind DL. Nutritional deficiencies during normal growth. Pediatr Clin North Am.
602 2009;56(5):1035–1053

603 61. Muangthai P, Nookaew P. Monitoring on Some Organic Acids in Fresh and Processed
604 Rural Plant Leaves in Thailand. Asian J Nat Appl Sci. 2015;4(1):82-89

605 62. Ayaz FA, Glew RH, Millson M, Huang HS, Chuang LT, Sanz C, Hayrliglu-Ayaz S.
606 Nutrient contents of kale (*Brassica oleraceae* L. var. *acephala* DC.). Food Chem. 2006;96:
607 572-579

608 63. Poyrazoglu E, Gokmen V, Artik N. Organic acids and phenolic compounds in
609 pomegranates (*Punica granatum* L.) grown in Turkey, J Food Compos Anal. 2002;15:567-
610 575.

611 64. Tchegang C, Kitkil A. Données ethnonutritionnelles et caractéristiques physicochimiques
612 des légumes feuilles consommés dans la savane de l'Adamaoua (Cameroun). Tropicultura.
613 2004;22(1):11-18. French

614 65. Koné MB, Traore S, Brou K. Use of SAIN and LIM System for Determination of
615 Nutritional Profile of Foods Consumed by Under-five Children in the District of Abidjan, Ivory
616 Coast. Global J Biol, Agric Health Sci. 2016;5(1):1-6

617 66. Darmon N. Les profils nutritionnels: concept, validation, utilisation. Unité de Recherche
618 sur la Prévention Nutritionnelle des Maladies Métaboliques Inra/Faculté de Médecine de la
619 Timone, Marseille; 2010.

620 Available : nicole.darmon@univmed.fr.

- 621 67. Pem D, Jeewon R. Fruit and Vegetable Intake: Benefits and Progress of Nutrition
622 Education Interventions- Narrative Review Article. Iran J Public Health. 2015;44(10):1309-
623 1321
- 624 68. Tohill BC, Seymour J, Serdula M, Kettel-Khan L, Rolls BJ. What Epidemiologic Studies
625 Tell Us about the Relationship between Fruit and Vegetable Consumption and Body Weight.
626 Nutr Rev. 2004;62(10):365-374.