

1 **GC-MS Determination of Bioactive Constituents**  
2 **of the methanolic fractions of**  
3 ***Cnidoscolusaconitifolius***

4  
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12 **ABSTRACT**  
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**Background:**

Diabetes mellitus is a major metabolic disorder affecting a huge population all over the world. *Cnidoscolus* species have been extensively used for the management of diabetes in folkloric medicine. The presence of diverse secondary metabolites has been reported from species of the genus *Cnidoscolus*. However, there has not been much information available on phytochemical components and biological activity in the leaf methanol extract of *Cnidoscolusaconitifolius*

**Objective:**

This study was designed to extract and identify some bioactive compounds in the leaf methanol fractions of *C. aconitifolius* which may provide insight on its pharmacological properties and its use in traditional medicine.

**Place and Duration of Study:**

Department of Biochemistry, Michael Okpara University of Agriculture, Umudike and

National Cereals Research Institute Zaria, Nigeria between June 2012 and July 2013

### **Methodology:**

Twenty grams of the powdered sample were subjected to column chromatography over silica gel (60-120 mesh) and eluted with 100ml each of n-hexane, petroleum ether, chloroform, methanol and respectively at the rate of 1mL/min. The eluates were concentrated and labeled as F<sub>1</sub>, F<sub>2</sub>, F<sub>3</sub> and F<sub>4</sub>. The percentage yields of the fractions were 7.55(%), 6.00(%), 15.5(%), and 65.00%(w/w) respectively. n-hexane and petroleum ether did not elute much of the compounds. The active methanol fraction of *C. aconitifolius* extract (F<sub>4</sub>) which showed the highest hypoglycaemic effect as identified by Oral Glucose Tolerance Test (OGTT) in rats was taken for Gas Chromatography Mass Spectroscopy (GC-MS) analysis for separation of the bio-active components. GC-MS analysis was performed using a GC-MS (Model: QP2010 PLUS SHIMADZU, JAPAN) comprising a AOC-20i auto-sampler and gas-chromatograph interfaced to a mass spectrometer.

### **Results:**

The biologically active organic components of the GC-MS analysis provided peaks of six different phytochemical compounds, with their retention time (RT) and peak area (PA) in addition to minor constituents. The major compounds are dodecanoic acid-1, 2, 3-propanetriyl ester (RT:25.74, PA: 51.18%), cyclotetradecane (RT: 23.39, PA:15.59%), eicosanoic acid (RT:20.61, PA:18.47%); octadecanoic acid (RT: 16.82, PA:1.21%), 4-nitrosophenyl-beta-phenyl propionate (RT: 11.53, PA: 4.38%), benzene acetic acid, phenyl malonic acid and 3-oxo-4-phenylbutyronitrile (RT:10.34, PA: 9.17%). The presence of these compounds in the plant extract may at least be responsible for one of the pharmacological properties of *C. aconitifolius* and thus recommended as plant of phyto-

pharmaceutical importance

14

15 **KEYWORDS:** *Phytochemicals, Bioactive compounds, GC-MS analysis,*  
16 *Cnidoscolumaconitifolius*

## 17 1. INTRODUCTION

18 Use of plants as a source of medicine has been inherited and is an important component  
19 of the health care system. Herbal medicines derived from plant extracts are being utilized  
20 increasingly to treat a wide variety of clinical diseases, although relatively little is known  
21 regarding their modes of action. Studies have shown that commonly consumed medicinal  
22 plants are good sources of polyphenols, saponins, flavonoids and phenyl  
23 propanoids(1). These compounds display a vast variety of pharmacological activities such as anti-  
24 inflammatory, anticancer, anticarcinogenic, antibacterial, antioxidant, antifungal, antiviral activities  
25 etc. In 2002, World Health Organization (WHO) estimated that more than 80% of the  
26 world's population depends on traditional herbal medicine for the treatment of different  
27 ailments (2).

28 Diabetes mellitus is one of the most severe and incurable metabolic disorders  
29 characterized by increased blood glucose level as a result of an absolute or relative lack  
30 of insulin and failure of insulin to act on its target tissue (3). According to the World Health  
31 Organization (WHO), almost 70% of the diabetic patients use plants as a primary source  
32 of antidiabetic agents in order to satisfy their principal health needs (4).

33 . With the increasing demands for herbal medicinal products in healthcare all over the  
34 world, medicinal plant extract manufacturers have started using the most appropriate  
35 extraction technologies in order to identify and isolate the chemical entities present in  
36 them. The purpose of identification of phytochemicals in plants is to attain the  
37 therapeutically desired active portion and to eliminate unwanted materials (5). A special  
38 feature of higher plants is their capacity to produce a large number of organic chemicals of

39 high structural density called secondary metabolites. A knowledge of the chemical  
40 constituents of plants is desirable not only for the discovery of therapeutic agent, but also  
41 for disclosing new sources of economic phyto-compounds for the synthesis of complex  
42 chemical substances and for discovering the actual significance of folklorics.  
43 *C. aconitifolius* (Miller) of family Euphorbiaceae is commonly referred to as '*Chaya*', '*Tree*  
44 *spinach*' in Mexico, '*Efo-lyanaipaja*' or '*Efo-Jerusalem*' in southwest Nigeria and '*Hospital*  
45 *Too Far*' in eastern part of Nigeria. (6). It is an ornamental evergreen drought  
46 deciduous shrub of about 5m tall with 32cm long and 30cm wide palmate leaves  
47 alternately arranged (7). The leaves are commonly eaten as vegetable, serve as blood  
48 builder (8) and possess most essential amino acids thus, making the leavea potential  
49 panacea for kwashiorkor and other related protein-deficiency diseases (9). A wide variety  
50 of the folkloric use of this herb in ethno medicine includes treatment for alcoholism,  
51 diabetes, kidney stone, insomnia, gout, scorpion stings and as cure for brain and vision  
52 improvements (10). Meanwhile it has been utilized extensively as a major component for  
53 the treatment of noninsulin-dependent diabetes mellitus. It has also been reported  
54 recently that the use resulted in a satisfactory hypoglycemic effect in diabetic animal  
55 models(11). Basic research involving animal models have shown that this herb attenuates  
56 renal dysfunction caused by ethanol toxicity, and also exhibits insulinogenic property in  
57 inbred type-2 diabetic mice (12). It also reported to elicit hepatoprotective activity in rats  
58 intoxicated with mega dose of paracetamol (13). The intrinsic potency of medicinal  
59 plants is attributable to the chemical constituents present. Evaluating the biological  
60 potency provides a direct assessment of its pharmacological quality.

61 In order to validate the pharmacological properties of this plant, there is need to identify  
62 the chemical components and bioactive principles present. Therefore the present study  
63 was aimed at identification of the phytochemical constituents present in the methanol  
64 fraction of *Cnidocolusaconitifolius* leaves using GC-MS analysis.

65

66 **2. MATERIALS AND METHODS**

67 **2.1 Collection and identification of Plant Material**

68 Fresh leaves of *Cnidocolusaconitifolius* (CA) were collected with hand in glove from  
69 Amaekpu in Ohafia Local Government Area of Abia State, Nigeria, in the morning hours  
70 between the month of October and December 2012. Samples were identified and  
71 authenticated by Mr. IbeNdukwe, a taxonomist in the Herbarium Section of the  
72 Department of Forestry and Environmental Management, Michael Okpara University of  
73 Agriculture Umudike, Abia State, Nigeria. A voucher specimen was kept at the herbarium  
74 in the same Department (Specimen No. FHI-107727).

75 **2.1.1 Preparation of plant material**

76 The leaves of *C. aconitifolius* were sorted, washed thoroughly with distilled water to remove  
77 dirt and debris, cut into smaller pieces before it was shade dried for 3 weeks at room  
78 temperature ( $28 \pm 3^{\circ}\text{C}$ ). The dried leaves were pulverized into fine powder using electric  
79 blender (CORONA-REF. 121, Landers and Qlink blender, Model No. OBL-15L40). The  
80 powdered materials were stored in air tight polyethene bags protected from direct sunlight  
81 until required for use.

82 **2.1.2 Plant Sample Extraction**

83 One hundred grams of the powdered leaves was extracted with 500mL of 40% ethanol  
84 overnight in a stopped bottle and with occasional stirring at room temperature ( $28 \pm 3^{\circ}\text{C}$ ).  
85 The sample was first sieved using muslin cloth and then filtered using Whatman No.1 filter  
86 paper. This process was repeated three times. The filtrate was concentrated under  
87 reduced pressure at  $40^{\circ}\text{C}$  for 45min in a rotary vacuum evaporator, and then lyophilized  
88 to get a brown aromatic solid extract (ethanol extract of CA). The yield of the extract was  
89 expressed in terms of the percentage of the dry weight of initial plant material used (yield  
90 35.37% w/w). The dry extract obtained was kept in a refrigerator at  $4^{\circ}\text{C}$  until required for  
91 use.

92 **2.1.3 Column Fractionation of Ethanol extract**

93 The dry crude extract was subjected to column chromatography according to standard  
94 methods(14,15). The sample for the column was prepared by adsorbing 20g of the  
95 ethanol extract of *C. aconitifolius* with 60g of silica gel G (60-120 mesh). The mixture was  
96 air dried and carefully layered on top of the packed silica gel in the column (14cm length)  
97 using a glass funnel. The extract in the column was eluted with 100ml each of petroleum  
98 ether, chloroform, methanol and n-hexane respectively at the rate of 1mL/min. The  
99 eluates were concentrated and labeled as F<sub>1</sub>, F<sub>2</sub>, F<sub>3</sub> and F<sub>4</sub>. The percentage yields of the  
100 fractions were 7.55%, 6.00%, 15.5%, and 65.00% (w/w) respectively. Petroleum ether  
101 and hexane did not elute much of the compounds. The methanol fraction of *C.*  
102 *aconitifolius* leaves which showed the highest hypoglycaemic effect as identified by Oral  
103 Glucose Tolerance Test (OGTT) in rats was further analyzed by Gas Chromatography-  
104 Mass Spectrometry (GC-MS).

#### 105 **2.1.4 Gas Chromatography-Mass Spectroscopy (GC-MS)**

106 GC-MS analysis was carried out on a GC-MS (Model: QP2010 PLUS Shimadzu, Japan)  
107 comprising a AOC-20i auto-sampler and gas-chromatograph interfaced to a mass  
108 spectrometer (GC-MS) The instrument is equipped with a VF 5 ms fused silica capillary  
109 column of 30 m length, 0.25 mm diameter and 0.25 µm film thickness. The temperatures  
110 employed were; column oven temperature 80°C, Injection Temp 250°C at a pressure of  
111 108.0kPa, with total flow and column flow of 6.20 ml/min and 1.58ml/min respectively. The  
112 linear velocity was 46.3 cm/sec and a purge flow of 3.0mL/min. The GC program ion  
113 source and interface temperature were 200.00°C and 250.00°C respectively with solvent  
114 cut time of 2.50min. The MS program starting time was 3.00min which ended at 30.00min.  
115 with event time of 0.50sec, scan speed of 1666 µL/sec, scan range 40-800u and an  
116 injection volume of 1 µL of the plant extract (split ratio 10:1). The total running time of GC-  
117 MS was 30 min. The relative percentage of the extract was expressed as percentage with  
118 peak area normalization.

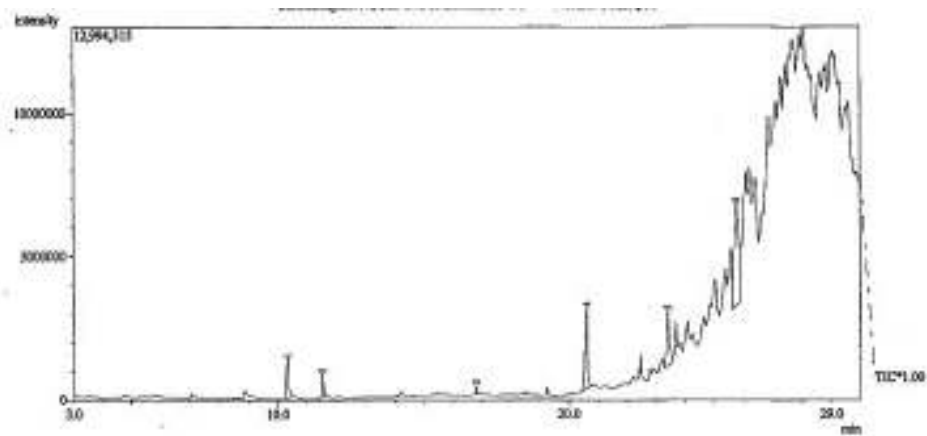
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120 **2.1.5 Identification of phytocompounds**

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122 Interpretation on the mass spectra was conducted using the database of National Institute  
123 Standard and Technology (NIST) having more than 62,000 patterns. The fragmentation  
124 pattern spectra of the unknown components were compared with those of known  
125 components stored in the NIST library (NIST Ver. 2.0 of 2005). The compound bioactivity  
126 prediction is based on Dr. Duke's Phytochemical and Ethnobotanical Databases (Dr.  
127 Duke Database, 2014 <http://www.ars-grin.gov/cgi-bin/duke/ethnobot..>(16).The relative  
128 percentage amount of each phyto-component was calculated by comparing its average  
129 peak area to the total area. The name, molecular weight and structure of the components  
130 of the test materials were ascertained.

131 **3. RESULTS AND DISCUSSION**

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133 Chromatographic purification of the ethanol extract of the leaves of *C. aconitifolius* with  
134 different solvents produced four fractions (F<sub>1</sub>-F<sub>4</sub>) with the methanol fraction producing the  
135 highest yield and hypoglycaemic effect as observed by oral glucose tolerance test in rats.  
136 This may suggest higher proportion of the polar plant component were extracted in the  
137 methanol fraction. The compounds present in the methanol extract of leaves of *C.*  
138 *aconitifolius* were identified by GC-MS analysis as shown in Fig.1.



139

140 **Fig. 1: GC-MS Chromatogram of methanol fraction of *Cnidocolusaconitifolius* leaves**

141 The active principle, area of peak concentration (%), retention time ((RT) molecular weight  
 142 (MW), and molecular formula (MF) in the methanol extract as identified through the NIST  
 143 database are listed in Table 1.

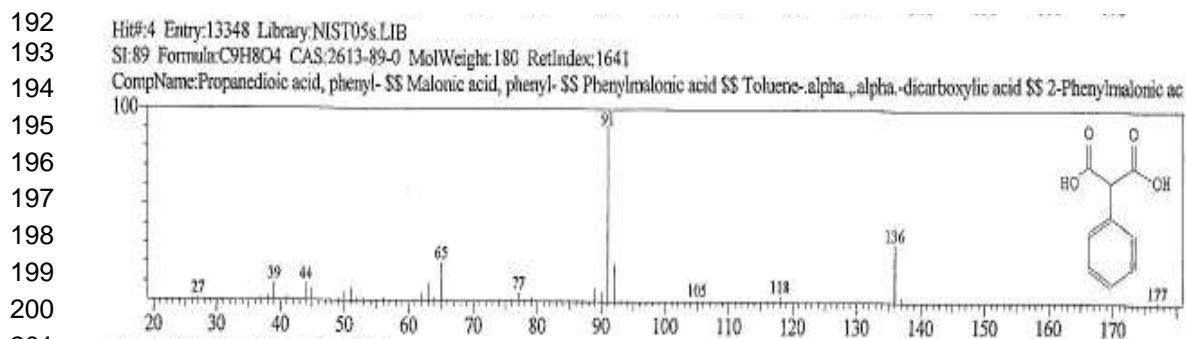
144 Table 1: Phyto-components identified in the methanol extract of *Cnidioscolusaconitifolius* by  
 145 GC-MS analysis

146	Peak	Retention	Name of Compound	Molecular	Molecular	Peak
147	No.	time (s)		Formula	Weight	Area
148	(RT)	(MF)	(MW)	(%)		(g/mol)
151	1	1249	Phenylmalonic acid	C <sub>9</sub> H <sub>8</sub> O <sub>4</sub>	180	9.17
152	1641		Benzene acetic acid	C <sub>8</sub> H <sub>8</sub> O <sub>2</sub>	136	9.17
153	1473		3-Oxo-4-phenylbutyronitrile	C <sub>10</sub> H <sub>9</sub> NO	159	9.17
154	718		Spiro (2, 4) hepta-4, 6-diene	C <sub>7</sub> H <sub>8</sub>	92	9.17
155	2. 000		4 Nitrosophenyl-β-phenyl propionate	C <sub>15</sub> H <sub>13</sub> NO <sub>3</sub>	255	4.38
157	1349		Benzene propanoic acid	C <sub>9</sub> H <sub>10</sub> O <sub>2</sub>	150	4.38
158	1394		3-Phenyl-Propionic acid	C <sub>12</sub> H <sub>16</sub> O <sub>2</sub>	192	4.38
160	31769		Tetradecanoic acid	C <sub>14</sub> H <sub>28</sub> O <sub>2</sub>	228	1.21
161	2167		Octadecanoic acid	C <sub>18</sub> H <sub>36</sub> O <sub>2</sub>	284	1.21
162	1968		n-Hexadecanoic acid	C <sub>16</sub> H <sub>32</sub> O <sub>2</sub>	256	1.21
164	4. 2366		Eicosanoic acid	C <sub>20</sub> H <sub>40</sub> O <sub>2</sub>	312	18.47
165	1869		Pentadecanoic acid	C <sub>15</sub> H <sub>30</sub> O <sub>2</sub>	242	18.47
167	5. 1679		Cyclo-tetradecane	C <sub>14</sub> H <sub>28</sub>	196	15.59
168	18185		Octadecene	C <sub>18</sub> H <sub>36</sub>	252	15.59
169	1620		7-Hexadecene	C <sub>16</sub> H <sub>32</sub>	224	15.59
170	2017		5-Eicosene	C <sub>20</sub> H <sub>40</sub>	280	15.59
172	64336		Do-decanoic acid-1, 2, 3- propane-triyl ester	C <sub>39</sub> H <sub>74</sub> O <sub>6</sub>	638	51.18
174	3218		Do-decanoic acid, 1-hydroxy Methyl-1, 2-diyl ester	C <sub>27</sub> H <sub>52</sub> O <sub>5</sub>	456	51.18
176	1570		Do-decanoic acid, ethenyl ester	C <sub>14</sub> H <sub>26</sub> O <sub>2</sub>	226	51.18

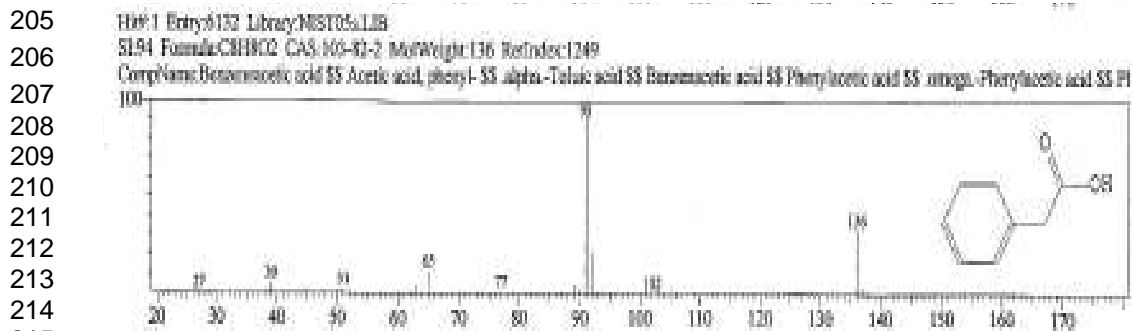
178 The main component of the methanol extract of the leaves was recognized as the  
 179 metabolites responsible for its antidiuretic effect. The organic compounds in methanol  
 180 extract of the leaves which were identified through their fragmentation patterns include 4-  
 181 dodecanoic acid-1, 2 3- propanetriyl ester (RT: 25.744, PA: 51.18%),



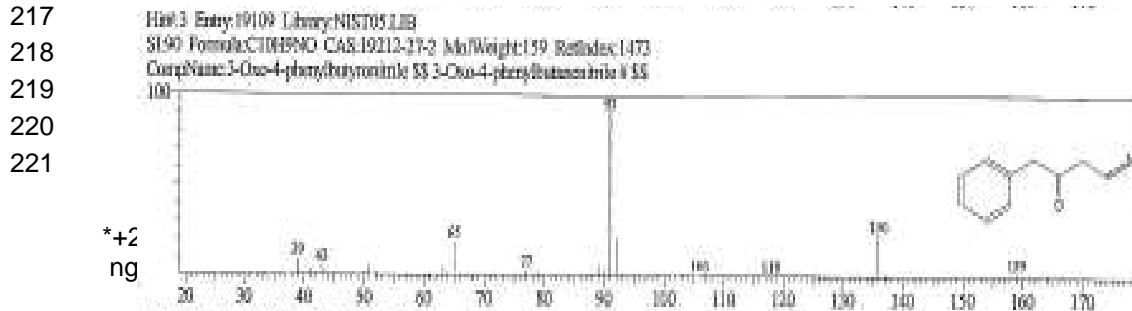
182 cyclotetradecane(RT: 23.39, PA: 15.59%), eicosanoic acid (RT: 20.61, PA: 18.47%) and  
 183 octadecanoic acid ( RT: 16.82, PA: 1.21%). Others are 4-Nitrosophenyl-beta-phenyl  
 184 propionate (RT: 11.53, PA: 4.38%), Benzene acetic acid,phenylmalonic acid and 3-Oxo-4-  
 185 phenylbutyronitrile (RT: 10.34, PA: 9.17). dodecanoic acid-1, 2, 3- propanetriyl ester have  
 186 high retention time (25.74min) and molecular weight (638), while benzene acetic acid is of  
 187 low molecular weight (136) and retention time (10.34 min). A similar trend of the presence  
 188 of fatty acids in extracts of dichloromethane and hexane of *C.aconitifoliushas* been  
 189 reported(17), particularly in studies correlatingthe comparative analysis of the nature of  
 190 the solvent of extraction and chemical composition of the leaves(18)..The mass spectrum  
 191 of the individual components are shown in Figure 2 (a-h)



202 **a.** Phenylmalonic acid



216 **b.**



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226 **c.3-Oxo-4-phenylbutyronitrile**

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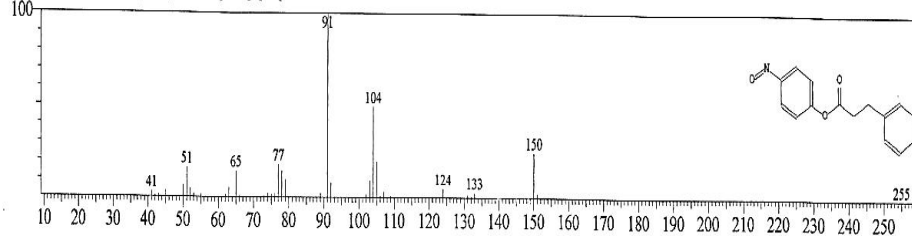
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Hit#1 Entry:74158 Library:NIST05.LIB  
SI:94 Formula:C15H13NO3 CAS:0-00-0 MolWeight:255 RetIndex:0  
CompName:4-Nitrosophenyl-.beta.-phenylpropionate



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237 **d.4 Nitrosophenyl-β-phenyl propionate**

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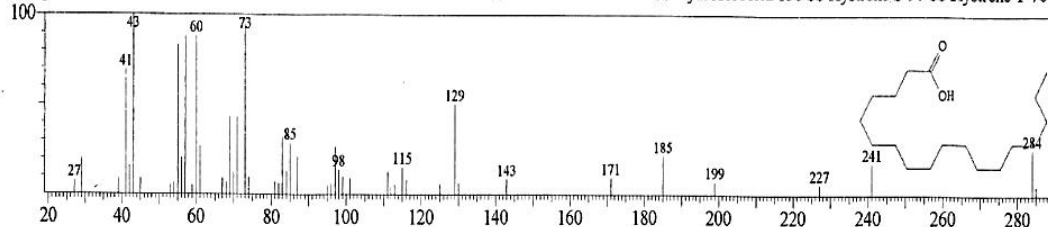
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Hit#4 Entry:22979 Library:NIST05s.LIB  
SI:89 Formula:C18H36O2 CAS:57-11-4 MolWeight:284 RetIndex:2167  
CompName:Octadecanoic acid \$\$ Stearic acid \$\$ n-Octadecanoic acid \$\$ Humko Industrere R \$\$ Hydrofol Acid 150 \$\$ Hystrene S-97 \$\$ Hystrene T-70 \$



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248 **e.Octadecanoic acid**

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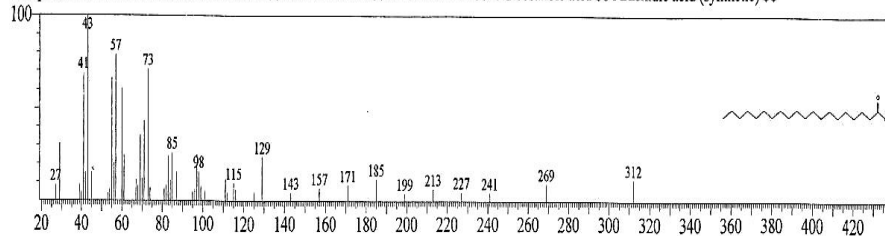
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Hit#4 Entry:108054 Library:NIST05.LIB  
SI:92 Formula:C20H40O2 CAS:506-30-9 MolWeight:312 RetIndex:2366  
CompName:Eicosanoic acid \$\$ Arachic acid \$\$ Arachidic acid \$\$ Icossanoic acid \$\$ n-Eicosanoic acid \$\$ Arachidic acid (synthetic) \$\$



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259 **f.Eicosanoic acid**

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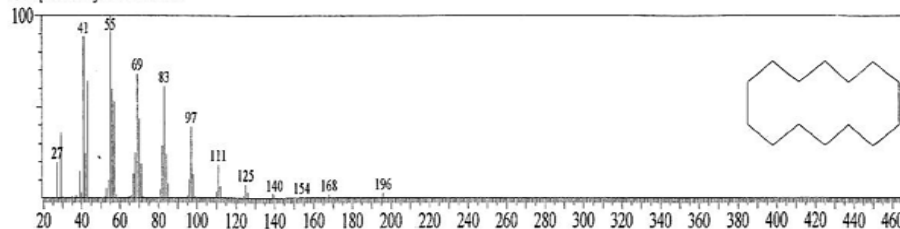
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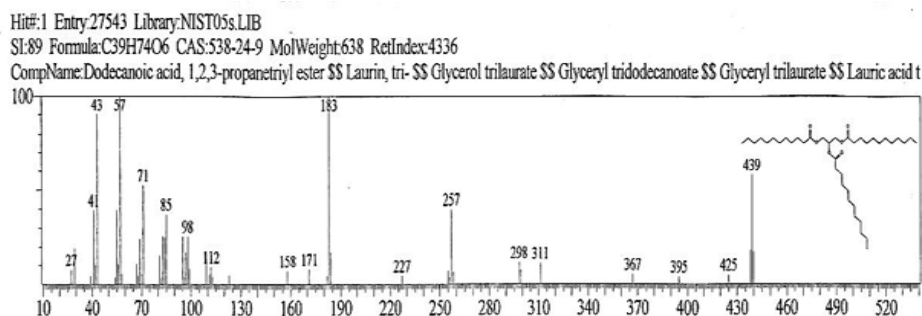
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Hit#4 Entry:15751 Library:NIST05s.LIB  
SI:85 Formula:C14H28 CAS:295-17-0 MolWeight:196 RetIndex:1679  
CompName:Cyclotetradecane



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g. Cyclo-tetradecane



h. Do-decanoic acid-1,2,3-propane-triyl ester

The peaks retention time ranged from 10.33 -25.00 while the peak percentage area ranged from 9.17–15.80. The compounds prediction and biological activities is based on Dr Duke/Phytochemical and ethno botanical databases as tabulated in Table 2.

**Table 2: Biological activities of some active principles present in methanol fraction of *Cnidioscolusaconitifolius* (20)**

Phyto-components	Nature of compound	Activities (16)
Tetradecanoic acid	Fatty acid	Antioxidant, cancer preventive, nematocide, hypercholesterolemic, lubricant
Do-decanoic acid,-1,2,3 –propane -triyl ester	Fatty acid	Hypercholesterolemic, antiarthritic, nematocide, hepatoprotective,
Octadecanoic acid	Fatty acid	b5- $\alpha$ reductase inhibitor, Cosmetic, Flavour, Hypocholesterolemic, Lubricant, Perfumery, Propepic and Suppository.
Eicosanoic acid	Fatty acid	Antiinflammatory, anti-therogenic
9-Octadecenoic acid	Fatty acid	Anti-inflammatory, Anti-alopecic, Anemiagenic, 5- $\alpha$ reductase inhibitor, $\alpha$ -reductase inhibitor lubricant, Antitumour, Choleric,

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			Dermatitigenic, Immunostimulant, Anti-leucotriene-D4, Antiandrogenic, Lipoxygenase inhibitor, Allergenic, Flavour, Hypocholesterolemic, Insectifuge, Irritant, Percutaneo-stimulant, Perfumery and Propecic
	n-Hexadecanoic acid	Fatty acid	Anti-alopecic, Anti-androgenic, Antioxidant, Haemolytic, Hypercholesterolemic, Lubricant, Nematicide, Pesticide, Propecic, Flavour 5- $\alpha$ reductase inhibitor. Anti-inflammatory,
	Dodecanoic acid -ethyl ester	Fatty acid ester	Hypercholesterolemic, Cancer preventive, Hepatoprotective, Nematicide, Insectifuge, Antihistamine, Anti-eczemic, Anti-acne, 5-Alpha reductase inhibitor, Antiandrogenic, Anti- arthritic, Anti-coronary.
	Hexadecanoic acid methyl ester	Fatty acid ester	Antioxidant, Hypercholesterolemic, Lubricant, Nematicide, Pesticide, Hemolytic 5-Alpha reductase inhibitor, Flavour, Antiandrogenic
293	5-Octadecene	Olefins	

294  
295 In the present study, six major compounds have been identified from the ethanol  
296 extract of the leaves of *C. aconitifolius* by Gas Chromatography-Mass Spectrometry  
297 analysis. Authentication of medicinal plants at genetic and chemical level is a critical step  
298 in the use of these botanical materials. Gas Chromatography-Mass Spectrometry (GC-  
299 MS) is a valuable tool for reliable identification of phytochemicals (19).  
300 The GC-MS analysis showed a fragmentation pattern characteristic of the presence of  
301 fatty acids such as octadecanoic acid, tetradecanoic acid, and dodecanoic acid. These  
302 phytochemicals are known to have antimicrobial activity, antioxidant,  
303 hypercholesterolemic, cancer preventive and hepatoprotective activity (20, 21). Similar  
304 compounds were observed in methanol fractions of *C. aconitifolius* with 9-octadecanoic  
305 acid as the prevailing compound (17).  
306 In addition, the methanolic fraction of *C. aconitifolius* also showed the presence of  
307 phenolic compounds, saturated and unsaturated fatty acids including eicosanoic acid, a

308 component of membrane and precursor of a group of hormones like prostaglandins,  
309 thromboxanes and prostacyclins which are important in regulation of diverse  
310 physiological processes(22). The eicosanoids in the extract is reported to have  
311 antiinflammatoryproperties(23).  
312 This action correlates with the active components isolated from this rhizome such as The  
313 study on the active principle of methanol fraction of *C. aconitifolius*revealed that the plant  
314 contains a wide range of phytochemicals which may contribute to its therapeutical value.  
315 The bioactivities of these compounds may depend on the lipophilic properties of their  
316 functional groups. Many metabolites have been found to possess interesting biological  
317 activities and find applications such as pharmaceuticals, insecticides, dyes, flavors and  
318 fragrances (20).

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#### 321 **4. CONCLUSION**

322  
323 From the present study, GC-MS analysis of the methanol fraction of *C. aconitifolius*  
324 *extracts* afforded active compounds which were easily distinguishable fatty acids that  
325 contribute to the therapeutic potential of the plant. This may justify its use as herbal  
326 therapy for the treatment of various diseases by traditional medical practitioners. It also  
327 suggests that further investigation on these phytochemicals will pave the way for the  
328 venture of cost effective drug with less side effect.

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#### 330 **CONSENT**

331 Not applicable.

#### 332 **ETHICAL APPROVAL**

333 Not applicable.

#### 334 **COMPETING INTERESTS**

335 This study will provide additional insights on the use of traditional herbal medicine and to  
336 being able to develop new cure for some ailments like diabetes as one of the leading  
337 diseases causing numerous deaths worldwide.

### 338 **Authors' contributions**

339 This work was carried out in collaboration between both authors. Author NKA participated  
340 in all operations of this manuscript. Author NKA wrote the first draft of the manuscript.  
341 Authors oo designed the study and wrote the protocol performed. Author NKA managed  
342 the analyses of the study and managed the literature searches. Author OO revised the  
343 manuscript and Author NKA has the final responsible for all information presented. Both  
344 authors read and approved the final manuscript

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